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Ganoderma lucidum: a comprehensive review of phytochemistry, efficacy, safety and clinical study



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ABSTRACT

Ganoderma lucidum, one of the most well-known edible fungi, is believed to be very beneficial for longevity and vitality. A long usage history suggests that *G. lucidum* has various clinical therapeutic effects. And experimental studies have confirmed that *G. lucidum* has multiple pharmacological effects, including antitumor, anti-microbial, anti-HIV protease, and antidiabetic activity and so on. With the deepening of research, more than 300 compounds have been isolated from *G. lucidum*. There is an increasing population of *G. lucidum*-based products, and its international development is expanding. Currently, *G. lucidum* has drawn much attention to its chemical composition, therapeutic effect, clinical value, and safety. This paper provides a comprehensive review of these aspects to enhance the global promotion of *G. lucidum*.

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1. Introduction

Ganoderma lucidum belongs to basidiomycete, with a woody texture, widely distributed in tropical and temperate regions in Europe, North America, and Asia^[1]. It is a kind of mushroom for both medicine and food, which has a long history in China, Japan, Korea, and other Asian countries. It has a common custom name in different countries, such as Lingzhi in China, Reishi in Japan, Youngzhi in Korea, and Linh chi in Vietnam. *G. lucidum* has been used medicinally in China for more than 2 000 years. And it was first described its medicinal value in *Shen Nong Ben Cao Jing*, the earliest pharmacopeia in China, written in the Eastern Han dynasty of China (25–220 AD). Chinese Pharmacopeia recorded that it has the effects

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of invigorating qi and tranquilizing spirit (补气安神), relieving cough and asthma (止咳平喘), and can be used for restlessness, insomnia, palpitations, lung deficiency, cough and asthma, consumption and shortness of breath, and loss of appetite^[2]. However, the medicinal value of *G. lucidum* was unknown to Western civilization until the 20^{th} century^[3]. *G. lucidum*, which is considered to be one of the most famous medicinal fungi in the world, is believed to be useful for prolonging longevity and maintaining vitality, and its industry value is estimated to be more than 2.5 billion U.S. dollars^[4].

Modern scientific research and clinical trials have confirmed the ancient knowledge of *G. lucidum* in Asian countries and provided a scientific basis. Modern research shows that *G. lucidum* contains polysaccharides, triterpenoids, steroids, sterols, nucleotides, fatty acids, and other active substances^[5-6]. Among them, polysaccharides and triterpenoids have been thoroughly investigated and widely regarded as the main bioactive components of *G. lucidum*. A large number of studies have confirmed that *G. lucidum* has a wide range of pharmacological effects, such as anti-tumor^[7-8],

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immunomodulation^[9-10], antioxidants^[11-12], antimicrobial^[13-14], anti-diabetes^[15-16], cardioprotective^[11,17], anti-inflammatory^[18-19], anti-androgenic^[20], antimutagenic^[21], neuroprotective^[12], and other pharmacological effects.

Recently, *G. lucidum* has become one of the most studied and product-developed medicinal fungi because of its numerous bioactive substances and extensive pharmacological effects. In the following review, we discuss the recent research status of *G. lucidum* in its chemistry and quality control, bioactivity and its mechanism, preclinical and clinical studies, safety, toxicity, and side effects evaluation.

2. Chemical constituents and quality control methods

According to the literature, there are many varieties of Ganoderma, including G. lucidum (Curtis) P. Karst., Ganoderma amboinase (Lam.) Pat., G. applanatum (Pers.) Pat., G. tsugae Murrill, G. atrum, G. pfeifferi Bres., G. sinense Zhao, Xu et Zhang and so on. Among them, only two species, namely G. lucidum (Curtis) P. Karst. and G. sinense Zhao, Xu et Zhang are recorded in the Pharmacopoeia of the People's Republic of China 2005-2020 edition under the heading of Ganoderma^[22]. G. lucidum as discussed in previous studies, has a wide variety of compounds, including triterpenoids, steroids, polysaccharides, fatty acids, amino acids, nucleosides, proteins, alkaloids and inorganic elements^[22-23]. And triterpenoids and polysaccharides are the main compounds because of their high content, diverse structures, and significant bioactivities^[24]. So far, more than 300 compounds have been isolated from the fruit body, mycelia, and spore of G. lucidum. The nonvolatile component analysis on G. lucidum showed that it is composed of several elements, including ash 0.72%-1.77%, carbohydrate 21.83%-27.78%, fat 1.1%-8.3%, fiber 59%-65%, protein 7%-8%^[25], etc. Different extraction and analytical methods have been selected by the physical and chemical properties of complicated compounds. Herein, various compounds and quality control methods of G. lucidum were summarized respectively in Tables 1-16.

2.1 Chemical constituents

2.1.1 Triterpenoids

Triterpenoids, as a kind of chemical substance prevalent in nature, are also the pharmacologically active components of *G. lucidum*. The fruit body and mycelium of *G. lucidum* contain different types and contents of triterpenoids^[26]. The structure of triterpenoid is derived from lanosterol, whose skeleton is a tetracyclic structure composed of $C_{30}H_{54}$. Most triterpenoids in *G. lucidum* exhibit highly oxidized characteristics^[27]. According to the different functional groups, triterpenoids can be divided into acid, alcohol, ketone, ester, aldehyde, and other types, and their multifarious substituent types in different positions result in a large number of triterpenoids. Over 260 triterpenoids have been identified from the fruit body, spore, and mycelia of *G. lucidum*.

Most triterpenoid skeletons in *G. lucidum* have 27 or 30 carbon atoms, but only a few possess 24 carbon atoms^[28], changing mostly on the substituent of C-17. A typical triterpenoid carbon skeleton marking with 30 carbon atom numbers in *G. lucidum* has shown in

Fig. 1. The skeletal types of Ganoderma triterpenoids in G. lucidum are also present in Fig. 1, which are classified by carbon-carbon double bond position in rings and side chain substituent situations. Moreover, reported compounds with different substituents and formulas are shown in Tables 1-11. Skeleton 1 corresponding to compounds 1-53 has not only a double bond between C-8 and C-9 but also a carbonyl group at C-23. And different substituents appear at C-3, 7, 11, 12, 15, 20, and 27. The substituents at the C-3 position include β -hydroxy, carbonyl group, and β -acetoxy group. There are two main substituents on C-7, C-11, and C-15, namely carbonyl and different configurations of hydroxyl group. In the C-11 position, all of these compounds possess a carbonyl group, except ganoderic acid Df^[29] possesses a β -hydroxy substituent. In C-12, complex substituents appear in this position, such as β -acetoxy, acetoxy, β -hydroxy, α -hydroxy, hydroxy group, and a hydrogen atom. There are two substituents attached to the same carbon in C-20, mainly methyl groups of different configurations and hydrogen atoms or hydroxyl groups. The carboxyl, formyl, acetyl, or butyryl moieties are all common substituents at C-25, which are the most common in the carboxyl group.

Different from the previous compounds in Table 1, compounds 54-92 corresponding to the skeleton 2 have two double bonds between C-8/9 and C-24/25. In addition, the substituents on C-3 and C-26 affect proliferation inhibitory activity, where the carbonyl group at C-3 is essential to inhibit cell proliferation. And the methyl group at C-26 enhances cell membrane permeability^[30]. All of the compounds 93-107 (Skeleton 3) have double bonds between C-8/9 and C-20/22 respectively, while their carbonyl groups at C-11 and C-23. Meanwhile, the skeleton 4 corresponding to compounds 108-115 has a double bond between C-8 and C-9 and a hydroxyl group at C-25, where lucidumol A has good inhibitory activity on α -glucosidase. And the hydroxyl group of C-3 may be an active functional group^[31]. Furthermore, compounds 116-121 corresponding to skeleton 5 share almost the same structure as skeleton 3, except for the double bond between C-16 and C-17 and the new substituent on C-28. The carbon number on the right-side branch of skeletons 6 and 7 is less than that of the previous skeleton, while the right-side branch of skeleton 8 has only an acetoxy group. Of all that skeleton with double bonds between C-8 and C-9, the carboxylic group in the side chain is also essential for recognizing aldose reductase inhibitory activity. Hydroxyl groups at C-3 and C-11, double bond moiety at C-20/C-22 and C-24/C-25 in the side chain are contributed to the promotion of aldose reductase inhibitory activity as well^[32]. Then, unlike the previous skeletons 1-7, skeletons 9 and 10 corresponding to compounds 179-234 have two double bonds between C-7/8 and C-9/11, respectively. The substituent at C-3 is a key group for various pharmacological activities and is essential for cytotoxicity^[33]. The inhibitory activity of sulfatase (STR) is affected by the two positions at C-3 and C-26, with ganodermenonol showing a good inhibitory effect^[34] and the inhibition of 5α reductase activity is probably caused by the C-3-carbonyl and C-26- α , β -unsaturated carbonyl groups, among which ganoderic acid TR exhibited the best inhibitory activity^[35]. In addition, compounds 235-266 are difficult to be classified on the skeleton, so all their structures are drawn in Fig. 2 and their chemical names and formulas are listed in Table 11.

2.1.2 Polysaccharides

Polysaccharides are polymeric carbohydrate molecules composed of long chains of at least ten monosaccharide units bound together by glycosidic linkages. In most of the previous research, polysaccharides have been extracted from the G. lucidum fruit body, spores, mycelia, and cultivation broth^[22]. The structural characterization of polysaccharides isolated from G. lucidum is listed in Table 12. The molecular weights of polysaccharides from different parts of G. lucidum have clear differences. The molecular weight of polysaccharides from the fruit body is between 10³ and 10⁶ Da^[36], while that of polysaccharides from spores and mycelia is about $10^5 - 10^6$ Da. The glucans exhibit complexity in polysaccharides. Glycosidic bonds of the main chain are composed of one single type or a mixture of α -(1 \rightarrow 3) glucan, α -(1 \rightarrow 6) glucan, mannan, and galactosan, with α , β -glucans or other linkages^[37]. Besides that, sugar components of polysaccharides in G. lucidum are glucose, mannose, rhamnose, and galactose.

2.1.3 Steroids

Steroids are also common compounds in G. lucidum, of which



Typical triterpenoid skeleton with carbon number.



Skeletal structure 3 for triterpenoids 93-107.



Skeletal structure 6 for triterpenoids 122-167.



Skeletal structure 9 for triterpenoids 179-226. Fig. 1 \triangle tyr

 R_{4} R_{4} R_{4} R_{6} R_{7} R_{7} R_{7} R_{8} R_{8} R_{8} R_{8} R_{8}

Skeletal structure 1 for triterpenoids 1-53.



Skeletal structure 4 for triterpenoids 108-115.



Skeletal structure 7 for triterpenoids 168-175.





2.1.4 Others

As a member of Chinese medicinal materials, there are many other types of ingredients in *G. lucidum*, such as meroterpenoids, alkaloids and nucleosides, which contain approximately more than 10 compounds in each category. Meroterpenoids are hybrid natural products that partially originate from the terpenoid pathway. Meroterpenoid is composed of a 1,2,4-trisubstituted phenyl group and a polyunsaturated terpenoid part. The diversity of the terpene moiety may be formed through oxidation, cyclization, isomerization, polymerization, etc^[38]. Up to now, no more than 10 alkaloids have been obtained in *G. lucidum*, which include polycyclic alkaloids, purine, pyrimidine, and cerebrosides. The chemical names, structural formulas, references and other information of all other types of ingredients are listed in Table 14.



Skeletal structure 2 for triterpenoids 54-92.



Skeletal structure 5 for triterpenoids 116-121.



Skeletal structure 8 for triterpenoids 176-178.



Table	1
1 4010	

G. lucidum triterpenoids 1–53 with skeletal structure 1.

Triterpenoid	General name	R ₁	R_2	R ₃	R_4	R ₅	R ₆	R ₇	R ₈	Formula	Ref.
1	n-Butyl ganoderate H	β -OH	=0	=O	β-OAc	=0	α -CH ₃	Н	COOBu	$C_{36}H_{52}O_9$	[39]
2	Butyl ganoderate A	=0	β -OH	=0	Н	α-OH	a-CH ₃	Н	COOBu	$C_{34}H_{52}O_7$	[40]
3	Butyl ganoderate B	β -OH	β -OH	=O	Н	=0	a-CH ₃	Н	COOBu	$C_{34}H_{52}O_7$	[40]
4	Ganoderic acid α	β -OH	=O	=O	β -OAc	α-OH	a-CH ₃	Н	COOH	$C_{32}H_{46}O_9$	[41]
5	Ganolucidic acid A	=0	Н	=O	Н	α-OH	α -CH ₃	Н	COOH	C ₃₀ H ₄₄ O ₆	[42]
6	Methyl ganolucidate A	=O	Н	=O	Н	α-OH	a-CH3	Н	COOCH ₃	C31H46O6	[42-43]
7	Ganolucidic acid B	<i>β</i> -OH	Н	=O	Н	α-OH	a-CH ₃	Н	COOH	C ₃₀ H ₄₆ O ₆	[42]
8	Methyl ganolucidate B	<i>β</i> -OH	Н	=O	Н	α-OH	a-CH ₃	Н	COOCH ₃	C ₃₁ H ₄₈ O ₆	[42-43]
9	Ganoderic acid A	=0	<i>β</i> -OH	=O	Н	α-OH	a-CH ₃	Н	СООН	C ₃₀ H ₄₄ O ₇	[44-45]
10	Methyl ganoderate A	=O	, <i>в</i> -он	=0	Н	α-OH	a-CH ₂	Н	COOCH,	C ₂₁ H ₄₄ O ₇	[45]
11	Ganoderic acid B	<i>β</i> -OH	β-OH	=0	Н	=0	a-CH ₂	н	COOH	C ₂₀ H ₄₄ O ₇	[45]
12	Methyl ganoderate B	β-OH	β-OH	=0	н	=0	a-CH.	н	COOCH	C.,H.O.	[44]
13	Ganoderic acid C	β-OH	<i>β</i> -ОН	=0	н	<i>α</i> -OH	a-CH.	н	соон	C ₁₀ H ₄₀ O ₇	[45]
14	Methyl ganoderate C	<i>β</i> ОН	<i>в</i> он	-0	н	a OH	a CH	н	СООСН	$C_{30}H_{42}O_7$	[45]
14	Ganadaria acid D	-0	<i>β</i> -ОН	-0	и и	-0	a CH	н ц		C H O	[45]
15	Methyl ganoderate D	-0	<i>р</i> -ОП <i>в</i> ОН	-0	н	-0	a CH	н	СООСН	$C_{30}H_{42}O_7$	[45]
10	Methyl ganoderate C2	-0 8 OU	<i>р</i> -ОП <i>в</i> ОЦ	-0	п	-0- 01	а-СП ₃	п	COOCH ₃	$C_{31}H_{44}O_7$	[45]
10	Methyl ganoderate C2	<i>р-</i> ОН <i>в</i> ОЦ	р-Оп	=0	п	и-ОП	<i>и</i> -СП ₃	п	COOCH ₃	$C_{31}\Pi_{48}O_7$	[40]
18	Methyl ganoderate K	β-OH	=0	=0	H	α-ΟΗ	a-CH ₃	н	COOCH ₃	$C_{31}H_{44}O_7$	[46-47]
19	/	β-OAc	<i>β</i> -OH	=0	β-OH	=0	α-CH ₃	н	СООН	$C_{31}H_{44}O_9$	[48]
20	Ganoderic acid B9	=0	α-OH	=0	Н	α-ΟΗ	a-CH ₃	н	СООН	C ₃₀ H ₄₄ O ₇	[49]
21	Ethyl ganoderate F	=0	=0	=0	β-OAc	=0	a-CH ₃	Н	COOCH ₃	$C_{34}H_{46}O_9$	[33]
22	/	β -OH	=0	=0	=0	=0	α -CH ₃	Н	COOH	$C_{30}H_{40}O_8$	[50]
23	Ganoderic acid Df	=O	β -OH	β -OH	Н	=0	α -CH ₃	Н	COOH	$C_{30}H_{44}O_7$	[29]
24	Ganoderic acid H	-OH	=O	=O	-OAc	=0	-CH ₃	Н	COOH	$C_{32}H_{44}O_9$	[51]
25	Ganoderic acid F	=O	=O	=O	β -OAc	=0	α -CH ₃	Н	COOH	$C_{32}H_{42}O_9$	[52]
26	Ganoderic acid E	=O	=O	=O	Н	=O	α -CH ₃	Н	COOH	$C_{30}H_{40}O_7$	[52]
27	Ganoderic acid K	-OH	-OH	=O	-OAc	=0	-CH ₃	Н	COOH	$C_{32}H_{46}O_9$	[53]
28	Ganoderic acid AM1	β -OH	=O	=O	Н	=O	α -CH ₃	Н	COOH	$C_{30}H_{42}O_7$	[53]
29	Ganoderic acid J	=O	=0	=O	Н	α-OH	α -CH ₃	Н	COOH	$C_{30}H_{42}O_7$	[53]
30	Ganoderic acid C2	β -OH	β -OH	=0	Н	α-OH	α -CH ₃	Н	COOH	$C_{30}H_{46}O_7$	[46]
31	Ganoderic acid G	-OH	-OH	=0	-OH	=0	-CH ₃	Н	COOH	$C_{30}H_{44}O_8$	[52]
32	/	=O	=0	=0	β -OH	=0	-CH ₃	Н	COOH	$C_{29}H_{38}O_8$	[33]
33	Methyl ganoderate E	=O	=0	=0	Н	=0	α -CH ₃	Н	$COOCH_3$	$\mathrm{C}_{31}\mathrm{H}_{42}\mathrm{O}_{7}$	[54-55]
34	Methyl ganoderate F	=0	=O	=0	β -OAc	=0	α -CH ₃	Н	$\rm COOCH_3$	$C_{33}H_{44}O_9$	[56]
35	Methyl ganoderate H	β -OH	=O	=0	β -OAc	=0	α -CH ₃	Н	$COOCH_3$	$C_{33}H_{46}O_9$	[51,56]
36	Methyl ganoderate G	β -OH	β -OH	=O	β -OH	=0	α -CH ₃	Н	COOCH ₃	$C_{31}H_{46}O_8$	[43]
37	Methyl ganoderate C6	β -OH	=O	=O	β -OH	=0	a-CH ₃	Н	COOCH ₃	C31H44O8	[51]
38	3-O-Acetylganoderic acid B	β-OAc	β -OH	=O	Н	=0	α -CH ₃	Н	COOH	$C_{32}H_{46}O_8$	[57]
39	3-O-Acetylganoderic acid K	β-OAc	=O	=0	Н	α-OH	a-CH3	Н	СООН	C32H46O8	[57]
40	Ethyl 3-O-acetylganoderate B	β-OAc	β -OH	=O	Н	=0	a-CH3	Н	COOCH ₃	C34H50O8	[57]
41	Ethyl ganoderate J	=O	=0	=O	Н	α-OH	a-CH ₃	Н	COOCH ₃	C32H46O7	[57]
42	Ganoderic acid C6	<i>β</i> -OH	=O	=O	<i>β</i> -OH	=0	a-CH ₃	Н	COOH	C ₃₀ H ₄₂ O ₈	[46]
43	/	<i>β</i> -OH	<i>β</i> -OH	=O	β-OAc	=0	a-CH ₃	Н	COOBu	C ₃₆ H ₅₄ O ₀	[58]
44	/	, =0	, =0	=0	β-OAc	=0	a-CH ₂	Н	COOBu	C ₂₆ H ₅₀ O ₀	[58]
45	1	=0	<i>β</i> -OH	=0	ß-OH	=0	a-CH.	н	COOCH.	C.,H.,O.	[51]
46		=0	<i>β</i> -ОН	=0	a-OH	=0	a-CH.	н	соосн.	C.,H.,O.	[18]
47		_0н	р он ß-Он	-0	н	_0H	a-CH.	н	-OH	C. H. O.	[10]
-19	, Methyl ganoderate I	<i>β</i> ОН	<i>в</i> он	-0	н	-0	a CH	R OH	СООН	C H O	[37]
40	Methyl ganoderata N	-0	р-011 8 ОН	-0	и ц	-0	-CH	-0H	COOCH	C. H. O	[+5,40] [60]
49 50	88 0a Dibudroconstante esi 4 I	-0	<i>р</i> -Оп	-0	п	-0 a OU	a CII	л	COOLI	$C_{31}H_{44}O_8$	[00]
50	op, 94-Dinyuloganoderic acid J	=0	=0	=0	п	a-OH	и-СП ₃	п	COOCU	$C_{30}\Pi_{44}O_7$	[01]
50	Concentration of the second se	=0	=0	=0	п	α-0H	α-CH	п	COOLH3	$C_{31}\Pi_{46}O_7$	[01]
52	Ganosporeric acid A	=U	9 CU	=0	=U		α-CH ₃	н	COORT	$C_{30}H_{38}O_8$	[02]
53	Methyl ganoderate G1	p-OH	ρ -OH	=0	ρ -OH	α-OH	α -CH ₃	н	COOCH ₃	$C_{31}H_{48}O_8$	[12]

G. lucidum triterpenoids 54–92 with skeletal structure 2.

Triterpenoid	General name	R_1	R_2	R ₃	R_4	R_5	R_6	R_7	R_8	R_9	Formula	Ref.
54	/	=0	=0	=O	Н	=0	-CH ₃	β -OH	β -OH	COOH	C29H38O7	[53]
55	7-Oxoganoderic acid Z	β -OH	=O	Н	Н	Н	-CH ₃	Н	Н	COOH	$C_{30}H_{46}O_4$	[63]
56	Ganoderic acid LM ₂	=0	β -OH	=O	Н	=0	α -CH ₃	Н	a-OH	COOH	$C_{30}H_{42}O_7$	[64]
57	Lucialdehyde B	=O	=O	Н	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{44}O_{3}$	[49]
58	Lucialdehyde C	β -OH	=O	Н	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{46}O_3$	[49]
59	Ganoderic acid y	=O	β -OH	=O	Н	α-OH	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{44}O_7$	[65]
60	Ganoderic acid δ	=0	α-OH	=O	Н	α-OH	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{44}O_7$	[65]
61	Ganoderic acid ε	β -OH	β -OH	=O	Н	=O	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{44}O_7$	[65]
62	Ganoderic acid ζ	β -OH	=O	=0	Н	=0	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{42}O_7$	[65]
63	Ganoderic acid η	β -OH	β -OH	=0	β -OH	=0	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{44}O_8$	[65]
64	Ganoderic acid θ	β -OH	=O	=O	β -OH	=O	α -CH ₃	Н	β -OH	COOH	$C_{30}H_{42}O_8$	[65]
65	Ganoderic acid β	β -OH	β -OH	=O	Н	=O	α -CH ₃	Н	Н	COOH	$C_{30}H_{44}O_{6}$	[66]
66	Ganolucidic acid E	=O	Н	=0	Н	a-OH	α -CH ₃	Н	Н	COOH	$C_{30}H_{44}O_5$	[67]
67	Ganoderal B	=0	α-OH	Н	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{46}O_3$	[68]
68	Ganoderic acid Ma	α-OAc	a-OAc	Н	Н	α-OH	α -CH ₃	Н	Н	COOH	$\mathrm{C}_{34}\mathrm{H}_{52}\mathrm{O}_{7}$	[69]
69	Ganoderic acid Mi	α-OAc	α -OCH ₃	Н	Н	a-OH	α -CH ₃	Н	Н	COOH	$C_{33}H_{52}O_{6}$	[70]
70	/	=0	=O	α -OH	Н	Н	α -CH ₃	Н	Н	COOH	$C_{30}H_{44}O_5$	[33]
71	/	=0	=O	β -OH	Н	Н	α -CH ₃	Н	Н	COOH	$C_{30}H_{44}O_5$	[33]
72	Lucidadiol	β -OH	=0	Н	Н	Н	α -CH ₃	Н	Н	$CH_{3}OH$	$C_{30}H_{48}O_3$	[71]
73	Lucidal	=0	=O	Н	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{46}O_3$	[71]
74	Ganoderic acid DM	=0	=O	Н	Н	Н	α -CH ₃	Н	Н	COOH	$C_{30}H_{44}O_4$	[72]
75	Lucialdehyde E	=0	β -OH	=0	Н	α-OH	α -CH ₃	Н	Н	CHO	$C_{30}H_{44}O_5$	[73]
76	Ganolucidic acid D	=0	Н	=O	Н	α -OH	α -CH ₃	Н	α -OH	COOH	$C_{30}H_{44}O_{6}$	[65]
77	Ganoderic acid W	α-OAc	α-ОН	=O	Н	α-OAc	α -CH ₃	Н	Н	COOH	$\mathrm{C}_{34}\mathrm{H}_{52}\mathrm{O}_{7}$	[69]
78	Ganoderic acid Mb	-OAc	-OH	Н	Н	-OAc	$-CH_3$	-OAc	Н	COOH	$C_{36}H_{54}O_9$	[69]
79	Ganoderic acid Mc	-OAc	-OAc	Н	Н	-OH	$-CH_3$	-OAc	Н	COOH	$C_{36}H_{54}O_9$	[69]
80	Ganoderic acid Md	-OAc	-OCH ₃	Н	Н	Н	$-CH_3$	-OAc	Н	COOH	$C_{35}H_{54}O_7$	[69]
81	Ganoderic acid Mg	-OAc	-OCH ₃	Н	Н	-OH	$-CH_3$	-OAc	Н	COOH	$C_{35}H_{54}O_8$	[70]
82	Ganoderic acid Mh	-OAc	-OH	Н	Н	-OH	$-CH_3$	-OAc	Н	COOH	$C_{34}H_{52}O_8$	[70]
83	Ganoderic acid Mj	-OH	-OCH ₃	Н	Н	Н	$-CH_3$	-OAc	Н	COOH	$C_{33}H_{52}O_{6}$	[70]
84	Ganoderic acid DH	α-OAc	α-OH	Н	Н	Н	α -CH ₃	β -OAc	Н	COOH	$C_{34}H_{52}O_7$	[74]
85	7-O-Methylganoderic acid O	α-OAc	β -OCH ₃	Н	Н	α-OAc	α -CH ₃	-OAc	Н	COOH	$C_{37}H_{56}O_9$	[75]
86	Ganoderic acid U	α-OH	α-OH	Н	Н	Н	α -CH ₃	Н	Н	COOH	$C_{30}H_{48}O_4$	[76]
87	Ganoderic acid V	=0	a-OH	Н	Н	α-OAc	α -CH ₃	Н	Н	COOH	$C_{32}H_{48}O_6$	[76]
88	Ganoderic acid Z	β -OH	=O	=O	Н	=0	α -CH ₃	Н	α-OH	COOH	$C_{30}H_{48}O_3$	[76]
89	7-O-Ethyl ganoderic acid O	α-OAc	α-OEt	Н	Н	α-OAc	α -CH ₃	OAc	Н	COOH	$C_{38}H_{58}O_9$	[77]
90	Lucialdehyde D	=0	=0	=O	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{42}O_4$	[73]
91	Ganoderic aldehyde A	β -OH	Н	=0	Н	Н	α -CH ₃	Н	Н	CHO	$C_{30}H_{46}O_3$	[78]
92	/	β -OH	α-OH	=O	Н	α-OH	α -CH ₃	Н	Н	-CH ₃	$C_{30}H_{48}O_4$	[59]

Table 3

G. lucidum triterpenoids 93-107 with skeletal structure 3.

	1							
Triterpenoid	General name	R ₁	R_2	R ₃	R_4	R ₅	Formula	Ref.
93	1	β -OH	=0	β-OAc	=O	COOH	$C_{32}H_{42}O_9$	[53]
94	Ganoderenic acid A	=O	β -OH	Н	α-OH	COOH	$C_{30}H_{42}O_7$	[52]
95	Ganoderenic acid B	β -OH	β -OH	Н	=O	COOH	$C_{30}H_{42}O_7$	[52]
96	Ganoderenic acid C	β -OH	β -OH	Н	α-OH	COOH	$C_{30}H_{44}O_7$	[52]
97	Ganoderenic acid D	=O	β -OH	Н	=O	COOH	$C_{30}H_{40}O_7$	[52]
98	1	=O	β -OH	β -OAc	=O	COOH	$C_{32}H_{42}O_9$	[33]
99	Ganoderenic acid H	-OH	=O	Н	=O	COOH	$C_{30}H_{40}O_7$	[53]
100	Ganoderenic acid K	β -OH	β -OH	β -OAc	=O	COOH	$C_{32}H_{44}O_9$	[33]
101	Ganoderenic acid E	=O	β -OH	β -OH	=O	COOH	$C_{30}H_{40}O_8$	[60]
102	Elfvingic acid A	=O	=O	a-OH	β -OH	COOH	$C_{30}H_{40}O_8$	[79]
103	/	=O	=O	β -OH	=O	COOH	C30H38O8	[80]
104	Methyl ganoderenate E	=O	β -OH	β -OH	=O	COOCH ₃	$C_{31}H_{42}O_8$	[60]
105	/	β -OH	=O	β -OAc	=O	COOCH ₃	$C_{33}H_{44}O_9$	[81]
106	/	=O	Н	Н	α-OH	COOCH ₃	$C_{31}H_{44}O_6$	[81]
107	/	=O	β -OH	Н	=O	COOCH ₃	$C_{31}H_{42}O_7$	[81]

G. lucidum triterpenoids 108–115 with skeletal structure 4.

r i i i i i i i i i i i i i i i i i i i								
Triterpenoid	General name	R ₁	R ₂	R ₃	R_4	R ₅	Formula	Ref.
108	Lucidumol A	=O	=0	α -CH ₃	<i>β</i> -OH	-CH ₃	C30H48O4	[66]
109	Ganoderiol C	=O	-OCH ₂ CH ₃	-CH ₃	-OH	-CH ₂ OH	C32H54O5	[67]
110	Ganoderiol D	=O	=O	α -CH ₃	-OH	-CH ₂ OH	C30H48O5	[67]
111	Ganoderiol G	=O	-OCH ₃	-CH ₃	-OH	-CH ₂ OH	C31H52O5	[67]
112	Ganoderiol H	-OH	=O	-CH ₃	-OH	-CH ₂ OH	$C_{30}H_{50}O_5$	[67]
113	Ganoderitriol M	β -OH	=O	α -CH ₃	α-OH	-CH ₃	$C_{30}H_{50}O_4$	[82]
114	/	β -OH	α -OCH ₃	α -CH ₃	α-OH	-CH ₂ OH	C31H54O5	[83]
115	15β -hydroxy-lucidumol A	=O	=O	α -CH ₃	α-OH	-CH ₃	$C_{30}H_{48}O_5$	[83]

Table 5

G. lucidum triterpenoids 116-121 with skeletal structure 5.

	1						
Triterpenoid	Chemical name	R_1	R_2	R ₃	R_4	Formula	Ref.
116	(3β,7β,25R)-3,7-Dihydroxy-11,15,23-trioxolanosta-8,16-dien-26-oic acid	Н	Н	=O	COOH	$C_{30}H_{42}O_7$	[84]
117	3β , 7β -Dihydroxy-11,15,23-trioxolanost-8,16-dien-26-oic acid methyl ester	Н	Н	=O	COOCH ₃	$C_{31}H_{44}O_7$	[84]
118	3β , 7β -Dihydroxy-11,15,23-trioxolanost-8,16-dien-26-oic acid methyl ester	Н	β -OAc	=O	COOH	$C_{32}H_{44}O_9$	[84]
119	3β , 7β , 15β -trihydroxy-11,23-dioxo-lanost-8,16-dien-26-oic acid methyl ester	Н	Н	β -OH	COOCH ₃	$C_{31}H_{46}O_7$	[59]
120	3β , 7β , 15β -trihydroxy-11,23-dioxo-lanost-8,16-dien-26-oic acid	Н	Н	β -OH	COOH	$C_{30}H_{44}O_7$	[59]
121	3β , 7β , 15α ,28-tetrahydroxy-11,23-dioxo-lanost-8,16-dien-26-oic acid	-OH	Н	α-OH	COOH	$C_{30}H_{44}O_8$	[59]

Table 6

G. lucidum triterpenoids 122–167 with skeletal structure 6.

Triterpenoid	General name	R ₁	R_2	R ₃	R_4	R ₅	R ₆	R ₇	R ₈	Formula	Ref.
122	Butyl lucidenate N	<i>β</i> -OH	<i>β</i> -OH	=0	Н	=0	a-CH3	Н	COOBu	C31H48O6	[40]
123	Butyl lucidenate A	=O	β -OH	=O	Н	=O	a-CH ₃	Н	COOBu	$C_{31}H_{46}O_{6}$	[40]
124	Methyl lucidenate D	=O	=O	=0	β -OAc	=0	α -CH ₃	Н	COOCH ₃	$C_{30}H_{40}O_8$	[54,85]
125	20(21)-Dehydrolucidenic acid A	=O	β -OH	=0	Н	=0	$\Delta^{20,21}$	$\Delta^{20,21}$	COOH	C27H36O6	[86]
126	Methyl 20(21)-dehydrolucidenate A	=O	β -OH	=O	Н	=0	$\Delta^{20,21}$	$\Delta^{20,21}$	COOCH ₃	$C_{28}H_{38}O_6$	[86]
127	Lucidenic acid N	β -OH	β -OH	=0	Н	=0	α -CH ₃	Н	COOH	$C_{27}H_{40}O_6$	[87-88]
128	Lucidenic acid D	=O	=0	=0	β -OAc	=0	α -CH ₃	Н	COOH	C29H38O8	[52]
129	Methyl lucidenate E	β -OH	=O	=O	β -OAc	=0	α -CH ₃	Н	COOCH ₃	$C_{30}H_{42}O_8$	[54]
130	Methyl lucidenate F	=O	=0	=O	Н	=O	α -CH ₃	Н	COOCH ₃	$C_{28}H_{38}O_6$	[47,54]
131	Ethyl lucidenates A	=O	β -OH	=O	Н	=O	α -CH ₃	Н	COOCH ₂ CH ₃	$C_{29}H_{40}O_6$	[88]
132	/	β -OCHO	β -OH	=0	-OH	=0	α -CH ₃	Н	COOH	$C_{28}H_{40}O_8$	[48]
133	Lucidenic acid A	=O	β -OH	=0	Н	=0	α -CH ₃	Н	COOH	$C_{27}H_{38}O_6$	[89]
134	Lucidenic acid B	=O	β -OH	=0	β -OH	=0	α -CH ₃	Н	COOH	C27H38O7	[89]
135	Lucidenic acid C	β -OH	β -OH	=0	β -OH	=0	α -CH ₃	Н	COOH	$C_{27}H_{40}O_7$	[89]
136	/	=O	=0	Н	Н	Н	α -CH ₃	Н	COOH	$C_{27}H_{40}O_4$	[33]
137	Lucidenic acid P	β -OH	β -OH	=O	β -OAc	=0	α -CH ₃	Н	COOH	$C_{29}H_{42}O_8$	[90]
138	Methyl lucidenate P	β -OH	β -OH	=O	β -OAc	=0	α -CH ₃	Н	COOCH ₃	$C_{30}H_{44}O_8$	[90]
139	Methyl lucidenate Q	=O	β -OH	=0	Н	α-OH	α -CH ₃	Н	COOCH ₃	$C_{28}H_{42}O_{6}$	[90]
140	/	β -OH	=O	=O	Н	=0	α -CH ₃	Н	COOH	C27H38O6	[50]
141	Methyl lucidenate D2	=O	=O	=O	β -OAc	=0	α -CH ₃	Н	COOCH ₃	$C_{30}H_{40}O_8$	[51]
142	Methyl lucidenate N	β -OH	β -OH	=O	Н	=0	α -CH ₃	Н	COOCH ₃	$C_{28}H_{42}O_{6}$	[91]
143	t-Butyl lucidenate B	=O	β -OH	=O	β -OH	=O	α -CH ₃	Н	COOBu	$C_{31}H_{46}O_7$	[91]
144	Methyl lucidenate A	=O	β -OH	=O	Н	=0	α -CH ₃	Н	COOCH ₃	$C_{28}H_{40}O_6$	[88]
145	20-Hydroxylucidenic acid D2	=O	=O	=0	β -OAc	=0	-OH	-CH ₃	COOH	C29H38O9	[86]
146	20-Hydroxylucidenic acid F	=O	=O	=O	Н	=O	-OH	-CH ₃	COOH	C27H36O7	[86]
147	20-Hydroxylucidenic acid E2	β -OH	=O	=0	β -OAc	=0	-OH	-CH ₃	COOH	$C_{29}H_{40}O_9$	[86]
148	20-Hydroxylucidenic acid N	β -OH	β -OH	=O	Н	=O	-OH	-CH ₃	COOH	C27H40O7	[86]
149	20-Hydroxylucidenic acid P	β -OH	β -OH	=0	β -OAc	=0	-OH	-CH ₃	COOH	$C_{29}H_{42}O_9$	[86]
150	Lucidenic acid F	=O	=O	=O	Н	=0	α -CH ₃	Н	COOH	$C_{27}H_{36}O_{6}$	[46]
151	Methyl lucidenate C	β -OH	β -OH	=0	β -OH	=0	α -CH ₃	Н	COOCH ₃	$C_{28}H_{42}O_7$	[33]
152	Lucidenic acid E2	β -OH	=O	=0	β -OAc	=0	α -CH ₃	Н	COOH	$C_{29}H_{40}O_8$	[90]
153	Lucidenic acid L	β -OH	=O	=0	β -OH	=0	α -CH ₃	Н	COOH	C27H38O7	[60]
154	Lucidenic acid K	=O	=O	=O	α-OH	=O	α -CH ₃	Н	COOH	C27H36O7	[60]
155	Lucidenic acid M	β -OH	α-OH	=0	Н	OH	a-CH3	Н	COOH	C27H42O6	[60]
156	Methyl lucidenate K	=O	=O	=0	α-OH	=0	α -CH ₃	Н	COOCH ₃	C28H38O7	[18]
157	Methyl lucidenate L	β -OH	=O	=0	β -OH	=O	a-CH3	Н	COOCH ₃	$C_{28}H_{40}O_7$	[18]
158	Methyl lucidenate M	β -OH	α -OH	=0	Н	OH	α -CH ₃	Н	COOCH ₃	$C_{28}H_{40}O_{6}$	[60]
159	Ethyl lucidenate A	=O	β -OH	=0	Н	=O	a-CH3	Н	COOCH ₂ CH ₃	$C_{29}H_{42}O_{6}$	[88]
160	Butyl lucidenate P	<i>β</i> -OH	β -OH	=0	β-OAc	=O	α -CH ₃	Н	COOBu	C33H50O8	[92]
161	Butyl lucidenate Q	=O	<i>β</i> -OH	=O	Н	α-OH	a-CH ₃	Н	COOBu	C31H48O6	[92]
162	Butyl lucidenate D2	=O	=O	=0	β-OAc	=O	α -CH ₃	Н	COOBu	C33H46O8	[92]
163	Butyl lucidenate E2	β -OH	=O	=O	β-OAc	=O	a-CH ₃	Н	COOBu	C33H48O8	[92]
164	- /	β-ΟСНО	<i>β</i> -OH	=O	OH	=O	a-CH ₃	Н	COOH	C28H40O8	[48]
165	Lucidenic acid D1	=0	=0	=O	=0	=0	a-CH ₃	Н	COOH	C27H34O7	[93]
166	Lucidenic acid E1	=O	<i>β</i> -OH	=O	α-OH	=O	a-CH ₃	Н	COOH	C27H38O7	[93]
167	/	=O	=0	Н	Н	Н	a-CH ₃	Н	COOH	C27H40O4	[33]

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Table 7

G. lucidum triterpenoids 168–175 with skeletal structure 7.

Triterpenoid	General name	R ₁	R ₂	R ₃	R_4	R ₅	Formula	Ref.
168	Methyl lucidenate H	<i>β</i> -OH	β -OH	Н	=O	COOCH ₃	$C_{28}H_{42}O_7$	[60]
169	Methyl lucidenate I	β -OH	=O	Н	=O	COOCH ₃	$C_{28}H_{40}O_7$	[60]
170	Methyl lucidenate J	β -OH	=O	β -OH	=O	COOCH ₃	$C_{28}H_{40}O_8$	[60]
171	Lucidenic acid H	β -OH	β -OH	Н	=O	COOH	$C_{27}H_{40}O_7$	[60]
172	Lucidenic acid I	β -OH	=O	Н	=O	COOH	C27H38O7	[60]
173	Lucidenic acid J	β -OH	=O	β -OH	=O	COOH	C27H38O8	[60]
174	Methyl lucidenate G	=O	β -OH	Н	a-OH	COOCH ₃	$C_{28}H_{42}O_7$	[94]
175	Lucidenic acid G	=O	β -OH	Н	a-OH	COOH	$C_{27}H_{40}O_7$	[94]

Table 8

G. lucidum triterpenoids 176–178 with skeletal structure 8.

Triterpenoid	General name	R ₁	R_2	Formula	Ref.
176	Lucidone A	<i>β</i> -OH	=O	$C_{24}H_{34}O_5$	[93]
177	Lucidone B	=O	=O	$C_{24}H_{32}O_5$	[93]
178	Lucidone C	OH	OH	$C_{24}H_{36}O_5$	[94]

Table 9

G. lucidum triterpenoids 179–226 with skeletal structure 9.

Triterpenoid	General name	R_1	R ₂	R ₃	R_4	R ₅	R ₆	Formula	Ref.
179	15-Hydroxy-ganoderic acid S	<i>β</i> -OH	a-OH	Н	Н	-CH ₃	COOH	C30H46O4	[63]
180	Ganoderic acid S1	=0	Н	Н	Н	-CH ₃	COOH	$C_{30}H_{44}O_{3}$	[95]
181	Ganoderic acid SZ	=0	Н	Н	Н	-CH ₃	COOH	$C_{30}H_{44}O_{3}$	[96]
182	Ganoderic acid Me	α-OAc	a-OAc	Н	Н	-CH ₃	COOH	$C_{34}H_{50}O_{6}$	[69]
183	Ganoderic acid Mf	α-OAc	α-OH	Н	Н	-CH ₃	COOH	$C_{32}H_{48}O_5$	[69]
184	Ganodermenonol	=0	Н	Н	Н	-CH ₃	CH_2OH	$C_{30}H_{46}O_2$	[97]
185	Ganodermadiol	β -OH	Н	Н	Н	-CH ₃	CH_2OH	$C_{30}H_{48}O_2$	[97]
186	Ganodermatriol	β -OH	Н	Н	Н	CH_2OH	CH_2OH	$C_{30}H_{48}O_3$	[98]
187	Ganodermic acid S	β -OAc	α-OAc	Н	Н	-CH ₃	COOH	$C_{34}H_{50}O_{6}$	[99]
188	Ganoderic acid Mk	-OAc	-OH	-OAc	Н	-CH ₃	COOH	$C_{34}H_{50}O_7$	[70]
189	Ganoderiol B	=O	α-OH	Н	Н	CH ₂ OH	CH_2OH	$C_{30}H_{46}O_4$	[98]
190	Ganoderic acid T	α-OAc	α-OAc	α-OAc	Н	-CH ₃	COOH	$C_{36}H_{52}O_8$	[100]
191	Ganoderic acid S	=O	Н	Н	Н	-CH ₃	COOH	$C_{30}H_{44}O_{3}$	[47,100]
192	Ganoderic acid R	α-OAc	Н	β-OAc	Н	-CH ₃	COOH	$C_{34}H_{50}O_{6}$	[100]
193	/	β -OAc	α-OAc	β -OAc	Н	-CH ₃	COOH	$C_{36}H_{52}O_8$	[101]
194	/	α-OH	α-OAc	=O	Н	-CH ₃	COOH	$C_{32}H_{46}O_{6}$	[101]
195	/	α-OAc	α-OAc	Н	=0	-CH ₃	COOH	$C_{34}H_{48}O_7$	[101]
196	/	α-OAc	α-OH	Н	=0	-CH ₃	COOH	$C_{32}H_{46}O_{6}$	[101]
197	/	α-OAc	α-OH	β -OH	Н	-CH ₃	COOH	$C_{32}H_{48}O_6$	[101]
198	Ganodermic acid T-N	β -OH	a-OAc	Н	Н	-CH ₃	COOH	$C_{32}H_{48}O_5$	[102]
199	Ganodermic acid T-O	β -OAc	α-OH	Н	Н	-CH ₃	COOH	$C_{32}H_{48}O_5$	[102]
200	Ganodermic acid T-Q	=O	α-OAc	Н	Н	-CH ₃	COOH	$C_{32}H_{46}O_5$	[102]
201	Ganoderic acid P	α-OH	α-OAc	β-OAc	Н	-CH ₃	COOH	$C_{34}H_{50}O_7$	[103]
202	Ganoderic acid Q	α-OAc	α-OH	β -OAc	Н	-CH ₃	COOH	$C_{34}H_{50}O_7$	[103]
203	Ganoderiol F	=O	Н	Н	Н	CH ₂ OH	CH_2OH	$C_{30}H_{46}O_3$	[67]
204	/	=O	Н	=O	Н	CH_2OH	CH_2OH	$C_{30}H_{44}O_4$	[104]
205	/	=O	Н	=O	Н	-CH ₃	CH_2OH	$C_{30}H_{44}O_3$	[104]
206	Ganodermic acid P2	β -OH	α-OAc	β-OAc	Н	-CH ₃	COOH	$C_{34}H_{50}O_7$	[105]
207	Ganoderic acid Y	=O	a-OH	Н	Н	-CH ₃	COOH	$C_{30}H_{46}O_3$	[106]
208	Ganoderic acid X	β -OH	Н	Н	Н	-CH ₃	CHO	$C_{32}H_{48}O_5$	[106]
209	Ganoderic acid TR1	α-OH	α-OAc	Н	Н	-CH ₃	COOH	$C_{30}H_{44}O_4$	[107]
210	23-Hydroxyganoderic acid S	=O	β -OH	Н	Н	-CH ₃	COOH	$C_{30}H_{46}O_4$	[107]
211	Ganoderic aldehyde TR	OH	Н	Н	OH	-CH ₃	COOH	$C_{30}H_{44}O_3$	[107]
212	/	α-OH	a-OH	a-OH	Н	-CH ₃	COOH	$C_{30}H_{46}O_5$	[108]
213	/	β -OH	a-OH	β -OH	Н	-CH ₃	COOH	$C_{30}H_{46}O_5$	[108]
214	/	α-OAc	a-OAc	a-OH	Н	$-CH_3$	COOH	$C_{34}H_{50}O_7$	[108]
215	/	β -OAc	a-OAc	a-OH	Н	-CH ₃	COOH	$C_{34}H_{50}O_7$	[108]
216	/	α-OH	a-OH	β -OAc	Н	-CH ₃	COOH	$C_{32}H_{48}O_6$	[108]
217	/	β -OAc	a-OAc	Н	Н	-CH ₃	COOH	$C_{34}H_{52}O_{6}$	[108]
218	/	β -OH	a-OH	β -OAc	Н	$-CH_3$	COOH	$C_{32}H_{48}O_6$	[108]
219	Lucialdehyde A	β -OH	Н	Н	Н	-CH ₃	CHO	$C_{30}H_{46}O_2$	[49]
220	Ganoderal A	=O	Н	Н	Н	-CH ₃	CHO	$C_{30}H_{44}O_2$	[47]
221	Ganoderic acid TR	=O	a-OH	Н	Н	-CH ₃	COOH	$C_{30}H_{44}O_4$	[35]
222	Ganodermic acid Ja	a-OH	a-OH	Н	Н	$-CH_3$	COOH	$C_{30}H_{46}O_4$	[105]
223	Ganodermic acid Jb	β -OH	a-OH	Н	Н	$-CH_3$	COOH	$C_{30}H_{46}O_4$	[105]
224	/	=O	a-OH	Н	Н	$-CH_3$	COOH	$C_{30}H_{44}O_4$	[33]
225	/	=O	a-OH	Н	Н	$-CH_3$	CH_2OH	$C_{30}H_{46}O_{3}$	[33]
226	/	<i>β</i> -OH	Н	Н	Н	-CH ₃	COOH	$C_{30}H_{46}O_3$	[33]

Table 10	
G. lucidum triterpenoids 227–234 with skeletal structure 10	

Triterpenoid	General name	R_1	R_2	R ₃	R_4	R ₅	R_6	R ₇	Formula	Ref.
227	Lucidumol B	β -OH	Н	Н	<i>β</i> -OH	-OH	-CH ₃	-CH ₃	$C_{30}H_{50}O_{3}$	[66]
228	Ganodermanontriol	=0	Н	Н	β -OH	α-OH	CH_2OH	β -CH ₃	$C_{30}H_{48}O_4$	[62]
229	Ganoderiol A	β -OH	Н	Н	β -OH	-OH	CH_2OH	$-CH_3$	$C_{30}H_{50}O_4$	[98]
230	Ganodermanondiol	=0	Н	Н	β -OH	-OH	$-CH_3$	$-CH_3$	$C_{30}H_{48}O_3$	[109]
231	Ganodermanontetrol	=0	Н	Н	α-ОН	CH ₂ OH	CH_2OH	-OH	$C_{30}H_{48}O_5$	[83]
232 12 <i>α</i> -	Methoxy-ganodermanondiol	=0	α -OCH ₃	Н	α-ОН	-OH	$-CH_3$	$-CH_3$	$C_{31}H_{50}O_4$	[83]
233 15α-	Hydroxy-ganodermanontriol	=0	Н	α-ОН	α-ОН	α-OH	CH_2OH	β -CH ₃	$C_{30}H_{48}O_5$	[83]
234	1	β -OH	Н	Н	α-ОН	-OCH ₃	CH ₂ OH	-CH ₃	$C_{31}H_{52}O_2$	[81]

G. lucidum triterpenoids 235–266.

Triterpenoid	Chemical name	General name		Ref.
235	(7β,12β,16α,23β,25R)-7,12,23-trihydroxy-3,11,15-trioxo- 16,23-cyclolanost-8-en-26-oic acid γ-lactone	Ganosporelactone A	$C_{30}H_{40}O_7$	[110]
236	(3β,7β,12β,16α,23β,25R)-3,7,12,23-tetrahydroxy-11,15-dioxo- 16,23-cyclolanost-8-en-26-oic acid γ-lactone	Ganosporelactone B	$C_{30}H_{42}O_{7} \\$	[110]
237	(24S,25S)-24,25-epoxy-26-hydroxylanosta-7,9(11)-dien-3-one	Epoxyganoderiol B	$C_{30}H_{46}O_3$	[68]
238	(3β,24S,25S)-24,25-epoxylanosta-7,9(11)-diene-3,26-diol	Epoxyganoderiol C	$C_{30}H_{48}O_3$	[68]
239	(3β,7β,12β)-3,7,12,20-tetrahydroxy-11,15,23-trioxolanost-8-en-26-oic acid	20-Hydroxylganoderic acid G	$C_{30}H_{44}O_{9}$	[61]
240	(3 <i>β</i> ,7 <i>β</i> ,25 <i>R</i>)-3,7,20-trihydroxy-11,15,23-trioxolanost-8-en-26-oic acid	Ganoderic acid I	$C_{30}H_{44}O_8$	[46]
241	(3β) -3,26,27-trihydroxylanosta-8,24-dien-7-one	Ganoderiol E	$C_{30}H_{48}O_4$	[67]
242	$(7\alpha, 15\alpha)$ -15,26,27-trihydroxy-7-methoxylanosta-8,24-dien-3-one	Ganoderiol I	$C_{31}H_{50}O_5$	[67]
243	(7a,24S,25S)-24,25-epoxy-7,26-dihydroxylanost-8-en-3-one	Epoxyganoderiol A	$C_{30}H_{48}O_4$	[68]
244	$(3\beta, 20\xi, 24E)$ -3,20-dihydroxy-7,11,15-trioxolanosta-8,24-dien-26-oic acid	Ganoderic acid V1	$C_{30}H_{42}O_{7}$	[111]
245	(12β) -12-hydroxy-3,7,11,15,23-pentaoxolanostan-26-oic acid	8β ,9 α -Dihydroganoderic acid C	$C_{30}H_{42}O_8$	[57]
246	(5α)-4,4,14-trimethyl-3-oxo-chola-7,9(11)-dien-24-oic acid	/	$C_{27}H_{40}O_3$	[95]
247	Methyl 7 β ,15 α -isopropylidenedioxy3,11,23-trioxo-5 α -lanost-8-en-26-oate	Methyl ganoderate A acetonide	$C_{34}H_{50}O_7$	[39]
248	(5 <i>α</i> ,7 <i>β</i> ,20ζ)-7-hydroxy-4,4,14-trimethyl-20,24-epoxychol-8-ene-3,11,15,24-tetrone	Ganolactone	$C_{27}H_{36}O_{6}$	[112]
249	$(3\beta, 12\beta, 25R)$ -methyl 3,12-bis(acetyloxy)-7,11,15,23-tetraoxolanostan-26-oate	Methyl O-acetylganoderate C	$C_{35}H_{50}O_{10}$	[113]
250	$(3\beta,4\alpha,5\alpha,7\beta,15\alpha)$ -3,7,15-trihydroxy-4-(hydroxymethyl)-4,14-dimethyl-11-oxochola-8,20-dien-24-oic acid	Lucidenic acid O	$C_{27}H_{40}O_7$	[114]
251	(25R)-3,7,11,15,23-pentaoxolanostan-26-oic acid	8β ,9 α -Dihydroganoderic acid C	$C_{30}H_{42}O_7$	[57]
252	(8α,20ξ)-8,9-epoxy-3,7,11,15,23-pentaoxolanostan-26-oic acid	1	$C_{30}H_{40}O_8$	[115]
253	$(3\beta, 4\beta, 15\alpha)$ -methyl-3,15,28-trihydroxy-11,23-dioxolanost-8-en-26-oate	Methyl Ganolucidate C	$C_{31}H_{48}O_7$	[116]
254	$(3\beta,4\beta,15\alpha)$ -3,15,28-trihydroxy-11,23-dioxolanost-8-en-26-oic acid	Ganolucidic acid C	$C_{30}H_{46}O_7$	[116]
255	12β -acetoxy- 3β ,25-dihydroxy-7,11,15-trioxo-lanost-8-en-26-oic acid	/	$C_{32}H_{46}O_9$	[117]
256	1	Ganosidone A	$C_{24}H_{32}O_5$	[118]
257	1	Ganotropic acid	$C_{30}H_{44}O_7$	[59]
258	$(3\beta,5\alpha,7\beta,12\beta)-\gamma-lactone-3,7,12,20-tetrahydroxy-4,4,14-trimethyl-11,15-dioxochol-8-en-24-oic acid$	Ganoderlactone D	$C_{27}H_{38}O_7$	[80]
259	(3β,5α,17E)-3-hydroxy-7,11,15,23-tetraoxolanosta-8,17(20)-dien-26-oic acid	/	$C_{30}H_{40}O_7$	[80]
260	$(3\beta,12\beta,17E)-12-(acetyloxy)-3-hydroxy-7,11,15,23-tetraoxolanosta-8,17(20)-dien-26-oic acid$	/	$C_{32}H_{42}O_9$	[80]
261	$(4\alpha, 15\alpha, 20\xi, 24E)$ -15,20,28-trihydroxy-3,11-dioxolanosta-8,24-dien-26-oic acid	Ganoderic acid	$C_{30}H_{44}O_7$	[119]
262	20,21,22,23,24,25,26,27-octo-nor-15α-hydroxylanosta-8-en-3,11,17-trione	Ganoluciduone A	$C_{22}H_{30}O_4$	[81]
263	(3β,23 <i>E</i>)-3-hydroxy-27-norlanosta-7,9(11),23-trien-25-one	Ganoluciduone B	$C_{29}H_{44}O_2$	[81]
264	$(3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetyloxy)-3,7-dihydroxy-4,4,14-trimethyl-pregn-8-ene-11,15,20-trioned (3\beta,7\beta,12\beta)-12-(acetylox)-12-(ac$	Ganolucidoid A	$C_{26}H_{36}O_7$	[81]
265	$(3\beta, 12\beta)$ -12-acetyloxy)-3-hydroxy-4,4,14-trimethyl-pregn-8-ene-7,11,15,20-tetrone	Ganolucidoid B	$C_{26}H_{34}O_7$	[81]
266	24,25-epoxy-26,27-dihydryoxy-lanosta-7,9(11)-dien-3-one	1	$C_{30}H_{46}O_4$	[81]





Table 12		
Polysaccharid	e isolated from G	E. lucidum.

No.	Origin	Main glycosidic bonds	Sugar composition	Molecular weight	Ref.
1	G. lucidum mycelium	1	G. lucidum polysaccharide (GLP) I: arabinose, rhamnose, xylose, mannose, glucose GLPII: arabinose, xylose, glucose GLPIII: arabinose, rhamnose, xylose, galactose, mannose, glucose GLPIV: arabinose, rhamnose, fucose, mannose, glucose	/	[120]
2	G lucidum fruit body	β (1-3)	Glucose	3.979×10^3 Da	[120]
3	<i>G</i> lucidum fruit body	$\beta(1-3)$ (1-4) (1-6)	GLP 1: glucose: GLP 2: glucose galactose mannose	GLPL 1: 52×10^3 Da: GLPL 2: 154×10^3 Da	[120]
4	G. lucidum fruit body	p (1 5), (1 -), (1 -))	Mannose	/	[120]
5	G. lucidum fruit body	/	Mannose	/	[120]
6	<i>G. lucidum</i> fruit body	/	Rhamnose, xylose, fructose, galactose, mannose, glucose	5.85×10^{2} Da	[120]
7	G. lucidum fruit body	β (1-3), (1-4), (1-6) α (1-4)	Glucose, xylose, arabinose	4×10^4 Da	[120]
8	G. lucidum fruit body	β (1–3), a few short (1–4)	Glucose, galactose, mannose, xylose, arabinose, fucose	/	[120]
9	G. lucidum mycelium	β (1-3)	Glucose	/	[120]
10	G. lucidum fruit body	β (1–3), with β (1–6) branches	Glucose	$GLPO < 1.2 \times 10^4 \text{ Da};$ $GLPI > 1.2 \times 10^4 \text{ Da}$	[120]
11	G. lucidum fruit body	β (1–3), (1–6)	Glucose	Ganoderan B: 7.41×10^3 Da; Ganoderan C: 5.81×10^3 Da	[121]
12	G. lucidum fruit body	/	GLB ₆ : arabinose, xylose, galactose, mannose, glucose GLB ₁₀ :rhamnose, arabinose, xylose, galactose, mannose, glucose	GLB6: 8.8×10^3 Da; GLB7: 9.0×10^3 Da; GLC1: 5.7×10^3 Da	[122]
13	G. lucidum fruit body	TGLP-2 (TGLP-6): β (1–3), (1–6) TGLP-3 (TGLP-7): β (1–3), β (1–4), (1–6)	TGLP-2 (TGLP-3, TGLP-6): glucose TGLP-7: galactose	TGLP-2: 20.9×10^4 Da; TGLP-3: 4.5 × 10^4 Da; TGLP-6: 3.2×10^4 Da; TGLP-7: 10.0×10^4 Da	[123]
14	G. lucidum fruit body	PL-1: β (1–4), (1–6) PL-3: β (1–3), (1–6) PL-4: β (1–3), (1–4), (1–6)	PL-1: glucose PL-3: glucose PL-4: glucose, mannose	PL-1: 8.3 × 10 ³ Da; PL-3: 6.3 × 10 ⁴ Da; PL-4: 2.0 × 10 ⁵ Da	[124]
15	G. lucidum spores	β (1-3)	Glucose	GSPL-I-1A: 7.8×10^5 Da	[125]
16	G. lucidum fruit body	/	Glucose, galactose, mannose	GLIS: -	[126]
17	G. lucidum fruit body	/	Rhamnose, xylose, fructose, galactose, mannose, glucose	GLPG: 5.13×10^{5} Da GLPW: 5.85×10^{5} Da	[127]
18	G. lucidum fruit body	β (1-3), (1-6)	Rhamnose, xylose, galactose, mannose, glucose	GL-PP-3A: 1.11×10^4 Da	[128]
19	<i>G</i> lucidum mycelia		Rhamnose xylose galactose glucose	SeGI P-2B-1: 1.06 x 10 ⁶ Da	[120]
20	<i>G. lucidum</i> fruit body		/	$\begin{array}{l} \text{GL-IV-1:} \ 13.3 \times 10^4 \ \text{Da; S-GL:} \ 10.1 \times 10^4 \ \text{Da;} \\ \text{CM-GL:} \ 6.3 \times 10^4 \ \text{Da; HE-GL:} \\ 7.2 \times 10^4 \ \text{Da; HP-GL:} \ 5.1 \times 10^4 \ \text{Da;} \end{array}$	[130]
21	G. lucidum fruit body	β (1-3), (1-6)	Glucose	M-GL: 14.1 × 10 ⁴ Da GLP20: 3.75 × 10 ⁶ Da	[131]
22	G. lucidum extract	β (1–3), (1–6)	GLP-1: glucose, galactose; GLP-2: glucose, galactose	GLP-1: 1.07×10^5 Da; GPL-2: 1.95×10^4 Da	[132]

Steroids in G. lucidum.

No.	Chemical name	General name	Formula	Ref.
1	(3β,22E)-ergosta-5,7,22-trien-3-ol	Ergosterol	$C_{28}H_{44}O$	[133-134]
2	$(3\beta,5\alpha,22E)$ -ergosta-7,22-dien-3-ol	Stellasterol	$C_{28}H_{46}O$	[134]
3	3β , 5α -dihydroxy- 6β -methoxy-ergosta-7,22-diene	6-O-Methylcerevisterol	$C_{29}H_{48}O_3$	[41]
4	$(3\beta, 5\alpha, 22E)$ -3,5-dihydroxy-ergosta-7,22-dien-6-one	6-Dehydrocerevisterol	$C_{28}H_{44}O_3$	[135]
5	$(3\beta,5\alpha,22E)$ -3,5,9-trihydroxyergosta-7,22-dien-6-one	/	$C_{28}H_{44}O_4$	[135]
6	(3β,5α,6β,22 <i>E</i>)-ergosta-7,22-diene-3,5,6-triol	Cerevisterol	$C_{28}H_{46}O_3$	[133]
7	(3β,5α,6β,22E)-ergosta-7,22-diene-3,5,6,9-tetrol	9-Hydroxycerevisterol	$C_{28}H_{46}O_4$	[135]
8	(3β,5α,6β,20R,22E)-ergosta-7,22-diene-3,5,6,9,14-pentol	/	$C_{28}H_{46}O_5$	[135]
9	(6α,22E)-6-hydroxy-ergosta-4,7,22-trien-3-one	/	$C_{28}H_{42}O_2$	[68]
10	(6β,22E)-6-hydroxyergosta-4,7,22-trien-3-one	/	$C_{28}H_{42}O_2$	[68]
11	Ergosta-4,7,22-triene-3,6-dione	/	$C_{28}H_{40}O_2$	[75]
12	(15α,22E)-15-hydroxy-ergosta-4,6,8(14),22-tetraen-3-one	Ganodermaside A	$C_{28}H_{40}O_2$	[136]
13	(15β,22E)-15-hydroxy-ergosta-4,6,8(14),22-tetraen-3-one	Ganodermaside B	$C_{28}H_{40}O_2$	[136]
14	(22E)-9-hydroxyergosta-4,6,8(14),22-tetraene-3,15-dione	Ganodermaside C	C28H38O3	[137]
15	(22E)-9-hydroxyergosta-4,6,8(14),22-tetraen-3-one	Ganodermaside D	$C_{28}H_{40}O_2$	[137]
16	Ergosta-7,22-dien-3-one	/	$C_{28}H_{44}O$	[93]
17	Ergosta-7,22-diene-3β-yl palmitate	/	$C_{44}H_{72}O_{2}$	[93]
18	Ergosta-7,22-dien-3β-yl linoleate	/	$C_{46}H_{76}O_2$	[138]
19	5α , 8α -epidioxyergosta-6, 22-dien- 3β -ol	Ergosterol peroxide	C28H44O3	[97]
20	Ergosta-5,7-dien-3β-ol	22,23-Dihydroergosterol	$C_{28}H_{46}O$	[71]
21	(3β)-stigmast-5-en-3-ol	β -Sitosterol	$C_{29}H_{50}O$	[133]
22	(3β,5α,8α,22E)-5,8-epidioxy-ergosta-6,9(11),22-trien-3-ol	9,11-Dehydroergosterol peroxide	C28H42O3	[139]
23	(22E)-ergosta-4,6,8(14),22-tetraen-3-one	/	$C_{28}H_{40}O$	[71]
24	Ergosta-7,22-diene-3 β -yl pentadecanoate	/	C43H74O2	[140]
25	5a,8a-epidioxyergosta-6,22-dien-3β-yl linoleate	/	C46H74O4	[138]
26	3β -linoleyloxyergost-7-ene	/	C46H78O2	[140]
27	$(3\beta,5\alpha)$ -ergost-7-en-3-ol	Fungisterol	C28H48O	[71]

Table 14Other compounds in G. lucidum.

No.	Chemical name	General name	Formula	Ref.
	Meroterpenoids			
1	(2Z,5E)-2-[2-(2,5-dihydroxyphenyl)ethylidene]-6,10-dimethyl-5,9-undecadienoic acid	Ganomycin B	$C_{21}H_{28}O_4$	[141]
2	2,5-dihydroxy-α-(4-methyl-3-penten-1-yl)-γ-oxo-benzenebutanoic acid	Chizhine D	$C_{16}H_{20}O_5$	[142]
3	(2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxy - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl) ethylidene] - 9, 10 - dihydroxyphenyl - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 9R) - 2 - [2 - (2, 5 - dihydroxyphenyl - 6, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E, 10 - dimethyl - 5 - undecenoic acid (2Z, 5E,	Ganomycin J	$C_{21}H_{30}O_6$	[141]
4	(2E) - 5 - [(5R) - 5 - (2,5 - dihydroxyphenyl) - 2,5 - dihydro - 2 - oxo - 3 - furanyl] - 2 - methyl - 2 - pentenal - 2 - pe	(+)-Chizhine E	$C_{16}H_{16}O_5$	[142]
5	(5R) - 5 - (2, 5 - dihydroxyphenyl) - 3 - [(3E) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 2(5H) - furanone - 2(5H) - 2	Ganomycin I	$C_{21}H_{26}O_5$	[142]
6	(5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 5 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 4,8 - dimethyl - 6 - 0xo - 3,7 - nonadien - 1 - yl] - 2(5H) - fur an one (5R) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - (2,5 - dihydroxyphenyl) - 3 - [(3E) - 2 - (2,5 - dihydroxyphenyl) - 3 - (3E) - ((+)-Chizhine F	$C_{21}H_{24}O_5$	[142]
7	(+) - 5 - (2, 5 - dihydroxyphenyl) - 3 - [(3E) - 4, 8 - dimethyl - 3, 7 - nonadien - 1 - yl] - 5 - methoxy - 2(5H) - fur an one of the second secon	Fornicin B	$C_{22}H_{28}O_5$	[142]
8	(2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydro-5-methoxy-2-oxo-3-furanyl]-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydro-5-methoxy-2-oxo-3-furanyl]-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydro-5-methoxy-2-oxo-3-furanyl]-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydro-5-methoxy-2-oxo-3-furanyl]-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydro-5-methoxy-2-oxo-3-furanyl]-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl-2-methyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl)-2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-(2,5-dihydroxyphenyl-2-pentenal (2E)-(+)-5-[5-(2,5-(2E)-2-pentenal (2E)-(+)-5-[5-(2,5-(2E)-2-pentenal (2E)-(+)-5-[5-(2E)-2-pentenal (2E)-(+)-5-[5-(2E)-2-pentenal (2E)-5-[5-(2E)-2-pentenal (2E)-5-[5-(2E)-5-[5-(2E)-2-pentenal (2E)-5-[5-(2E)-5-[5-(2E)-5-[5-(2E)-5-[5-(2E)-5-5-[5-(2E)-5-5-[5-(2E)-5-5-5-5-5-5-5-5-5-5-5-5-5-5-5-5-5-5-5	(+)-Lucidulactone B	$C_{17}H_{18}O_6$	[143]
9	(3R,5R) - rel - 3 - [2 - (2,5 - dihydroxyphenyl) - 2 - oxoethyl] dihydro - 5 - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - rel - 3 - [2 - (2,5 - dihydroxyphenyl) - 2 - oxoethyl] dihydro - 5 - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - rel - 3 - [2 - (2,5 - dihydroxyphenyl) - 2 - oxoethyl] dihydro - 5 - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - fur an one (3R,5R) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - (2 - methyl - 1 - propen - 1 - yl) - 2(3H) - 2(3	Chizhine A	$C_{16}H_{18}O_5$	[142]
10	$(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-rel-3-[2-(2,5-dihydroxyphenyl)-2-oxoethyl]dihydro-5-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-furanone(3R,5S)\-rel-3-(2-methyl-1-propen-1-yl)-2(3H)\-furanone(3R,5S)\-fura$	Chizhine B	$C_{16}H_{18}O_5$	[142]
11	(3R, 6R) - rel - 3 - [2 - (2, 5 - dihydroxyphenyl) - 2 - oxoethyl] tetrahydro - 3 - hydroxy - 6 - (1 - methylethenyl) - 2H - pyran - 2 - one (3R, 6R) - 2H - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2 - one (3R, 6R) - 2H - pyran - 2H -	Chizhine C	$C_{16}H_{18}O_{6}$	[142]
12	(1R,2S) - 1 - [2 - (2,5 - dihydroxyphenyl) - 2 - oxoethyl] - 2 - hydroxy - 3 - methyl - 3 - cyclohexene - 1 - carboxylic acid acid acid acid acid acid acid ac	Chizhiol A	$C_{16}H_{18}O_{6}$	[144]
13	rel-(3aR,9bR,10R)-6,9,10-trihydroxy-1H,3H-3a,9b-propanonaphtho [1,2-c] furan-3,5(4H)-dioned and a standard st	(±)-Lingzhiol	$C_{15}H_{14}O_{6}$	[145]
	Alkaloids			
1	(1S)-1,2,3,4-tetrahydro-1-methoxy [1] benzopyrano [4,3-b] cyclopenta [d] pyridin-8-old and a start of the s	Lucidimine A	$\mathrm{C_{16}H_{15}NO_{3}}$	[146]
2	1,2,3,4-tetrahydro[1]benzopyrano[4,3-b]cyclopenta[d]pyridin-8-ol	Lucidimine B	$\mathrm{C_{15}H_{13}NO_{2}}$	[146]
3	(-)-1,2,3,4-tetrahydro-4-methoxy [1] benzopyrano [4,3-b] cyclopenta [d] pyridin-8-old (-)-1,2,3,4-tetrahydro-8-b] cyclopenta [d] pyridin-8-b] cyclopenta [d] cyclopen	Lucidimine C	$\mathrm{C_{15}H_{15}NO_{3}}$	[146]
4	1	Lucidimine D	$\mathrm{C_{17}H_{17}NO_4}$	[146]
5	1,2,3,4-tetrahydropyrimidine-2,4-dione	Uracil	$C_4H_4N_2O_2$	[147]
6	1 - β - D -ribofuranosyl-2,4(1H,3H)-pyrimidinedione	Uridine	$C_9H_{12}N_2O_6$	[147]
7	9H-purin-6-amine	Adenine	$C_5H_5N_5$	[147]
8	1	Adenosine	$C_{10}H_{13}N_5O_4\\$	[147]
9	$(4E,8E)-N-D-2'-hydroxypalmitoyl-1-O-\beta-D-glucopyranosyl-9-methyl-4,8-sphingadienine$	/	C41H77NO9	[148]
10	$(4E, 8E)$ -N-D-2'-hydroxystearoyl-1-O- β -D-glucopyranosyl-9-methyl-4,8-sphingadienine	/	$\mathrm{C}_{43}\mathrm{H}_{81}\mathrm{NO}_9$	[148]

2.2 Quality control methods

2.2.1 Small molecular substances in G. lucidum

As mentioned before, the small molecule compounds in G. lucidum are composed of triterpenes, nucleosides, alkaloids, and sterols, as well as some other types of molecules. Among them, triterpenoids are a class of important biologically active ingredients, which have strong physiological activities, such as anti-inflammatory^[149-150], anti-tumor^[24], protecting liver^[151] and gallbladder^[152], cytotoxic^[33], α -glucosidase inhibitory^[153], anti-malaria^[154], lowering cholesterol^[155], lowering blood fat^[156], lowering blood sugar^[15], etc. Therefore, there are relatively more studies on the quality control of triterpenoids in G. lucidum at present. Moreover, some reports focus on polar compounds such as nucleosides and nucleobases, which can participate in the regulation of various physiological processes, such as inhibiting platelet aggregation^[157] and reducing memory deterioration in old senescence-accelerated mice^[158]. Most of the classical methods of quality control research in the past 15 years on G. lucidum small molecule substances are listed in Table 15, and the corresponding extraction and analytical methods are also summarized.

In general, the small molecules in *G. lucidum* are extracted by organic solvents or water, and the extraction solvent is selected by the requirement of polarity. Methanol^[159-162], chloroform^[162-166], ethanol^[167-169], 95% ethanol^[170-173] or acetonitrile/water^[174] are favored by most researchers when extracting triterpenoids, while water extraction^[168,175] is the most common method for extracting nucleosides. In addition, researchers often use ultrasonic extraction^[160,162,164-166,172-173,175], heating extraction^[170,173,176],

reflux extraction^[159,163,173], shaking^[169,171], vortexing^[176] and other methods such as accelerated solvent extraction (ASE) system^[161] to improve the extraction efficiency. After extraction, researchers would often use silica gel column chromatography or preparative high-performance liquid chromatography (HPLC) to further purify the isolated triterpene fractions due to the insufficient sensitivity of instrument^[22].

With the rise of fingerprints in the field of quality control of traditional Chinese medicine and the renewal of analytical instruments, researchers are more inclined to directly analyze the extracts of herbs through instruments. Many previous studies^[159-161,163-168,170-178], as shown in Table 15, have also focused on the quantitative measurement of the chemical composition of Ganoderma, to distinguish G. lucidum from different species or origins and to control the quality of herb of the same batch. And in terms of the instruments employed in the research, ultraviolet-visible spectroscopy (UV-Vis), HPLC, infrared spectroscopy (IR) and Raman spectra, near-infrared (NIR) spectroscopy, gas chromatography (GC), ultraperformance liquid chromatography (UPLC) and high-performance thin-layer chromatography (HPTLC) are the popular ones. Abundant detectors have also provided favorable conditions for the detection of different types of compounds, such as ultraviolet detector/ ultraviolet-visible detector (UV/UVD), mass spectrometry (MS), multilevel mass spectrometry (MS_n), etc. These instruments have their advantages and disadvantages. Among them, ultraviolet-visible spectrophotometry is an analytical method that has been utilized extensively for decades, which has the advantages of low analysis cost and wide applicable concentration range. For the quality control of Ganoderma in the 2020 edition of the Chinese Pharmacopoeia,

the total contents of triterpenoids and sterols were measured by ultraviolet-visible spectrophotometry^[179]. The authenticity of G. lucidum spore powder samples can be identified by NIR spectroscopy, and its adulteration content can be predicted^[180]. LC-MS can also achieve high sensitivity, improve the analysis efficiency and obtain specific compound information. This method has been used by the United States Pharmacopoeia to specify the retention time of specific ganoderic acid, ganoderic acid and monosaccharide, such as ganoderic acid A, B, C, D and ganoderenic acid B, C, D and mannose, galactose^[181] and etc. Zhang et al.^[162] combined the HPLC-MS analysis of the Ganoderma extract with the pharmacological activity research on three cancer cells and found that 6 compounds for example 12-acetoxyganoderic acid F and ganoderic acid A had significant anti-proliferative activities. Compare with LC-MS, GC-MS has higher separation efficiency for sterols analysis than LC-MS, and exhibits high sensitivity, high accuracy and high interference removal ability, which are very suitable for quantifying several steroid compounds simultaneously^[182]. Moreover, the sterol chromatogram combined with pattern recognition analysis,

Table 15

Small molecule marker compounds in G. lucidum.

such as hierarchical clustering analysis (HCA) and discriminant analysis (DA), could distinguish Ganoderma from different species and regions^[170]. IR and Raman spectroscopy could overcome the shortcomings of the large number of organic solvents required for LC with a simplified sample preparation process. HPTLC method also uses less solvent and reduced the time for analysis and sample preparation^[169]. The result of fingerprinting is a complex and broad data matrix, which is characterized by a large number of variables^[159]. Principal component analysis (PCA)^[159,166,168,172,177], partial least squares discriminant analysis (PLS-DA)^[159,162], HCA^[159,170-171,174-175], DA^[170-171,177], soft independent modeling of class analogy (SIMCA)^[159] and cluster analysis (CA)^[166,172-173] have been regarded as the feasible data analysis methods in massive and complex data. Among them, PCA and CA mainly focus on the features that contribute most to the variance by highlighting the high or low regulatory levels of certain metabolites. And HCA, DA and SIMCA could be used to distinguish or predict different samples^[183]. Furthermore, PLS-DA can quantitatively predict the target component in the mixture through a large amount of data.

	1					
No.	Sample	Extraction method	Instrument	Date analytical method	Marker compound or biomarker	Ref.
1	Fruit bodies	Chloroform; Ultrasonic extraction	RP-HPLC	/	Ganoderic acids C2, B, AM ₁ , K, H and D	[165]
2	176 Samples from different origins	Crushing and grinding samples to get powder	NIR	PCA; DPLS; DA	Total triterpenoid saponins	[177]
3	29 Fruit bodies from different cultivation places	Chloroform; Reflux extraction	HPLC-PAD;	/	The peaks of ganoderic acid B, C2, AM ₁ , H, F and ganoderic acid D, K in the fingerprint	[163]
4	80 Samples from different origins	Methanol; Reflux extraction	HPLC	HCA; PCA; PLS-DA; SIMCA	19 Common peaks without specific components in fingerprint	[159]
5	Fruit bodies of 15 strains	95% Ethanol; Shaking	HPLC	HCA; DA	32 Characteristic peaks of unconfirmed components in fingerprint	[171]
6	Fruit bodies of 23 batches	ACN/water (50:50, V/V)	LC-UV (HILIC/RP modes)	HCA	Characteristic peaks of unidentified components in fingerprint	[174]
7	Fruit bodies and whole spores from different provinces	Chloroform; Ultrasonic extraction	LC-MS/MS	/	Ganoderic acid C2, B, A, H, D	[164]
8	12 Batches of <i>G. lucidum</i> and 4 batches of <i>G. atrum</i>	95% Ethanol; Heating extraction	GC-MS	HCA; DA	Ergosterol, ergosta-7,22-dien-3β-ol, Ergosta-7-en-3β- ol, ergosta-4,6,8(14),22-tetraen-3-one	[170]
9	20 Samples from different origins	Water; Ultrasonic extraction	ZIC-HILIC	HCA	16 Nucleosides and nucleobases such as adenine, uracil, thymine	[175]
10	20 Batches of <i>G. lucidum</i> and 12 batches of <i>G. sinense</i>	Ethanol	HPLC-VWD; UPLC- PDA	PCA	Ganoderic acid C2, G, B, K, A, H, D, F and ganoderenic acid A, B, C, D	[168]
11	G. lucidum and G. sinense	Water; Heating and vigorous vortex	Multiple columns and detectors LC system	/	Uracil, cytidine, adenosine, ganoderic acid C2, G, B, A, K, D, E, ganoderenic acid B, K, D and so on	[176]
12	20 Batches of G. lucidum	Ethanol	UPLC-SSDMC	/	Ganoderenic acid C, B, D and ganoderic acid C2, G, B, A, H, D, F	[167]
13	15 Ganoderma species from different sources	Methanol; Ultrasonication method	HPLC-DAD	/	Uracil, adenosine, ergosterol and 12 triterpenes (for example: lucidenic acid N, E2, A, ganoderic acid A, E)	[160]
14	Fruit body of <i>G. sinensis</i> , and <i>G. lucidum</i>	Methanol; ASE	HPLC	/	Adenine and adenosine	[161]
15	15 Samples from different regions	Chloroform; Ultrasonic extraction	HPLC and HPLC-MS ⁿ	CA; PCA	Ganoderic acid B, 3,7,15-trihydroxy-11,23-dioxolanost- 8,16-dien-26-oic acid, lucidenic acid A, ganoderic acid G, and 3,7-oxo-12-acetylganoderic acid DM	[166]
16	39 Fruit bodies of different regions	95% Ethanol; Ultrasonic extraction	HPLC	PCA; CA	Ganoderic acid A and total triterpene	[172]
17	10 Ganoderma samples from different regions	95% Ethanol; Heating Reflux Extraction and ultrasonic Extraction	HPLC-LTQ Orbitrap- MS ⁿ ; TLC	CA	Lucidenic acid LM1, E, A, D, ganoderic acid G, B, A, D, F, ganoderenic acid A, D	[173]
18	Different types of ganoderic acids	Mixed with KBr	IR and Raman spectra	/	Ganoderic acid A and other ganoderic acids, ganoderenic acid A	[178]
19	6 Different cultivars and origins of Ganoderma species	Chloroform; Ultrasonication method	HPLC-UV/ESI-MS/MS	S PLS-DA	12-Acetoxyganoderic acid F, Ganoderic acid A, etc.	[162]
20	50 Samples of G. lucidum fruit body	Ethanol; Shaking	HPTLC	/	Ergosterol, ganoderic acid D, A, G, C2	[169]

However, due to the complexity of chemical components in Chinese medicinal materials, it is a challenge to achieve convincing quality control only from the content of certain compounds. Fingerprints can simultaneously judge the content of multiple complex components in traditional Chinese medicine, instead of just taking the content of a few specific components as the criterion^[159]. Since 2008, the fingerprint of the chloroform and methanol extracts of G. lucidum has been studied to distinguish the chemical composition of Ganoderma from different regions through similarity analysis^[159,163]. In the past decade, a variety of detectors and chromatographic columns have been successively used for fingerprint study. As more small molecular compounds in G. lucidum have been found, the compounds contained in the fingerprint have also shown the characteristics of its complexity and diversity. Nevertheless, studying the fingerprint of small-molecule compounds of G. lucidum merely based on the different chemical components could only provide chemical markers for identifying G. lucidum from different regions or different parts. With the proposal of the concept of quality markers, the original strategy based on representative chemical components for content determination has changed to an overall strategy based on chemical components combined with pharmacological activities. The extracts from different species and different parts of Ganoderma were quantitatively analyzed by HPLC-MS, and combined with the anti-proliferative activity study, quality markers for cancer treatment were found^[162]. When the fingerprint analysis was combined with the pharmacological study to form a spectrum-activity relationship, the obtained quality marker would be more convincing.

2.2.2 Macromolecular substances in G. lucidum

The macromolecular compounds in *G. lucidum* are mainly carbohydrates, including glycoproteins and polysaccharides^[184], in which polysaccharides represent one of the most abundant components in *G. lucidum*^[37]. Since Miyazaki et al.^[185] reported that a branched arabinoxyloglucan isolated from *G. lucidum* fruit body possesses the anti-tumor effect, researchers have become more interested in polysaccharides with more studies only their chemical structure and activity. Therefore, the quality control of polysaccharides in *G. lucidum* has become an important criterion of its quality.

The extraction methods of GLP mainly include hot water extraction, salt solution extraction, alkaline solution extraction and so on. Water extraction conforms to the traditional Chinese medicine decoction principle and is the most common basic extraction method. And the salt solution can remove the influence of protein by 0.9% sodium chloride^[186]. Water-insoluble polysaccharide could also be obtained by alkaline solution extraction, which needs to be combined with sulfate modification polysaccharide method^[187]. Since GLP are also insoluble in alcohol, the isolation method normally involves water extraction and alcohol precipitation^[37]. In addition, ultrasonic^[188] and microwave-assisted extraction^[189-191] are also new methods for isolating GLP. The commonly used method of purifying polysaccharides is column chromatography, including anion exchange column chromatography^[124,188-189], gel permeation chromatography^[124,188] and affinity chromatography. In the meantime,

researchers use fermentation methods to purify polysaccharides, such as fermented soybean curd residue^[192].

Different from the analysis of small molecular compounds with single type of analysis method, GLP requires the combination of multiple analysis methods to characterize the various information of it clearly. Research on the structure of GLP mainly includes the range of molecular weight, monosaccharide composition, glycosidic bond types, repeating units, etc. The main methods for analyzing the monosaccharides composition of polysaccharides include GC^[124,187,189,193-196], LC^[194-195,197-200] and high-performance anion-exchange chromatography (HPAEC)^[201]. High-performance thin-layer chromatography (HPTLC)^[202], carbohydrate gel electrophoresis (PACE)^[203], and HPSEC-ELSD^[124,187,189,193,204] methods are also used to detect polysaccharides. After the sample be processed by acid hydrolysis, the glycosidic bond of the polysaccharide is cleaved, so that the monosaccharide composition can be conveniently detected by the GC and GC-MS method. In addition, GC-MS combined with methylation treatment can help determine the connection mode of sugar residues in polysaccharides, and periodic acid oxidation and Smith degradation can determine the type of glycosidic bonds^[205]. Furthermore, by derivatizing the sample with 1-phenyl-3-methyl-5-pyrazolone (PMP), the monosaccharide molecules are given ultraviolet or fluorescent absorbing groups to determine the composition of monosaccharides by HPLC^[206]. As one of the factors affecting the biological activity of polysaccharides, molecular weight is mainly detected by size exclusion chromatography (SEC) and gel permeation chromatography (GPC)^[187]. Some researchers have utilized matrix-assisted laser desorption/ionization (MALDI) mass spectrometry to test molecular weight^[207]. The connection mode of glycosidic bonds can be analyzed by infrared spectroscopy, and NMR technology can help analyze the anomeric configuration, the connection mode of glycosidic bonds, the connection sequence of sugar residues, etc. The different techniques for determining the relevant information of polysaccharides are listed in Table 16, and the different pre-treatment methods required are also listed.

The activity of Ganoderma polysaccharide has been reported many times, and Ganoderma polysaccharide has been included in the Chinese Pharmacopoeia as a quality control marker^[179]. Many researchers have studied the fingerprint of triterpenoids and polysaccharides from Ganoderma and judged the total content of the composite components at the same time, in order to distinguish Ganoderma species from different origins and different growth conditions. At present, the research on quality control of G. lucidum polysaccharide is mainly about the monosaccharide or oligosaccharide composition of polysaccharide hydrolysate by acid hydrolysis combined with instrument detection, based on which G. lucidum in different regions could be distinguished. For instance, Zhao et al.^[212] discriminated G. lucidum origins by HILIC-ELSD/ ESI-TOF/MS combined with acid hydrolysis, and polysaccharides and D-galactose could be considered as promising quality control markers. The fingerprinting method of HPTLC can distinguish the polysaccharides of G. applanatum and G. lucidum. However, for G. lucidum, G. nigrolucidum and different parts (fruit bodies and spores) of Ganoderma, the difference in their fingerprints are not very obvious, and the differences between the samples could hardly be detected^[202]. In addition to monosaccharide composition analysis, molecular weight and conformation also affect the activity

	I J		
Method	Sample processing	Structure information	Ref.
GC	Complete acid hydrolysis & Acetylation	Monosaccharide composition	[124,189,193-196,208]
(HP)LC	Pre-column derivatization with PMP Acid hydrolysis & Alkali treatment	Monosaccharide composition Purity and molecular weight	[194-195,197-200]
GC-MS	Methylation analysis & Acid hydrolysis	Glycosidic bond type	[124, 189, 192-194, 196, 198-200, 205, 208-211]
(HP)SEC	Dissolve in eluent	Molecular weight	[124,189,193,204,208]
(HP)GPC	Dissolve in water	Molecular weight	[198,205,207]
HPAEC-PAD	Acid hydrolysis	Monosaccharide composition	[201]
NMR	Dissolve in D ₂ O	Anomer configuration, glycosidic bond connection mode, sugar residue connection sequence	[124,189,193-196,200,207-208,211]
IR	KBr	Glycosidic bond type, characteristic group	[124,189,192-196,199-200,205,207-208,211]
AFM	Dissolve in water	Macromolecular conformation	[208]
HPTLC	Acid hydrolysis	Polysaccharides fingerprint	[202]
PACE	Partial acid hydrolysis & Enzymatic digestion	Polysaccharides fingerprint	[203]

 Table 16

 Analytical method of polysaccharides in *G. lucidum*.

of GLP. Li et al.^[132] found that the structural features of GLP are related to immunoregulatory activity. The polysaccharides degraded by ultrasound exhibited higher lipid-lowering and antioxidant activities than natural polysaccharides^[207]. In recent years, with the advancement of analytical technology, breakthroughs have also been made in data processing methods. Multivariate statistical analysis is an analytical method suitable for quality control of Chinese herbal medicines, including PCA, hierarchical cluster analysis and (orthogonal) projection-to-latent-structure discrimination analysis ((O)PLS-DA)^[212]. Among them, the PCA and OPLS-DA methods are the most popular in traditional Chinese medicine fingerprint. Both the PLS-DA and OPLS-DA methods each have their advantages and disadvantages so the selection method should be based on the specific data situation^[213].

At present, most of the studies have focused on the chemical composition alone or the efficacy of the polysaccharides. And no consensus has been reached on the structure-activity relationship through the research on GLP. Further, studies on the activity of GLP and the summary of its chemical structure similarities require the future efforts of researchers.

3. Bioactivity and its mechanism

3.1 Anti-tumor effect

Numerous laboratory researchers and preclinical trials have shown that *G. lucidum* has extensive anticancer activity. It was reported that *G. lucidum* could induce cell-cycle arrest and apoptosis^[214], inhibit angiogenesis, invasion and metastasis, and tumor growth to exert its anticancer activity *in vitro* and *in vivo*^[215-216]. And there are accumulating evidence implicating that *G. lucidum* also exerts anticancer action by affecting autophagy^[217-22], immune function^[223-229], chemotherapy, and radiotherapy sensitivity^[230-236].

G. lucidum extract, which contains 6% triterpenes and 13.5% polysaccharides, inhibited the proliferation of ovarian cancer HOCC cells by up-regulating Cx43 expression and down-regulating the expression of VEGF^[237]. GLP and enzymatically hydrolyzed *G. lucidum* polysaccharide (EGLP) induced apoptosis in U14 cells and suppressed tumor growth in U14 cervical tumor-bearing mice, in which the anticancer activity of EGLP is better than GLP. Moreover, EGLP exerted anticancer activity by regulating the apoptotic process,

decreasing the expression of Bcl-2 and COX-2, and increasing the expression of Bax and cleaved caspase- $3^{[238]}$. Anti-angiogenesis of *G. lucidum* showed as the viability of HUVEC or the formation of HUVEC capillary tubes was inhibited *in vitro*^[239-240], and decreased the expression of VEGF and bFGF or microvessel density of tumor *in vivo*^[241]. *G. lucidum* inhibited breast cancer migration and invasion by regulating the Rac/Lamellipodin pathway^[242] and suppressed breast-to-lung cancer metastases^[243].

The extracts of G. lucidum fruiting body (GLE) induced HCT116 cell cycle arrested in G0/G1 correlated tightly with the decreasing gene expression of E2F-1, CDK2, CDK4, CDK6, Cyclin A2, Cyclin B1, Cyclin E1 and increasing expression of P21^[222]. Apoptosis was induced after GLE treatment via decreasing the ratio of Bcl-2/Bax and increasing cleaved caspase-3 and poly ADP-ribose polymerase protein expression^[222]. An increase in HCT116 cell autophagy was observed with the treatment of GLE, which was induced by regulating the related protein expression in the mTOR pathway^[222]. Consistently, the antitumor effect of G. lucidum is regulated by caspase-dependent apoptosis^[221,244-246] and inducing initiation of autophagy^[217,221] was proved in other studies. Sporoderm-broken spores of G. lucidum water extract (BSGLWE) was studied to show that it disrupted cell cycle progression and induced apoptosis in colorectal cancer HCT116 cells^[244]. Moreover, BSGLWE suppressed tumor growth in vivo by regulating the expression of genes and proteins associated with the cell cycle and apoptosis (Fig. 3)^[244]. G. lucidum exerts its anticancer activity for inhibiting proliferation and inducing cell death via caspase-dependent and cyclin-CDK2 pathways and regulating mTORC1/2-mediated signaling pathways by activating AMPK and inhibiting IGFR/PI3K/Rheb (Fig. 3)^[247]. Water extract from G. lucidum also induced mitochondriamediated apoptosis, arrested the cell cycle at the S phase via the cyclin-CDK2 pathway, and inhibited cell migration associated with EMT in glioblastoma cells GBM8901 and U87 (Fig. 3)^[248]. G. lucidum spore oil (GLSO) induced apoptosis in vitro and in vivo via the caspase pathway (Fig. 3)^[249].

Autophagy affected by *G. lucidum in vitro* and *in vivo*, which was increased autophagosome accumulation and blocked autophagic flux^[217,221,250]. Moreover, autophagosome accumulation is responsible for apoptosis in colorectal cancer, which is mediated via MAPK/ERK activation^[217]. Ganoderic acid D, a representative active triterpenoid from *G. lucidum*, would inhibit cell proliferation, cycle arrested,



Fig. 3 Anti-tumor mechanisms of G. lucidum.

induce apoptosis and autophagic cell death to exert its anticancer effect, which through the mTOR pathway to lead the synergistic effect on apoptosis and autophagic cell death in the esophageal squamous cell carcinoma (ESCC) cells^[250]. However, cell cycle arrest for antitumor activity of *G. lucidum* not only arrested in G0/G1 phase^[218-219,222,247] but also arrested in the G2/M phase^[221,244,250], S phase^[248] and subG1 phase^[245]. GLP conjugated with bismuth sulfide nanoparticles (GLP-BiNP) increased the sensitivity of radiotherapy to inhibit invasion, metastasis, and tumor growth, and alter the radiation-induced immunosuppression microenvironment^[235]. Moreover, hepatic carcinoma HepG2 cells could be suppressed by GLP, which regulates the expression of hepatic miRNAs and immune-related miRNAs^[226].

3.2 Anti-microbial activity

3.2.1 Antibacterial activity

Fungi are a widespread kind of microorganism. As a notable kingdom of fungi, mushrooms can be edible and play an indispensable role in alternative and supplementary medicine. Owing to a set of bioactive functions, *G. lucidum* is of significant ability against microbiota, including bacteria and fungi. The capacity of *G. lucidum* against bacteria is characterized by the inhibition of both Gram-positive bacteria and Gram-negative bacteria^[184]. Interestingly, reports facilitated by different extracts of *G. lucidum* commonly indicated anti-bacteria ability, which enhanced the practicability of *G. lucidum* but puzzled determination in defined bioactive compounds.

Extract parameter, yield region, and chosen organ of *G. lucidum* determine the resultant content of bioactive compounds and direct efficacy of anti-bacteria ability. Scientists have used water, methanol, dichloromethane^[251], acetone^[252], and chloroform as solutions to obtain a prepared extract of *G. lucidum*. An interesting design, in which dried *G. lucidum* powder was added to beef sausage, demonstrated the anti-bacteria ability of raw *G. lucidum* was similar

to nitrite^[253]. With a spectrum of components in different categories, *G. lucidum* showed to protect from oxidant stress, while its anti-bacteria ability is closely associated with the generation of reactive oxidant species oppositely. In addition, the leakage of bacterial inner proteins is another mechanism of the phenolic-rich fraction of *G. lucidum* water extract^[254]. Another report compared the antibacterial ability of per ether, chloroform, and methanol extract. Methanol extract exhibited the smallest IC₅₀ value implying variable anti-bacterial efficacy attributed to extraction conditions^[255]. Gokcen^[256] considered phenolic compound, β -carotene and lycopene components as significantly bioactive components of methanol extract.

When the discrimination in species was considered, the sensitivity of different tested bacteria to *G. lucidum* varied. To detect whether *G. lucidum* acted on drug resistance, multidrug-resistant tuberculosis was considered. After the administration of *G. lucidum* extracts in different concentrations, the growth of tuberculosis was inhibited^[257]. In addition, extended-spectrum β -lactamase-producing and multidrugresistant *Pseudomonas aeruginosa* was examined for supporting the specific antibacterial ability of *G. lucidum*^[13]. Several studies also established test models using an array of bacteria, which might provide a detailed description of *G. lucidum*. *G. lucidum* could suppress either special species, like drug-resistant ones or common species extract to widen its anti-bacteria applications. General lines, including *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus epidermis*, *Enterococcus raffinosus* were inhibited by *G. lucidum*, respectively^[258].

There remained several reports about mono-components for anti-bacterial ability despite raw extract of *G. lucidum*. Peptides of *G. lucidum* obtained from fruiting bodies and mycelium showed anti-bacterial potentially. The structural presence of cationic and hydrophobic amino acids facilitated its efficacy^[259]. This effect also appeared in another research focused on crude proteins of *G. lucidum*^[260]. *P*-hydroxybenzoic and cinnamic acids found in *G. lucidum* *G. lucidum*^[261]. Richly produced in the fruiting body of *G. lucidum*, polysaccharides, especially with non-reducing sugars, exhibited anti-bacterial performance directed to foodstuffs and human health-associated microorganisms^[262-263], while exopolysaccharide and chitosan did as well^[264-265]. These proofs congruously indicated the anti-bacterial ability of *G. lucidum*. However, fewer studies concentrated on the affirmatory bio-compounds with mechanisms of *G. lucidum* limited its applications. With anticipated extraction and separation methods improving, the effect and mechanisms of *G. lucidum* anti-bacterial will be further studied when the purified components can be prepared.

3.2.2 Anti-fungal activity

Although bacteria and fungi are different in structures and characters, both microorganisms are negatively regulated by G. lucidum. Research has wildly examined the suppressive effects of G. lucidum on an array of pathogenic fungi, including Candida albicans, Candida krusei, Candida glabrata^[251], and Aspergillus fumigatus^[261]. The anti-fungi ability of G. lucidum attracted growing interest in this natural substance rather than standard agents resisted by vicious fungi. Dejan S. compared the efficacy of G. lucidum from China with that from Serbia. The variable sources resulted in different sensitivity of tested fungi to the administration of G. lucidum, which performed better than bifonazole and ketoconazole. When treated by that from China, Trichoderma viride and Penicillium funiculosum were best suppressed while extract of G. lucidum from Serbia sensitively impacted Aspergillus versicolor^[266]. The hypothesis that diverse strains brought out different anti-fungi efficacy was also proved by Swati's discovery^[267]. Two strains of G. lucidum, DARL-4, and MS-1, showed moderate anti-fungi ability compared to fluconazole. Monica measured the performances of proteins of G. lucidum extract acting as deoxyribonuclease, ribonuclease, protease, glucanase, and chitinase. The anti-fungi efficacy of G. lucidum might partially associate with proteins of G. lucidum for the degradation of nutrient components that were necessary for phytopathogen fungi[268].

3.2.3 Anti-viral efficacy

People hitherto have found a variety of viruses and driven away at the development of broad-spectrum anti-viral agents. In addition to the frequent mutation of viruses, drug resistance is also a vital problem in clinical practice. Some active components isolated from fungi show potent pharmacological action in treating infectious diseases. G. lucidum has been proven with specific efficacy in treating various internal diseases for thousands of years, and its efficacy in anti-virus gradually comes into notice. Based on the development of modern pharmacology, several studies isolated and purified many active components from G. lucidum, and part of them have been verified with efficacy in anti-virus in vivo or in vitro. Some early studies show that ganoderic acid could inhibit the excretion of HBsAg and HBeAg in a dose-dependent manner and protect liver tissues from injury^[269-270]. Otherwise, the proteoglycans of G. lucidum prohibited herpes virus-associated pain was proved in vitro^[271] and in a clinical study on 5 patients in Japan^[272]. But the anti-hepatitis virus and antiherpes virus molecular mechanisms induced by G. lucidum still lack molecular level studies.

Dengue is prevalent in several countries and regions, and the symptoms are characterized by hyperpyrexia, fatigue, and bleeding. The replication and assembly of DENV viruses mainly depend on NS2B-NS3 protease (PR) activation, which becomes a potential target for treatment. By comparison with an inhibitor of NS2B-NS3 PR, four triterpenoids isolated from *G. lucidum* are probable inhibitors of NS2B-NS3 protease. In a consequential study *in vitro*, Ganodermanontriol was further put forward as a potent inhibitor^[273], but the interaction between Ganodermanontriol and DENV NS2B-NS3 PR is elusive. Six active components were isolated from another form of *G. lucidum* in another study that showed better inhibitory efficacy on DENV-2 NS2B-NS3 PL at $(84.6 \pm 0.7)\%^{[274]}$. Additionally, the molecular interactions between these components and DENV-2 NS2B-NS3 PL have been confirmed as van der Waals, hydrogen bonding, and pi-pi interactions.

Epstein-Barr virus (EBV) is a human herpes virus that exists as a DNA loop in the B lymphocyte. Infection of EBV contributed to about 1.5% of human cancer patients and is expressed in malignant tumors^[275], in which the treatment of anti-EBV has become a preventative strategy. Co-treatment of G. lucidum extracts and quercetin in a low concentration suppressed the development of EBV-associated gastric cancer and the increased PAPR1 cleavage, caspase 3 and decreased Bcl-2 were observed, which proved the G. lucidum extracts enhanced the quercetin induced apoptosis in a dose-dependent manner. Mechanically, G. lucidum extracts and quercetin upregulated the expression of EBV latent and lytic genes, triggering lytic reaction, and consequently leading to the apoptosis of EBV^[276]. Zheng et al.^[277] isolated 5 active components, including ganoderic acid A, ganoderic acid B, ganoderol B, ganodermanontriol, and ganodermanondiol from G. lucidum by dichloromethane, all of them inhibited the activity of telomerase and EBV capsid antigens to the almost same degree. But the ganoderic acid A and B exhibited more effectiveness than the other three in inhibiting EBV early antigens. Furthermore, the hydrogen bonds, van der Waals force, and hydrophobic interaction were confirmed in the combination of ganoderic acid A and amino acid residues.

In a clinical study containing 61 patients with gingivitis and positive for human papillomavirus 16 serotype (HPV16) or human papillomavirus 18 serotypes (HPV18), the group with combination therapy of G. lucidum and Trametes versicolor performed better than that with Laetiporus sulphureus only^[278], which indicated the potential anti-HPV efficacy of G. lucidum. Another study further confirmed the pharmacological effects of G. lucidum extracts in inhibiting the proliferation, apoptosis, and cell cycle of HPV transformed cells^[279]. Zhang et al.^[280] discovered G. lucidum triterpenoids (lanosta-7,9(11),24-trien-3-one,15;26-dihydroxy) and ganoderic acid Y exhibited anti-enterovirus 71(EV71) effects in vitro. This efficacy probably depended on the binding of active components and viral capsid proteins, associated with the adhesion of the EV71 to the cell membrane. In addition to the inhibition of adsorption of EV71, these two components also interfered with the replication of EV71 RNA. The G. lucidum induced immunomodulation is also related to the polysaccharide, which interacts with the innate immune receptors, including Dectin-1, DC-SIGN, Langerin, and TLR2^[281]. The relatively broad-spectrum anti-viral efficacy of active components in G. lucidum mostly depends on the interruption of adsorption, invasion, and cell cycle. Some studies also illustrated the molecular mechanisms in this progress, at least in part.

3.3 Anti-HIV protease efficacy

Although various anti-viral efficacy of *G. lucidum* has been confirmed in the past decade, the development of anti-human immunodeficiency virus (HIV) drugs is still a complicated issue^[282]. It has been found that the ganoderic acid β , lucidumol B, and ganolucidic acid A isolated from *G. lucidum* exhibited dramatic inhibition effects on HIV-1 protease activity as early as the end of the 20th century^[66], but the molecular mechanisms of these effects are still unclear.

The HIV-1 protease (PR) is associated with the proliferation of HIV-1, which cleaves precursor protein and promotes HIV maturity, and is targeted by some active components in G. lucidum. Five active components isolated from the fruiting bodies and spores of G. lucidum, namely ganoderic acid β , lucidumol B, ganodermanondiol, ganodermanontriol, and ganolucidic acid A, had potent anti-HIV1 PR activity in a value of IC_{50} within 20–90 μ mol/L^[66]. Another study proved extractive ganoderiol F inhibited HIV at a concentration of 7.8 mg/mL, and other active components such as ganoderiol B and ganoderic acid C1 also inhibited HIV PR moderately^[41]. In a molecular docking study on the interaction between ganoderic acid with 1HVR and 1DIF, which are 2 types of HIV-1 PR, ganoderic acid B performed the lowest cluster number and IC₅₀ value. Triterpenes in G. lucidum significantly inhibited α -glucosidase and α -amylase activity with IC₅₀ value of 0.1 µmol/L. Furthermore, the model of ganoderic acid B formed 4 hydrogen bonds with 1HVR in a manner of ILE50, ILE50, ASP29, and ASP30 residues^[283]. HIV-1 reverse transcriptase (RT) is another target for pharmacological inhibitor extract from G. lucidum. A laccase isolated from fresh fruiting bodies of G. lucidum is characterized by novel N-terminal, high molecular weight, and Con A-Sepharose adsorption only. Its anti-HIV RT efficacy is probably based on the interaction between protein-protein^[284].

The acquired immunodeficiency syndrome (AIDS) caused by HIV is still a serious illness in the world and inhibition of HIV PR and HIV RT are principal strategies in existing therapies. Whereas the side-effects elicited by tipranavir, saquinavir and ritonavir have become a new problem for AIDS treatments^[285] and complementary and alternative medicine therapies could provide more selectable solutions. Studies proved some active components isolated from *G. lucidum* have anti-HIV efficacy, but the molecular mechanisms of these effects are still unclear.

3.4 The regulation of G. lucidum in diabetes mellitus

Bioactive components isolated from *G. lucidum* exhibit potent anti-diabetic efficacy via various pharmacologic actions, but the protection of renal insufficiency is needed to study further. According to the International Diabetes Federation database, the burden of global diabetes mellitus (DM) has risen to 463 million adult patients in 2019. Although insulin injection and oral glucose-lowering drugs, including Metformin, Acarbose, DDP-4 inhibitors, and SGLT-2 inhibitors, are recommended as the first-line therapies, diet and regular exercises are still needed and undesirable side effects are inevitable^[286]. Most make us anticipate the exploration of more effective and glucose-lowering drugs and permanent remedies, which are ongoing and extracts isolated from plants play a vital role in the study of DM treatment^[287].

In addition to the pharmacological effects, the extraction process influences the efficacy of *G. lucidum*. In the condition of 65.8–70.0 °C for 2.8–3 h, the bioactive components acquired optimal anti-diabetes activity and more than 39% α -glucosidase inhibition effects^[288].

In a study with T2DM rats induced by streptozotocin, GLPs treatment dramatically reduced fasting blood glucose (FBG) and insulin, mediated the aberrant gut microbiota and enhanced the antioxidant ability^[289]. GLPs also involved in regulating lipid metabolism in db/db mice. In the high-fat diet db/db mice treated with GLPs, the body weight, FBG, and HbA1C levels decreased and aberrant lipid metabolism was corrected and consequently inhibited lipid anabolism^[290]. Triterpenes in G. lucidum significantly inhibited α -glucosidase and α -amylase activity with IC₅₀ values of (10.02 ± 0.95) and $(31.82 \pm 4.30) \ \mu g/mL$, respectively^[291], which indicated the triterpenes are promising to act as a substitution of acarbose and voglibose. Furthermore, the triterpenoids isolated from G. lucidum improve glucose consumption in insulin resistance cells. The concentrations of triterpenoids at 0.03 and 0.06 mg/mL contributed to glucose consumption values at (1.80 ± 0.12) and (2.21 ± 0.29) mmol/L, respectively, and without cytotoxicity in HepG2 cells^[292].

Impairment of the pancreas organ is a major feature of DM, especially in T1DM, which can be observed in the early years of the patients. The impaired islet β cells trigger the decreased secretion of insulin and consequently result in hyperglycemia, so restoring the pancreas function and regulating blood glucose are two principal strategies in treating DM. Protection of pancreas organ induced by recombinant LZ-8, an analog of immunomodulatory protein Ling Zhi-8, promotes insulin excretion, relieves symptoms of T1DM, decreases blood glucose and HbA1c and inhibits TNF- α and IL-1 β , which is mainly depended on its anti-inflammation and regulatory T cells mediation^[293]. A proteoglycan isolated from G. lucidum was named FYGL and had similar efficacy in inhibiting pancreatic β cells injury accompanying decreased ROS and NO levels^[294]. In an early study, researchers illustrated the mechanisms of insulin resistance restoration induced by FYGL. Activation of protein tyrosine phosphatase 1 B(PTP1B) dephosphorylated insulin receptor substrate (IRS) contributed to the dysfunction of the insulin signal pathway. This aberrant phenomenon was corrected by FYGL, accompanied by activation of PI3K/Akt, and finally restored the glucogen synthesis by insulin in HepG2 cells^[295]. Furthermore, treating myoblast L6 cells with FYGL also led to the activation of AMPK and increased the expression of GLUT4, promoting glucose uptake^[16].

A novel complex GLP-chromium (III) (GLP-Cr III) treated pre-diabetic mice with administration of 50 mg/kg per day decreased the level of FBG, total cholesterol (TC) and triglyceride (TG)B^[296], which provided a novel form of GLP. The vasculopathy is still a frequent pathologic change in DM patients with the long course, which results in increased risk factors of cardiovascular disease, renal insufficiency, and fundus lesions. In terms of these challenges, a clinical study confirmed that *G. lucidum* administration could not increase cardiovascular risk^[297]. *G. lucidum* mycelia mediated the leukocyte metabolism and oxidative stress in DM rats^[298]. The ROS and chronic inflammation induced by advanced glycation end products (AGE) are associated with

hyperglycemia and finally result in vasa vasorum angiogenesis, but this pathologic process can be interfered with by polysaccharide peptide (PSP) isolated from *G. lucidum*^[299]. PSP in *G. lucidum* also promotes the repair of blood vessels. A study with 35 Wistar rats showed that the PSP promoted endothelial repairment and downregulated the risk factors in an optimum dose with 300 mg/kg body weight, which was proved by the alteration of endothelial progenitor cells and circulating endothelial cells level^[300]. Xiao et al.^[301] confirmed an anti-diabetic bioactive component F31 in GLP. Mechanistically, F31 promoted the phosphorylation of AMPK and decreased the level of hepatic glucose regulatory enzyme mRNA in liver tissues. The increased GLUT4 was also observed in the T2DM mice and was accompanied by decreased epididymal fat/ body weight.

3.5 Liver and gastric injury

Protection from liver damage was another application of G. lucidum. There remained a set of articles using in vivo and in vitro models to assess how G. lucidum possessed anti-hepatitis ability^[151,302]. G. lucidum could treat acute liver damage induced by carbon tetrachloride^[303]. Shi^[304] and Han^[305] also discovered that treatment of G. lucidum ameliorated the insult of D-galactosamine and lipopolysaccharide in hepatitis mice. With good ability in radical scavenging, mitochondrial enzyme normalization, and membrane potential restoration, G. lucidum protects from disruption of liver injury-associated indications followed by attenuated MDA, superoxide dismutase (SOD), and glutathione (GSH)^[306]. As disruption of aminotransferase, aspartate aminotransferase, and lactate dehydrogenase were assessed after treatment of G. lucidum in preclinic^[307-308], a randomized controlled trial using a commercial drug mainly consisting of G. lucidum gave support for the translational medical potentiality of G. lucidum in liver protection. More attention was then put on triterpenoids^[309-310] and GLP^[311] because of structural identification and detectable antioxidant profile compared with raw extract of G. lucidum with water, ethanol, or methanol^[21].

Like the role in hepatocellular carcinoma, the *G. lucidum* and its extracts also can contribute to reversing the gastroblastoma, which may interfere with intracellular autophagy. Methanolic fruiting body extract of *G. lucidum* inhibited the growth of gastric cancer cells by prohibiting the cellular autophagy and cell cycle by increasing the LC-II and p62 adaptor^[220,312]. Meanwhile, *G. lucidum* and its extracts regulate the fungal immunomodulatory protein Lz-8 and induce endoplasmic reticulum (ER) stress-mediated autophagic cell death in the human gastric cancer cell line SGC-7901^[313]. Further investigations need to elucidate whether those autophagosome-lysosome procedure and mediate aiming protein degradation.

3.6. Cardiovascular potential

As a multi-pathway regulation, *G. lucidum* works on the cardioprotective effects^[314]. Selenium-enriched GLP (Se-GLP) was discovered to prevent oxidative damage in a mouse model of heart reperfusion injury and made the role of anti-oxidative regent. In such a model, the antioxidant enzymes, including SOD, catalase (CAT),

glutathione peroxidase (GSH-Px), and GSH were all compromised, which also contributed to reversing the heart injury-induced failure^[315]. Accordingly, Ganoderic acid A extracted from *G. lucidum* was correlated to a hypoxic injury of the heart going through the PI3K/AKT pathway mediated rat H92 cardiomyocyte proliferation and apoptosis attenuation^[316]. Furthermore, in the pressure-boosting irradiated cardiomyopathy mice model, one extract of spore oil was confirmed for the modification of cardiac function improvement through the circle RNA-FOXO3 axis, which is an important pathway associated with heart failure^[317].

G. lucidum and its extraction ingredients were not only directly used in myocardial disorders, but also effective in the following complications^[318-320]. Interestingly, G. lucidum and its ingredients can alleviate cardiovascular collapse and damage caused by diastolic dysfunction^[321]. At the same time, polysaccharide peptide extracted from G. lucidum was reported to hinder diabetes and reduce the cardiac disease risk by releasing the vascular damage level in Wistar rat models^[300]. Besides, G. lucidum spores declined the total cholesterol and triglycerides in diabetic rats through upregulating the acyl-CoA oxidase 1, acetyl-CoA carboxylase, INSIG1, and INSIG2 gene expression^[322]. Another major complication of cardiovascular disease is dyslipidemia, for instance, polysaccharide peptide was applied to protect the atherogenesis process in the context of dyslipidemia from atherosclerosis and blood vessel damage^[323]. Polysaccharides as hypolipidemic ingredients can also exhibit antioxidant and antiapoptotic effects in high-fat diet mice^[324].

4. Preclinical and clinical studies

Before the common era, *G. lucidum* has been widely used by Asia populations to improve general health. In recent years, novel applications of *G. lucidum* have been suggested, such as treatment alongside Western chemotherapy to inhibit cancer and reduce side effects. In addition, other diseases, such as brain, renal, immune, infection, and myopathy-related problems, have also been regulated by *G. lucidum*.

4.1 Protective effects of G. lucidum against cancer

G. lucidum was used as adjuvant therapy with chemotherapy or radiotherapy in cancer treatment to prolong long-term survival, promote life quality, and regulate immunity. To evaluate the effects of G. lucidum in cancer treatment, clinical studies since 1997 have been published^[325-329]. Alteration of the immune system after administration of G. lucidum is discussed most. Evidence revealed that cancer patients displayed a series of cellular immunological enhancements, such as NK cell activity and CD4/CD8 ratios were changed after Lingzhi capsule, G. lucidum extracts or Ganoderma spore powder supplementation^[325,330-332]. The quality of life is also evaluated after the administration of G. lucidum. Studies show that the administration of G. lucidum extracts improves the quality of life of patients with lung cancer^[333]. After oral administration of *Ganoderma* spore powder capsules, one hundred patients with digestive system tumors obtained more remarkable points of quality of life score than the control^[334]. In addition, oral administration with Ganoderma spore powder can also improve physical well-being and fatigue subscale for breast cancer patients undergoing endocrine therapy^[335]. Although accumulative clinical studies about *G. lucidum* in cancer therapy emerged, there is no sufficient evidence to encourage the use of *G. lucidum* as a first-line treatment for cancer. Also, long-term treatment has not been followed, and it remains uncertain whether the treatment can help prolong cancer patient survival. However, *G. lucidum* is an alternative adjunct that potentially enhances immune response, promotes quality of life, and shows minor adverse effects for patients with cancer.

4.2 Renal protective effects of G. lucidum

Shieh et al.^[336] first observed the protective function of *G. lucidum* on renal cells and hepatocytes. Subsequently, several clinical trials have been started to evaluate the effects. Futrakul et al.^[337] found that 5 patients with impaired renal function showed significantly decreased proteinuria after taking a crude extract of fungus *G. lucidum* for 1 year. In 2003, Xiao et al.^[338] observed that *G. lucidum* decoction is useful in markedly lower *Russula Subnigricans*-induced kidney injury in patients. In addition, Nephrotic patients with focal segmental glomerulosclerosis also ameliorated injured kidneys after administration of *G. lucidum*, such as inhibiting endothelial cell cytotoxicity, restoring immune-circulatory balance, and suppressing proteinuria^[339]. These clinical results confirmed the potential protective roles of *G. lucidum* in renal health and proved the safety of administration of *G. lucidum* in the human body.

4.3 Neuroprotective effects of G. lucidum

G. lucidum is neuroprotective in ischemia/reperfusion or traumatic spinal cord injury in experimental models, which drives clinical trials to determine its effects on neuroprotection. G. lucidum has been reported to show effects on pain relief. Administration of hot water extracts from various herbs, including G. lucidum has been tested to relieve herpes zoster pain within a few days. No patients developed post-herpetic neuralgia after more than one year of follow-up^[340]. In 2018, a clinical study reported that weekly seizure frequency was reduced in patients with epilepsy after administering G. lucidum spore powder^[341]. In addition, G. lucidum also shows curative effects on depression in combination therapy^[342]. However, the effects of G. lucidum spore powder on Alzheimer's disease are limited. G. lucidum spore powder treatment for 6 weeks did not show more encouraging results in symptom improvements^[343]. Taken together, the evaluation of the neuroprotective effects of G. lucidum is limited in clinical trials.

4.4 Protective effects of G. lucidum on other diseases

Considering the potential immune regulation effects of *G. lucidum*, immune status has been checked in rheumatoid arthritis (RA) patients and hemophiliacs with positive HIV antibodies after administration of *G. lucidum*^[344-345]. RA patients' global score improved significantly, but no obvious alteration in immune status for hemophiliacs after eating *G. lucidum*. In addition, Ji 731 injection, which contained *G. lucidum*, was evaluated to be useful for atrophic rhinitis^[346].

In conclusion, G. lucidum or its related products were potentially effective for several diseases, especially for immune regulation after

chemotherapy for cancer patients. Moreover, immune regulation has also been applied to other immune or inflammation-related diseases, such as RA or atrophic rhinitis. Not only immune regulation, but neuron system-related diseases were also possible to be modulated by *G. lucidum*, such as pain relief and neurodegenerative diseases. However, as the potential mechanism of *G. lucidum* in biological function and there is no long-term treatment trial, the effects of *G. lucidum* is hard to observe and evaluate.

5. Safety, toxicity, and side effects evaluation

G. lucidum includes triterpenes, polysaccharides, nucleosides, steroids, fatty acids, alkaloids, proteins, peptides, amino acids, and inorganic elements^[347]. Most reports illustrated that one or some of them are not causing apparent toxicity or safety problems. Rodent animals, for example, can tolerate up to 5 000 mg/kg dose of G. lucidum administration, and no mortality rate was reported during the integral progression^[348]. In addition, extracted ingredients from GLP were administrated in Wistar rats compared with a placebo. The result indicated no significant differences in abnormal clinical symptoms as well as body weight and food intake, even the same for the hematology and clinical chemistry values^[349]. This phenotype matched the data in multiple human clinical trials. Health volunteers 1.5 g/day and 4 consecutive weeks of exposure would not affect their hemostatic parameters as well as platelet and global hemostatic functions, while it cannot cause breeding or tissue damage problems^[350]. After a 12-week stable dose of G. lucidum treatment, 23 dyslipidemia volunteers accompanied mild hypertension that did not affect the physiological expression, only found a few side symptoms considered not clinically significant such as headache, influenza/running nose^[17]. However, minors reported higher doses of it to induce some hematologic abnormal and immune responses. For example, atherosclerotic patients who receive the above amount of G. lucidum treatment with 3 000 mg/day can inhibit their platelet aggregation progression^[351]. After a 10-day trial, participants supplemented with 2 g of G. lucidum were found to boost CD56, the cell surface glycoprotein involved in embryonic development and nerve growth in their blood samples^[352]. Besides, GLP can enhance the sheep's red blood cells though no phagocytic function or macrophages were manifested during the procedure^[349]. Overall, appropriate doses of G.</sup> lucidum under expert advice may not trigger toxicity, side effects, and safety problems for the patients.

6. Conclusion

G. lucidum, as a medicinal and edible plant, has been used for a long history. After years of research, more than 300 compounds have been isolated from the fruit body, mycelia and spore of *G. lucidum*. These ingredients have also been shown to have various pharmacological effects. One of the most studied is tumor and immune regulation. *G. lucidum* has also been developed into various products, of which *G. lucidum* spore-related functional food is the most famous. In these years of the outbreak of COVID-19, *G. lucidum* has also been studied on COVID-19, showing a good effect in inhibiting the COVID-19 virus. Despite the wide application of *G. lucidum*, the therapeutic mechanism of *G. lucidum* for various diseases is still unclear. Long-term follow-up is also lacking in clinical studies. As a popular medicinal and edible plant, *G. lucidum* needs more research on its therapeutic mechanism and clinical research.

Declaration of competing interests

The authors declare that they have no competing interests.

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References

- A. González, V. Atienza, A. Montoro, et al., Use of *Ganoderma lucidum* (Ganodermataceae, Basidiomycota) as radioprotector, Nutrients 12 (2020) 1143. https://doi.org/10.3390/nu12041143.
- [2] Chinese Pharmacopoeia Commission, Chinese Pharmacopoeia, China Medical Science Press, 2020.
- [3] J. Simonić, M. Stajic, J. Vukojevic, *Ganoderma lucidum* from tradition to modern medicine, Zb. Matice. Srp. Prir. Nauke. 133 (2017) 151-161. https:// doi.org/10.2298/ZMSPN1733151C.
- [4] J. Li, J. Zhang, H. Chen, et al., Complete mitochondrial genome of the medicinal mushroom *Ganoderma lucidum*, PLoS One 8 (2013) e72038. https://doi.org/10.1371/journal.pone.0072038.
- [5] F.Y. Sheng, S.S. Wang, X. Luo, et al., Simultaneous determination of ten nucleosides and bases in *Ganoderma* by micellar electrokinetic chromatography, Food Sci. Hum. Wellness 11 (2022) 263-268. https://doi. org/10.1016/j.fshw.2021.11.015.
- [6] C.Q. Li, Y.P. Cui, J. Lu, et al., Ionic liquid-based ultrasonic-assisted extraction coupled with HPLC and artificial neural network analysis for *Ganoderma lucidum*, Molecules 25 (2020) 1309. https://doi.org/10.3390/ molecules25061309.
- [7] R. Zhao, Q. Chen, Y.M. He, The effect of *Ganoderma lucidum* extract on immunological function and identify its anti-tumor immunostimulatory activity based on the biological network, Sci. Rep. 8 (2018) 12680. https:// doi.org/10.1038/s41598-018-30881-0.
- [8] Y. Fu, L. Shi, K. Ding, Structure elucidation and anti-tumor activity in vivo of a polysaccharide from spores of *Ganoderma lucidum* (Fr.) Karst, Int. J. Biol. Macromol. 141 (2019) 693-699. https://doi.org/10.1016/ j.ijbiomac.2019.09.046.
- [9] Z.H. Yin, Z.H. Liang, C.Q. Li, et al., Immunomodulatory effects of polysaccharides from edible fungus: a review, Food Sci. Hum. Wellness 10 (2021) 393-400. https://doi.org/10.1016/j.fshw.2021.04.001.
- [10] C.Q. Li, Y.P. Cui, J. Lu, et al., Spectrum-effect relationship of immunologic activity of *Ganoderma lucidum* by UPLC-MS/MS and component knockout method, Food Sci. Hum. Wellness 10 (2021) 278-288. https://doi. org/10.1016/j.fshw.2021.02.019.
- [11] C. Dai, L. He, B. Ma, et al., Facile nanolization strategy for therapeutic *Ganoderma lucidum* spore oil to achieve enhanced protection against radiation-induced heart disease, Small 15 (2019) e1902642. https://doi. org/10.1002/smll.201902642.
- [12] C. Wang, X. Liu, C. Lian, et al., Triterpenes and aromatic meroterpenoids with antioxidant activity and neuroprotective effects from *Ganoderma lucidum*, Molecules 24 (2019) 4353. https://doi.org/10.3390/molecules24234353.
- [13] N.A. El-Zawawy, S.S. Ali, Anti-proteolytic activity of *Ganoderma lucidum* methanol extract against *Pseudomonas aeruginosa*, J. Infect. Dev. Ctries. 10 (2016) 1020-1024. https://doi.org/10.3855/jidc.6929.
- [14] B. Ergun, Evaluation of antimicrobial, cytotoxic and genotoxic activities of *Ganoderma lucidum* (Reishi mushroom), Pak. J. Pharm. Sci. 30 (2017) 1991-1995. https://doi.org/10.4172/2155-9600.C1.032.

- [15] H.T. Ma, J.F. Hsieh, S.T. Chen, Anti-diabetic effects of Ganoderma lucidum, Phytochemistry 114 (2015) 109-113. https://doi.org/10.1016/ j.phytochem.2015.02.017.
- [16] Z. Yang, F. Wu, Y. He, et al., A novel PTP1B inhibitor extracted from *Ganoderma lucidum* ameliorates insulin resistance by regulating IRS1-GLUT4 cascades in the insulin signaling pathway, Food Funct. 9 (2018) 397-406. https://doi.org/10.1039/c7fo01489a.
- [17] T.T. Chu, I.F. Benzie, C.W. Lam, et al., Study of potential cardioprotective effects of *Ganoderma lucidum* (Lingzhi): results of a controlled human intervention trial, Br. J. Nutr. 107 (2012) 1017-1027. https://doi.org/10.1017/ s0007114511003795.
- [18] Y.L. Wu, F. Han, S.S. Luan, et al., Triterpenoids from *Ganoderma lucidum* and their potential anti-inflammatory effects, J. Agric. Food Chem. 67 (2019) 5147-5158. https://doi.org/10.1021/acs.jafc.9b01195.
- [19] L.R. Wen, Z.L. Sheng, J.P. Wang, et al., Structure of water-soluble polysaccharides in spore of *Ganoderma lucidum* and their anti-inflammatory activity, Food Chem. 373 (2022) 131374. https://doi.org/10.1016/ j.foodchem.2021.131374.
- [20] B. Zhang, R.W. Zhang, X.Q. Yin, et al., Inhibitory activities of some traditional Chinese herbs against testosterone 5α-reductase and effects of *Cacumen platycladi* on hair re-growth in testosterone-treated mice, J. Ethnopharmacol. 177 (2016) 1-9. https://doi.org/10.1016/j.jep.2015.11.012.
- [21] B. Lakshmi, T.A. Ajith, N. Jose, et al., Antimutagenic activity of methanolic extract of *Ganoderma lucidum* and its effect on hepatic damage caused by benzo[a]pyrene, J. Ethnopharmacol. 107 (2006) 297-303. https://doi. org/10.1016/j.jep.2006.03.027.
- [22] Z. Lin, B. Yang, Ganoderma and health: biology, chemistry and industry, Springer Nature, 2019.
- [23] C.W. Huie, X. Di, Chromatographic and electrophoretic methods for Lingzhi pharmacologically active components, J. Chromatogr. B Analyt. Technol. Biomed. Life Sci. 812 (2004) 241-257. https://doi.org/10.1016/ j.jchromb.2004.08.038.
- [24] C. Liang, D. Tian, Y. Liu, et al., Review of the molecular mechanisms of *Ganoderma lucidum* triterpenoids: ganoderic acids A, C2, D, F, DM, X and Y, Eur. J. Med. Chem. 174 (2019) 130-141. https://doi.org/10.1016/ j.ejmech.2019.04.039.
- [25] C. Zhao, C. Zhang, Z. Xing, et al., Pharmacological effects of natural Ganoderma and its extracts on neurological diseases: a comprehensive review, Int. J. Biol. Macromol. 121 (2019) 1160-1178. https://doi. org/10.1016/j.ijbiomac.2018.10.076.
- [26] L. Gu, Y. Zheng, D. Lian, et al., Production of triterpenoids from *Ganoderma lucidum*: elicitation strategy and signal transduction, Process Biochem. 69 (2018) 22-32. https://doi.org/10.1016/j.procbio.2018.03.019.
- [27] S. Baby, A.J. Johnson, B. Govindan, Secondary metabolites from Ganoderma, Phytochemistry 114 (2015) 66-101. https://doi.org/10.1016/ j.phytochem.2015.03.010.
- [28] Q. Xia, H. Zhang, X. Sun, et al., A comprehensive review of the structure elucidation and biological activity of triterpenoids from *Ganoderma* spp., Molecules 19 (2014) 17478-17535. https://doi.org/10.3390/ molecules191117478.
- [29] S. Fatmawati, K. Shimizu, R. Kondo, Ganoderic acid Df, a new triterpenoid with aldose reductase inhibitory activity from the fruiting body of *Ganoderma lucidum*, Fitoterapia 81 (2010) 1033-1036. https://doi. org/10.1016/j.fitote.2010.06.025.
- [30] J. Liu, K. Shimizu, A. Tanaka, et al., Target proteins of ganoderic acid DM provides clues to various pharmacological mechanisms, Sci. Rep. 2 (2012) 905. https://doi.org/10.1038/srep00905.
- [31] B. Chen, J. Tian, J. Zhang, et al., Triterpenes and meroterpenes from *Ganoderma lucidum* with inhibitory activity against HMGs reductase, aldose reductase and α-glucosidase, Fitoterapia 120 (2017) 6-16. https://doi. org/10.1016/j.fitote.2017.05.005.
- [32] S. Fatmawati, R. Kondo, K. Shimizu, Structure-activity relationships of lanostane-type triterpenoids from *Ganoderma* lingzhi as α-glucosidase inhibitors, Bioorg. Med. Chem. Lett. 23 (2013) 5900-5903. https://doi. org/10.1016/j.bmcl.2013.08.084.
- [33] C.R. Cheng, Q.X. Yue, Z.Y. Wu, et al., Cytotoxic triterpenoids from *Ganoderma lucidum*, Phytochemistry 71 (2010) 1579-1585. https://doi. org/10.1016/j.phytochem.2010.06.005.

- [34] U. Grienke, T. Kaserer, B. Kirchweger, et al., Steroid sulfatase inhibiting lanostane triterpenes - structure activity relationship and *in silico* insights, Bioorg. Chem. 95 (2020) 103495. https://doi.org/10.1016/ j.bioorg.2019.103495.
- [35] J. Liu, K. Kurashiki, K. Shimizu, et al., Structure-activity relationship for inhibition of 5alpha-reductase by triterpenoids isolated from *Ganoderma lucidum*, Bioorg. Med. Chem. 14 (2006) 8654-8660. https://doi.org/10.1016/ j.bmc.2006.08.018.
- [36] Q. Wang, F. Wang, Z. Xu, et al., Bioactive mushroom polysaccharides: a review on monosaccharide composition, biosynthesis and regulation, Molecules 22 (2017) 955. https://doi.org/10.3390/molecules22060955.
- [37] J.H. Lu, R.J. He, P.L. Sun, et al., Molecular mechanisms of bioactive polysaccharides from *Ganoderma lucidum* (Lingzhi), a review, Int. J. Biol. Macromol. 150 (2020) 765-774. https://doi.org/10.1016/j.ijbiomac.2020.02.035.
- [38] X. Peng, M. Qiu, Meroterpenoids from *Ganoderma* species: a review of last five years, Nat. Prod. Bioprosp. 8 (2018) 137-149. https://doi.org/10.1007/ s13659-018-0164-z.
- [39] I. Lee, B. Ahn, J. Choi, et al., Selective cholinesterase inhibition by lanostane triterpenes from fruiting bodies of *Ganoderma lucidum*, Bioorg. Med. Chem. Lett. 21 (2011) 6603-6607. https://doi.org/10.1016/j.bmcl.2011.04.042.
- [40] I. Lee, J. Seo, J. Kim, et al., Lanostane triterpenes from the fruiting bodies of *Ganoderma lucidum* and their inhibitory effects on adipocyte differentiation in 3T3-L1 Cells, J. Nat. Prod. 73 (2010) 172-176. https://doi.org/10.1021/ np900578h.
- [41] S. El-Mekkawy, M.R. Meselhy, N. Nakamura, et al., Anti-HIV-1 and anti-HIV-1-protease substances from *Ganoderma lucidum*, Phytochemistry 49 (1998) 1651-1657. https://doi.org/10.1016/s0031-9422(98)00254-4.
- [42] T. Kikuchi, S. Kanomi, Y. Murai, et al., Constituents of the fungus Ganoderma lucidum (FR.) KARST. III.: structures of ganolucidic acids A and B, new lanostane-type triterpenoids, Chem. Pharm. Bull. 34 (1986) 4030-4036. https://doi.org/10.1248/cpb.34.4030.
- [43] T. Kikuchi, S. Matsuda, Y. Murai, et al., Ganoderic acid G and I and ganolucidic acid A and B, new triterpenoids from *Ganoderma lucidum*, Chem. Pharm. Bull. 33 (1985) 2628-2631. https://doi.org/10.1248/ cpb.33.2628.
- [44] T. Kubota, Y. Asaka, I. Miura, et al., Structures of ganoderic acid A and B, two new lanostane type bitter triterpenes from *Ganoderma lucidum* (FR.) KARST, Helv. Chim. Acta. 65 (1982) 611-619. https://doi.org/10.1002/ hlca.19820650221.
- [45] H. Kohda, W. Tokumoto, K. Sakamoto, et al., The biologically active constituents of *Ganoderma lucidum* (Fr.) Karst. Histamine release-inhibitory triterpenes, Chem. Pharm. Bull. (Tokyo) 33 (1985) 1367-1374. https://doi. org/10.1248/cpb.33.1367.
- [46] T. Kikuchi, S. Kanomi, S. Kadota, et al., Constituents of the fungus Ganoderma lucidum (Fr.) Karst. I structures of ganoderic acids C2, E, I, and K, lucidenic acid F and related compounds, Chem. Pharm. Bull. 34 (1986) 3695-3712. https://doi.org/10.1248/cpb.34.3695.
- [47] A. Morigiwa, K. Kitabatake, Y. Fujimoto, et al., Angiotensin converting enzyme-inhibitory triterpenes from *Ganoderma lucidum*, Chem. Pharm. Bull. (Tokyo) 34 (1986) 3025-3028. https://doi.org/10.1248/cpb.34.3025.
- [48] J. Luo, Z. Lin, Structure identification of triterpenes from fruiting bodies of *Ganoderma lucidum* by NMR spectra and X-ray diffraction analysis, Zhong Cao Yao 33 (2002) 197-200. https://doi.org/10.3321/ j.issn:0253-2670.2002.03.002.
- [49] J.J. Gao, B.S. Min, E.M. Ahn, et al., New triterpene aldehydes, lucialdehydes A-C, from *Ganoderma lucidum* and their cytotoxicity against murine and human tumor cells, Chem. Pharm. Bull. (Tokyo) 50 (2002) 837-840. https:// doi.org/10.1248/cpb.50.837.
- [50] S.H. Guan, M. Yang, X. Liu, et al., Two new lanostanoid triterpenes from the fruit body of *Ganoderma lucidum*-the major component of SunRecome®, Nat Prod Commun. 1 (2006) 177-181. https://doi. org/10.1177/1934578X0600100301.
- [51] T. Kikuchi, S. Kanomi, Y. Murai, et al., Constituents of the fungus Ganoderma lucidum (FR.) KARST. II.: structures of ganoderic acids F, G, and H, lucidenic acids D2 and E2, and related compounds, Chem. Pharm. Bull. 34 (1986) 4018-4029. https://doi.org/10.1248/cpb.34.4018.
- [52] Y. Komoda, H. Nakamura, S. Ishihara, et al., Structures of new terpenoid constituents of *Ganoderma lucidum* (Fr.) Karst (Polyporaceae), Chem. Pharm. Bull. 33 (1985) 4829-4835. https://doi.org/10.1248/cpb.33.4829.

- [53] S.H. Guan, J.M. Xia, M. Yang, et al., Cytotoxic lanostanoid triterpenes from *Ganoderma lucidum*, J. Asian Nat. Prod. Res. 10 (2008) 695-700. https://doi. org/10.1080/10286020802016297.
- [54] T. Kikuchi, S. Matsuda, S. Kadota, et al., Ganoderic acid D, E, F, and H and lucidenic acid D, E, and F, new triterpenoids from *Ganoderma lucidum*, Chem. Pharm. Bull. 33 (1985) 2624-2627. https://doi.org/10.1248/ cpb.33.2624.
- [55] M. Hirotani, T. Furuya, Ganoderic acid derivatives, highly oxygenated lanostane-type triterpenoids, from *Ganoderma lucidum*, Phytochemistry 25 (1986) 1189-1193. https://doi.org/10.1016/S0031-9422(00)81578-2.
- [56] C.F. Wang, J.Q. Liu, Y.X. Yan, et al., Three new triterpenoids containing four-membered ring from the fruiting body of *Ganoderma sinense*, Org. Lett. 12 (2010) 1656-1659. https://doi.org/10.1021/ol100062b.
- [57] Y.Y. Li, Z.Y. Mi, Y. Tang, et al., Lanostanoids isolated from *Ganoderma lucidum* mycelium cultured by submerged fermentation, Helv. Chim. Acta. 92 (2009) 1586-1593. https://doi.org/10.1002/hlca.200900028.
- [58] D.Z. Liu, Y.Q. Zhu, X.F. Li, et al., New triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their bioactivities, Chem. Biodivers. 11 (2014) 982-986. https://doi.org/10.1002/cbdv.201400004.
- [59] Y. Shao, L. Qiao, L. Wu, et al., Structure identification and anti-cancer pharmacological prediction of triterpenes from *Ganoderma lucidum*, Molecules 21 (2016) 678. https://doi.org/10.3390/molecules21050678.
- [60] T. Nishitoba, H. Sato, S. Sakamura, Triterpenoids from the fungus Ganoderma lucidum, Phytochemistry 26 (1987) 1777-1784. https://doi. org/10.1016/S0031-9422(00)82287-6.
- [61] J. Ma, Q. Ye, Y. Hua, et al., New lanostanoids from the mushroom *Ganoderma lucidum*, J. Nat. Prod. 65 (2002) 72-75. https://doi.org/10.1021/np010385e.
- [62] R.Y. Chen, D.Q. Yu, Studies on the triterpenoid constituents of the spores from *Ganoderma lucidum* Karst, J. Chin. Pharm. Sci. 2 (1993) 91-96. https:// doi.org/CNKI:SUN:XYGZ.0.1993-02-000.
- [63] C. Li, Y. Li, H.H. Sun, New ganoderic acids, bioactive triterpenoid metabolites from the mushroom *Ganoderma lucidum*, Nat. Prod. Res. 20 (2006) 985-991. https://doi.org/10.1080/14786410600921466.
- [64] J. Luo, Y.Y. Zhao, Z.B. Li, A new lanostane-type triterpene from the fruiting bodies of *Ganoderma lucidum*, J. Asian Nat. Prod. Res. 4 (2002) 129-134. https://doi.org/10.1080/10286020290027416.
- [65] B.S. Min, J.J. Gao, N. Nakamura, et al., Triterpenes from the spores of *Ganoderma lucidum* and their cytotoxicity against meth-A and LLC tumor cells, Chem. Pharm. Bull (Tokyo) 48 (2000) 1026-1033. https://doi. org/10.1248/cpb.48.1026.
- [66] B.S. Min, N. Nakamura, H. Miyashiro, et al., Triterpenes from the spores of *Ganoderma lucidum* and their inhibitory activity against HIV-1 protease, Chem. Pharm. Bull. (Tokyo) 46 (1998) 1607-1612. https://doi.org/10.1248/ cpb.46.1607.
- [67] T. Nishitoba, K. Oda, H. Sato, et al., Novel triterpenoids from the fungus Ganoderma lucidum, Agric. Biol. Chem. 52 (1988) 367-372. https://doi.org/ 10.1080/00021369.1988.10868655.
- [68] T. Nishitoba, H. Sato, K. Oda, et al., Novel triterpenoids and a steroid from the fungus *Ganoderma lucidum*, Agric. Biol. Chem. 52 (1988) 211-216. https://doi.org/10.1080/00021369.1988.10868604.
- [69] T. Nishitoba, H. Sato, S. Shirasu, et al., Novel triterpenoids from the mycelial mat at the previous stage of fruiting of *Ganoderma lucidum*, Agric. Biol. Chem. 51 (1987) 619-622. https://doi.org/10.1080/00021369.1987.10868026.
- [70] T. Nishitoba, H. Sato, S. Sakamura, Novel mycelial components, ganoderic acid Mg, Mh, Mi, Mj and Mk, from the fungus *Ganoderma lucidum*, Agric. Biol. Chem. 51 (1987) 1149-1153. https://doi.org/10.1080/00021369.1987.1 0868141.
- [71] A.G. Gonzalez, F. Leon, A. Rivera, et al., Lanostanoid triterpenes from Ganoderma lucidum, J. Nat. Prod. 62 (1999) 1700-1701. https://doi. org/10.1021/np990295y.
- [72] H. Cai, F.S. Wang, J.S. Yang, et al., Studies on the triterpenoid constituents from the fruiting body of *Ganoderma lucidum* (FR) Karst, Chin. J. Vet. Sci. 17 (1997) 511-513. https://doi.org/10.16303/j.cnki.1005-4545.1997.05.030.
- [73] B.J. Ma, Y. Zhou, Y. Ruan, et al., Lanostane-type triterpenes from the sporoderm-broken spores of *Ganoderma lucidum*, J. Antibiot. 65 (2012) 165-167. https://doi.org/10.1038/ja.2011.135.
- [74] Y.B. Li, R.M. Liu, J.J. Zhong, A new ganoderic acid from *Ganoderma lucidum* mycelia and its stability, Fitoterapia 84 (2013) 115-122. https://doi.org/10.1016/j.fitote.2012.11.008.

- [75] M. Hirotani, I. Asaka, C. Ino, et al., Ganoderic acid derivatives and ergosta-4,7,22-triene-3,6-dione from *Ganoderma lucidum*, Phytochemistry 26 (1987) 2797-2803. https://doi.org/10.1016/s0031-9422(00)83593-1.
- [76] J. Toth, B. Luu, J. Beck, et al., Chemistry and biochemistry of oriental drugs. Part IX. Cytotoxic triterpenes from *Ganoderma lucidum* (Polyporaceae): structures of ganoderic acids U-Z, J. Chem. Res. (1983) 299-299.
- [77] J.L. Wang, Y.B. Li, R.M. Liu, et al., A new ganoderic acid from *Ganoderma lucidum* mycelia, J. Asian Nat. Prod. Res. 12 (2010) 727-730. https://doi.org/ 10.1080/10286020.2010.493506.
- [78] H.H. Ko, C.F. Hung, J.P. Wang, et al., Antiinflammatory triterpenoids and steroids from *Ganoderma lucidum* and *G. tsugae*, Phytochemistry 69 (2008) 234-239. https://doi.org/10.1016/j.phytochem.2007.06.008.
- [79] M. Yang, X. Wang, S. Guan, et al., Analysis of triterpenoids in *Ganoderma lucidum* using liquid chromatography coupled with electrospray ionization mass spectrometry, J. Am. Soc. Mass Spectrom. 18 (2007) 927-939. https://doi.org/10.1016/j.jasms.2007.01.012.
- [80] X.R. Zhao, B.J. Zhang, S. Deng, et al., Isolation and identification of oxygenated lanostane-type triterpenoids from the fungus *Ganoderma lucidum*, Phytochem. Lett. 16 (2016) 87-91. https://doi.org/10.1016/ j.phytol.2016.03.007.
- [81] H.G. Su, X.R. Peng, Q.Q. Shi, et al., *Lanostane triterpenoids* with antiinflammatory activities from *Ganoderma lucidum*, Phytochemistry 173 (2020) 112256. https://doi.org/10.1016/j.phytochem.2019.112256.
- [82] M. Chen, M. Zhang, S. Sun, et al., A new triterpene from the fruiting bodies of *Ganoderma lucidum*, Yao Xue Xue Bao 44 (2009) 768-770. https://doi. org/10.3321/j.issn:0513-4870.2009.07.015.
- [83] S. Chen, X. Li, T. Yong, et al., Cytotoxic lanostane-type triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their structure-activity relationships, Oncotarget 8 (2017) 10071-10084. https://doi.org/10.18632/ oncotarget.14336.
- [84] S.H. Guan, M. Yang, X.M. Wang, et al., Structure elucidation and complete NMR spectral assignments of three new lanostanoid triterpenes with unprecedented Delta (16,17) double bond from *Ganoderma lucidum*, Magn. Reson. Chem. 45 (2007) 789-791. https://doi.org/10.1002/mrc.2046.
- [85] R.X. Hao, J.S. Zhang, Q. Tang, et al., Isolation, purification and identification of two new triterpenoid constituents from the fruiting bodies of *Ganoderma lucidum*, Mycosystema 25 (2006) 599-602. https://doi. org/10.1016/S1872-2075(06)60070-8.
- [86] T. Akihisa, M. Tagata, M. Ukiya, et al., Oxygenated lanostane-type triterpenoids from the fungus *Ganoderma lucidum*, J. Nat. Prod. 68 (2005) 559-563. https://doi.org/10.1021/np040230h.
- [87] C.J. Weng, C.F. Chau, K.D. Chen, et al., The anti-invasive effect of lucidenic acids isolated from a new *Ganoderma lucidum* strain, Mol. Nutr. Food Res. 51 (2007) 1472-1477. https://doi.org/10.1002/mnfr.200700155.
- [88] P. Li, Y.P. Deng, X.X. Wei, et al., Triterpenoids from *Ganoderma lucidum* and their cytotoxic activities, Nat. Prod. Res. 27 (2013) 17-22. https://doi.org /10.1080/14786419.2011.652961.
- [89] T. Nishitoba, H. Sato, T. Kasai, et al., New bitter C27 and C30 terpenoids from the fungus *Ganoderma lucidum* (Reishi), Agric. Biol. Chem. 49 (1985) 1793-1798. https://doi.org/10.1080/00021369.1985.10866955.
- [90] K. Iwatsuki, T. Akihisa, H. Tokuda, et al., Lucidenic acids P and Q, methyl lucidenate P, and other triterpenoids from the fungus *Ganoderma lucidum* and their inhibitory effects on Epstein-Barr virus activation, J. Nat. Prod. 66 (2003) 1582-1585. https://doi.org/10.1021/np0302293.
- [91] I. Lee, H. Kim, U. Youn, et al., Effect of lanostane triterpenes from the fruiting bodies of *Ganoderma lucidum* on adipocyte differentiation in 3T3-L1 cells, Planta. Med. 76 (2010) 1558-1563. https://doi.org/10.1055/s-0030-1249827.
- [92] N.T. Tung, T.D. Cuong, T.M. Hung, et al., Inhibitory effect on NO production of triterpenes from the fruiting bodies of *Ganoderma lucidum*, Bioorg. Med. Chem. Lett. 23 (2013) 1428-1432. https://doi.org/10.1016/ j.bmcl.2012.12.066.
- [93] T. Nishitoba, S. Sato, S. Sakamura, New terpenoids from *Ganoderma lucidum* and their bitterness, Agric. Biol. Chem. 49 (1985) 1547-1549. https://doi.org/10.1080/00021369.1985.10866944.
- [94] T. Nishitoba, H. Sato, S. Sakamura, New terpenoids, ganolucidic acid D, ganoderic acid L, lucidone C and lucidenic acid G, from the fungus *Ganoderma lucidum*, Agric. Biol. Chem. 50 (1986) 809-811. https://doi.org/ 10.1080/00021369.1986.10867474.

- [95] X.Q. Zhang, F.C.F. Ip, D.M. Zhang, et al., Triterpenoids with neurotrophic activity from *Ganoderma lucidum*, Nat. Prod. Res. 25 (2011) 1607-1613. https://doi.org/10.1080/14786419.2010.496367.
- [96] C. Li, J. Yin, F. Guo, et al., Ganoderic acid Sz, a new lanostanoid from the mushroom *Ganoderma lucidum*, Nat. Prod. Res. 19 (2005) 461-465. https:// doi.org/10.1080/14786410412331272077.
- [97] M. Arisawa, A. Fujita, M. Saga, et al., Three new lanostanoids from Ganoderma lucidum, J. Nat. Prod. 49 (1986) 621-625. https://doi. org/10.1021/np50046a010.
- [98] H. Sato, T. Nishitoba, S. Shirasu, et al., Ganoderiol A and B, new triterpenoids from the fungus *Ganoderma lucidum* (Reishi), Agric. Biol. Chem. 50 (1986) 2887-2890. https://doi.org/10.1080/00021369.1986.10867818.
- [99] M.S. Shiao, L.J. Lin, S.F. Yeh, et al., Two new triterpenes of the fungus Ganoderma lucidum, J. Nat. Prod. 50 (1987) 886-890. https://doi. org/10.1021/np50053a019.
- [100] M. Hirotani, C. Ino, T. Furuya, et al., Ganoderic acids T, S and R, new triterpenoids from the cultured mycelia of *Ganoderma lucidum*, Chem. Pharm. Bull. 34 (1986) 2282-2285. https://doi.org/10.1248/cpb.34.2282.
- [101] M.S. Shiao, L.J. Lin, S.F. Yeh, Triterpenes from *Ganoderma lucidum*, Phytochemistry 27 (1988) 2911-2914. https://doi.org/10.1016/0031-9422(88)80687-3.
- [102] L.J. Lin, M.S. Shiao, S.F. Yeh, Triterpenes from *Ganoderma lucidum*, Phytochemistry 27 (1988) 2269-2271. https://doi.org/10.1016/0031-9422(88)80140-7.
- [103] M. Hirotani, I. Asaka, C. Ino, et al., Studies on the metabolites of higher fungi .7. ganoderic acid-derivatives and ergosta-4,7,22-triene-3,6-dione from *Ganoderma-lucidum*, Phytochemistry 26 (1987) 2797-2803. https://doi. org/10.1016/S0031-9422(00)83593-1.
- [104] C. Gerhäuser, W.D. Zhang, N. Ho-Chong-Line, et al., New lanostanoids from *Ganoderma lucidum* that induce NAD (P) H: qui-none oxidoreductase in cultured Hepalclc7 murine hepatoma cells, Planta Med. 66 (2000) 681-684. https://doi.org/10.1055/s-2000-8647.
- [105] M.S. Shiao, L.J. Lin, S.F. Yeh, Triterpenes in *Ganoderma lucidum*, Phytochemistry 27 (1988) 873-875. https://doi.org/10.1016/0031-9422(88)84110-4.
- [106] J. Toth, B. Luu, G. Ourisson, Ganoderic acid T and Z: cytotoxic triterpenes from *G. lucidum*, Tetrahedron. Lett. 24 (1983) 1081-1084. https://doi. org/10.1016/S0040-4039(00)81610-X.
- [107] M. Adams, M. Christen, I. Plitzko, et al., Antiplasmodial lanostanes from the *Ganoderma lucidum* mushroom, J. Nat. Prod. 73 (2010) 897-900. https://doi. org/10.1021/np100031c.
- [108] L.J. Lin, M.S. Shiao, S.F. Yeh, Seven new triterpenes from Ganoderma lucidum, J. Nat. Prod. 51 (1988) 918-924. https://doi.org/10.1021/ np50059a017.
- [109] A. Fujita, M. Arisawa, M. Saga, et al., Two new lanostanoids from Ganoderma lucidum, J. Nat. Prod. 49 (1986) 1122-1125. https://doi. org/10.1021/np50048a029.
- [110] R.Y. Chen, D.Q. Yu, Application of 2d NMR techniques in the structure determination of ganosporelactone A and B, Yao Xue Xue Bao 26 (1991) 430-436.
- [111] M. Hirotani, C. Ino, T. Furuya, Comparative study on the strain-specific triterpenoid components of *Ganoderma lucidum*, Phytochemistry 33 (1993) 379-382. https://doi.org/10.1016/0031-9422(93)85523-T.
- [112] F. Wang, H. Cai, J. Yang, et al., Triterpenoids from the fruiting body of *Ganoderma lucidum*, J. Chin. Pharm. Sci. 4 (1997) 20-25.
- [113] M. Hirotani, T. Furuya, M. Shiro, A ganoderic acid derivative, a highly oxygenated lanostane-type triterpenoid from *Ganoderma lucidum*, Phytochemistry 24 (1985) 2055-2061. https://doi.org/10.1016/S0031-9422(00)83121-0.
- [114] Y. Mizushina, N. Takahashi, L. Hanashima, Lucidenic acid O and lactone, new terpene inhibitors of eukaryotic DNA polymerases from a basidiomycete, Bioorg. Med. Chem. 7 (1999) 2047-2052. https://doi. org/10.1016/S0968-0896(99)00121-2.
- [115] S. Joseph, K.K. Janardhanan, V. George, et al., A new epoxidic ganoderic acid and other phytoconstituents from *Ganoderma lucidum*, Phytochem. Lett. 4 (2011) 386-388. https://doi.org/10.1016/j.phytol.2011.08.011.
- [116] T. Nishitoba, H. Sato, S. Sakamura, New terpenoids, ganoderic acid J and ganolucidic acid C, from the fungus *Ganoderma lucidum*, Agric. Biol. Chem. 49 (1985) 3637-3638. https://doi.org/10.1080/00021369.1985.10867324.

- [117] X.Q. Che, S.P. Li, J. Zhao, Ganoderma triterpenoids from aqueous extract of Ganoderma lucidum, Zhongguo Zhong Yao Za Zhi 42 (2017) 1908-1915. https://doi.org/10.19540/j.cnki.cjcmm.20170412.001.
- [118] M.H. Koo, H.J. Chae, J.H. Lee, et al., Antiinflammatory lanostane triterpenoids from *Ganoderma lucidum*, Nat. Prod. Res. 35 (2019) 4295-4302. https://doi.org/10.1080/14786419.2019.1705815.
- [119] C. Murata, Q.T. Tran, S. Onda, et al., Extraction and isolation of ganoderic acid sigma from *Ganoderma lucidum*, Tetrahedron Lett. 57 (2016) 5368-5371. https://doi.org/10.1016/j.tetlet.2016.10.072.
- [120] I.C. Ferreira, S.A. Heleno, F.S. Reis, et al., Chemical features of *Ganoderma* polysaccharides with antioxidant, antitumor and antimicrobial activities, Phytochemistry 114 (2015) 38-55. https://doi.org/10.1016/ j.phytochem.2014.10.011.
- [121] M. Tomoda, R. Gonda, Y. Kasahara, et al., Glycan structures of ganoderans B and C, hypoglycemic glycans of *Ganoderma lucidum* fruit bodies, Phytochemistry 25 (1986) 2817-2820. https://doi.org/10.1016/S0031-9422(00)83748-6.
- [122] Q. Chen, R. Li, Y. He, Studies on anti-aging polysaccharides GLB GLC of Ganoderma lucidum Δ, Beijing Yike Daxue Xuebao 25 (1993) 303-305. https://doi.org/10.1007/BF02005919.
- [123] T. Li, Y. He, R. Li, The study of polysaccharides of *Ganoderma lucidum* from Tai Mountain, China J. Chinese Matera. Medica. 22 (1997) 487-489. https://doi.org/10.3321/j.issn:1001-5302.1997.08.016.
- [124] X.F. Bao, X.S. Wang, Q. Dong, et al., Structural features of immunologically active polysaccharides from *Ganoderma lucidum*, Phytochemistry 59 (2002) 175-181. https://doi.org/10.1016/s0031-9422(01)00450-2.
- [125] X.F. Bao, Y. Zhen, L. Ruan, et al., Purification, characterization, and modification of T lymphocyte-stimulating polysaccharide from spores of *Ganoderma lucidum*, Chem. Pharm. Bull. (Tokyo) 50 (2002) 623-629. https://doi.org/10.1248/cpb.50.623.
- [126] J. Zhang, Q. Tang, M. Zimmerman-Kordmann, et al., Activation of B lymphocytes by GLIS, a bioactive proteoglycan from *Ganoderma lucidum*, Life Sci. 71 (2002) 623-638. https://doi.org/10.1016/s0024-3205(02)01690-9.
- [127] S. Lin, S. Wang, Z. Lin, et al., Isolation and identification of active components of *Ganoderma lucidum* cultivated with grass and wood log I. extraction, purification and characterization of glycopeptide, Zhong Cao Yao 34 (2003) 872-874. https://doi.org/10.3321/j.issn:0253-2670.2003.10.003.
- [128] S.Z. Wang, K. Ding, S.Q. Lin, et al., Isolation, purification and structural analysis of GL-PP-3A, an active polysaccharide peptide from *Ganoderma lucidum*, Yao Xue Xue Bao 42 (2007) 1058-1061. https://doi.org/10.3321/ j.issn:0513-4870.2007.10.010.
- [129] D. Shang, J. Zhang, L. Wen, et al., Preparation, characterization, and antiproliferative activities of the Se-containing polysaccharide SeGLP-2B-1 from Se-enriched *Ganoderma lucidum*, J. Agric. Food Chem. 57 (2009) 7737-7742. https://doi.org/10.1021/jf9019344.
- [130] J. Wang, L. Zhang, Structure and chain conformation of five water-soluble derivatives of a beta-D-glucan isolated from *Ganoderma lucidum*, Carbohydr. Res. 344 (2009) 105-112. https://doi.org/10.1016/j.carres.2008.09.024.
- [131] Y. Liu, J. Zhang, Q. Tang, et al., Physicochemical characterization of a high molecular weight bioactive β-D-glucan from the fruiting bodies of *Ganoderma lucidum*, Carbohydr. Polym. 101 (2014) 968-974. https://doi. org/10.1016/j.carbpol.2013.10.024.
- [132] J. Li, F. Gu, C. Cai, et al., Purification, structural characterization, and immunomodulatory activity of the polysaccharides from *Ganoderma lucidum*, Int. J. Biol. Macromol. 143 (2020) 806-813. https://doi.org/10.1016/ j.ijbiomac.2019.09.141.
- [133] R.Y. Chen, Y.H. Wang, D.Q. Yu, Studies on the chemical constituents of the spores from *Ganoderma lucidum*, J. Integr. Plant Biol. 33 (1991) 65-68.
- [134] H.W. Seo, T.M. Hung, M. Na, et al., Steroids and triterpenes from the fruit bodies of *Ganoderma lucidum* and their anti-complement activity, Arch. Pharm. Res. 32 (2009) 1573-1579. https://doi.org/10.1007/s12272-009-2109-x.
- [135] C.R. Zhang, S.P. Yang, J.M. Yue, Sterols and triterpenoids from the spores of *Ganoderma lucidum*, Nat. Prod. Res. 22 (2008) 1137-1142. https://doi. org/10.1080/14786410601129721.
- [136] Y. Weng, L. Xiang, A. Matsuura, et al., Ganodermasides A and B, two novel anti-aging ergosterols from spores of a medicinal mushroom *Ganoderma lucidum* on yeast via UTH1 gene, Bioorg. Med. Chem. 18 (2010) 999-1002. https://doi.org/10.1016/j.bmc.2009.12.070.

- [137] Y. Weng, J. Lu, L. Xiang, et al., Ganodermasides C and D, two new antiaging ergosterols from spores of the medicinal mushroom *Ganoderma lucidum*, Biosci. Biotechnol. Biochem. 75 (2011) 800-803. https://doi. org/10.1271/bbb.100918.
- [138] C.N. Lin, W.P. Tome, S.J. Won, Novel cytotoxic principles of Formosan Ganoderma lucidum, J. Nat. Prod. 54 (1991) 998-1002. https://doi. org/10.1021/np50076a012.
- [139] Y.K. Chen, Y.H. Kuo, B.H. Chiang, et al., Cytotoxic activities of 9,11-dehydroergosterol peroxide and ergosterol peroxide from the fermentation mycelia of *Ganoderma lucidum* cultivated in the medium containing leguminous plants on Hep 3B cells, J. Agric. Food Chem. 57 (2009) 5713-5719. https://doi.org/10.1021/jf900581h.
- [140] F.C. Ziegenbein, H.P. Hanssen, W.A. König, Secondary metabolites from Ganoderma lucidum and Spongiporus leucomallellus, Phytochemistry 67 (2006) 202-211. https://doi.org/10.1016/j.phytochem.2005.10.025.
- [141] R.A. Mothana, R. Jansen, W.D. Jülich, et al., Ganomycins A and B, new antimicrobial farnesyl hydroquinones from the basidiomycete *Ganoderma pfeifferi*, J. Nat. Prod. 63 (2000) 416-418. https://doi.org/10.1021/np990381y.
- [142] Q. Luo, X.L. Wang, L. Di, et al., Isolation and identification of renoprotective substances from the mushroom *Ganoderma lucidum*, Tetrahedron. 71 (2015) 840-845. https://doi.org/10.1016/j.tet.2014.12.052.
- [143] X.F. Wang, Y.M. Yan, X.L. Wang, et al., Two new compounds from *Ganoderma lucidum*, J. Asian Nat. Prod. Res. 17 (2015) 329-332. https://doi. org/10.1080/10286020.2014.960858.
- [144] F.J. Zhou, X.L. Wang, S.M. Wang, et al., A new meroterpenoid from *Ganoderma lucidum*, Nat. Prod. Res. Dev. 27 (2015) 22-25. https://doi. org/10.16333/j.1001-6880.2015.01.004.
- [145] Y.M. Yan, J. Ai, L.L. Zhou, et al., Lingzhiols, unprecedented rotary doorshaped meroterpenoids as potent and selective inhibitors of p-Smad3 from *Ganoderma lucidum*, Org. Lett. 15 (2013) 5488-5491. https://doi. org/10.1021/ol4026364.
- [146] Z.Z. Zhao, H.P. Chen, T. Feng, et al., Lucidimine A-D, four new alkaloids from the fruiting bodies of *Ganoderma lucidum*, J. Asian Nat. Prod. Res. 17 (2015) 1160-1165. https://doi.org/10.1080/10286020.2015.1119128.
- [147] A. Shimizu, T. Yano, Y. Saito, et al., Isolation of an inhibitor of platelet aggregation from a fungus, *Ganoderma lucidum*, Chem. Pharm. Bull. 33 (1985) 3012-3015. https://doi.org/10.1248/cpb.33.3012.
- [148] Y. Mizushina, L. Hanashima, T. Yamaguchi, et al., A mushroom fruiting body-inducing substance inhibits activities of replicative DNA polymerases, Biochem. Biophys. Res. Commun. 249 (1998) 17-22. https://doi. org/10.1006/bbrc.1998.9091.
- [149] Y. Jiao, T. Xie, L.H. Zou, et al., Lanostane triterpenoids from *Ganoderma curtisii* and their NO production inhibitory activities of LPS-induced microglia, Bioorg. Med. Chem. Lett. 26 (2016) 3556-3561. https://doi.org/10.1016/j.bmcl.2016.06.023.
- [150] S. Joseph, S. Baby, V. George, et al., Antioxidative and antiinflammatory activities of the chloroform extract of *Ganoderma lucidum* found in South India, Sci. Pharm. 77 (2009) 111-121. https://doi.org/10.3797/scipharm.0808-17.
- [151] J.G. Wu, Y.J. Kan, Y.B. Wu, et al., Hepatoprotective effect of *Ganoderma* triterpenoids against oxidative damage induced by tert-butyl hydroperoxide in human hepatic HepG2 cells, Pharm. Biol. 54 (2016) 919-929. https://doi. org/10.3109/13880209.2015.1091481.
- [152] L. Li, H.J. Guo, L.Y. Zhu, et al., A supercritical-CO2 extract of *Ganoderma lucidum* spores inhibits cholangiocarcinoma cell migration by reversing the epithelial-mesenchymal transition, Phytomedicine 23 (2016) 491-497. https://doi.org/10.1016/j.phymed.2016.02.019.
- [153] X.R. Zhao, X.K. Huo, P.P. Dong, et al., Inhibitory effects of highly oxygenated lanostane derivatives from the fungus *Ganoderma lucidum* on P-glycoprotein and α-glucosidase, J. Nat. Prod. 78 (2015) 1868-1876. https:// doi.org/10.1021/acs.jnatprod.5b00132.
- [154] D. Kang, M. Mutakin, J. Levita, Computational study of triterpenoids of *Ganoderma lucidum* with aspartic protease enzymes for discovering HIV-1 and plasmepsin inhibitors, Int. J. Chem. 7 (2015) 62. https://doi.org/10.5539/ ijc.v7n1p62.
- [155] A. Berger, D. Rein, E. Kratky, et al., Cholesterol-lowering properties of *Ganoderma lucidum in vitro*, ex vivo, and in hamsters and minipigs, Lipids Health Dis. 3 (2004) 2. https://doi.org/10.1186/1476-511x-3-2.
- [156] Y. Kabir, S. Kimura, T. Tamura, Dietary effect of *Ganoderma lucidum* mushroom on blood pressure and lipid levels in spontaneously hypertensive

rats (SHR), J. Nutr. Sci. Vitaminol (Tokyo) 34 (1988) 433-438. https://doi. org/10.3177/jnsv.34.433.

- [157] J.L. Gao, K.S. Leung, Y.T. Wang, et al., Qualitative and quantitative analyses of nucleosides and nucleobases in *Ganoderma* spp. by HPLC-DAD-MS, J. Pharm. Biomed. Anal. 44 (2007) 807-811. https://doi.org/10.1016/ j.jpba.2007.03.012.
- [158] T.H. Chen, M.F. Wang, Y.F. Liang, et al., A nucleoside-nucleotide mixture may reduce memory deterioration in old senescence-accelerated mice, J. Nutr. 130 (2000) 3085-3089. https://doi.org/10.1093/jn/130.12.3085.
- [159] Y. Chen, S.B. Zhu, M.Y. Xie, et al., Quality control and original discrimination of *Ganoderma lucidum* based on high-performance liquid chromatographic fingerprints and combined chemometrics methods, Anal. Chim. Acta. 623 (2008) 146-156. https://doi.org/10.1016/j.aca.2008.06.018.
- [160] D.T. Ha, L.T. Loan, T.M. Hung, et al., An improved HPLC-DAD method for quantitative comparisons of triterpenes in *Ganoderma lucidum* and its five related species originating from Vietnam, Molecules 20 (2015) 1059-1077. https://doi.org/10.3390/molecules20011059.
- [161] M.S. Khan, R. Parveen, K. Mishra, et al., Determination of nucleosides in *Cordyceps sinensis* and *Ganoderma lucidum* by high performance liquid chromatography method, J. Pharm. Bioallied Sci. 7 (2015) 264-266. https:// doi.org/10.4103/0975-7406.168022.
- [162] C. Zhang, D. Fu, G. Chen, et al., Comparative and chemometric analysis of correlations between the chemical fingerprints and anti-proliferative activities of ganoderic acids from three *Ganoderma* species, Phytochem. Anal. 30 (2019) 474-480. https://doi.org/10.1002/pca.2830.
- [163] Y. Chen, Y. Yan, M.Y. Xie, et al., Development of a chromatographic fingerprint for the chloroform extracts of *Ganoderma lucidum* by HPLC and LC-MS, J. Pharm. Biomed. Anal. 47 (2008) 469-477. https://doi. org/10.1016/j.jpba.2008.01.039.
- [164] Y. Liu, Y. Liu, F. Qiu, et al., Sensitive and selective liquid chromatographytandem mass spectrometry method for the determination of five ganoderic acids in *Ganoderma lucidum* and its related species, J. Pharm. Biomed. Anal. 54 (2011) 717-721. https://doi.org/10.1016/j.jpba.2010.11.002.
- [165] X.M. Wang, M. Yang, S.H. Guan, et al., Quantitative determination of six major triterpenoids in *Ganoderma lucidum* and related species by high performance liquid chromatography, J. Pharm. Biomed. Anal. 41 (2006) 838-844. https://doi.org/10.1016/j.jpba.2006.01.053.
- [166] L. Wu, W. Liang, W. Chen, et al., Screening and analysis of the marker components in *Ganoderma lucidum* by HPLC and HPLC-MS(n) with the aid of chemometrics, Molecules 22 (2017) 584. https://doi.org/10.3390/ molecules22040584.
- [167] J. Da, C.R. Cheng, S. Yao, et al., A reproducible analytical system based on the multi-component analysis of triterpene acids in *Ganoderma lucidum*, Phytochemistry 114 (2015) 146-154. https://doi.org/10.1016/ j.phytochem.2014.08.007.
- [168] J. Da, W.Y. Wu, J.J. Hou, et al., Comparison of two officinal Chinese pharmacopoeia species of *Ganoderma* based on chemical research with multiple technologies and chemometrics analysis, J. Chromatogr. A. 1222 (2012) 59-70. https://doi.org/10.1016/j.chroma.2011.12.017.
- [169] D.A. Frommenwiler, D. Trefzer, M. Schmid, et al., Comprehensive HPTLC fingerprinting: a novel economic approach to evaluating the quality of *Ganoderma lucidum* fruiting body, J. Liq. Chromatogr. Relat. Technol. 43 (2020) 414-423. https://doi.org/10.1080/10826076.2020.1725560.
- [170] M.Y. Shen, M.Y. Xie, S.P. Nie, et al., Discrimination of different *Ganoderma* species and their region based on GC-MS profiles of sterols and pattern recognition techniques, Anal. Lett. 44 (2011) 863-873. https://doi. org/10.1080/00032711003790007.
- [171] X.M. Shi, J.S. Zhang, Q.J. Tang, et al., Fingerprint analysis of Lingzhi (*Ganoderma*) strains by high-performance liquid chromatography coupled with chemometric methods, World J. Microbiol. Biotechnol. 24 (2008) 2443-2450. https://doi.org/10.1007/s11274-008-9766-7.
- [172] H. Zhang, H. Jiang, Y. Chen, et al., Quality evaluation of triterpenoids in *Ganoderma* and related species by the quantitative analysis of multicomponents by single marker method, J. Liq. Chromatogr. Relat. Technol. 41 (2018) 860-867. https://doi.org/10.1080/10826076.2018.1531292.
- [173] H. Zhang, H. Jiang, X. Zhang, et al., Development of global chemical profiling for quality assessment of *Ganoderma* species by ChemPattern software, J. Anal. Methods Chem. 2018 (2018) 1675721. https://doi. org/10.1155/2018/1675721.

- [174] Y. Chen, W. Bicker, J. Wu, et al., *Ganoderma* species discrimination by dual-mode chromatographic fingerprinting: a study on stationary phase effects in hydrophilic interaction chromatography and reduction of sample misclassification rate by additional use of reversed-phase chromatography, J. Chromatogr. A. 1217 (2010) 1255-1265. https://doi.org/10.1016/ j.chroma.2009.12.024.
- [175] Y. Chen, W. Bicker, J. Wu, et al., Simultaneous determination of 16 nucleosides and nucleobases by hydrophilic interaction chromatography and its application to the quality evaluation of *Ganoderma*, J. Agric. Food Chem. 60 (2012) 4243-4252. https://doi.org/10.1021/jf300076j.
- [176] Z. Qian, J. Zhao, D. Li, et al., Analysis of global components in *Ganoderma* using liquid chromatography system with multiple columns and detectors, J. Sep. Sci. 35 (2012) 2725-2734. https://doi.org/10.1002/jssc.201200441.
- [177] Y. Chen, M.Y. Xie, Y. Yan, et al., Discrimination of *Ganoderma lucidum* according to geographical origin with near infrared diffuse reflectance spectroscopy and pattern recognition techniques, Anal. Chim. Acta. 618 (2008) 121-130. https://doi.org/10.1016/j.aca.2008.04.055.
- [178] G. Yao, Y. Ma, M. Muhammad, et al., Understanding the infrared and Raman spectra of ganoderic acid A: an experimental and DFT study, Spectrochim. Acta A Mol. Biomol. Spectrosc. 210 (2019) 372-380. https:// doi.org/10.1016/j.saa.2018.11.019.
- [179] National Pharmacopoeia Committee, Pharmacopoeia of the People's Republic of China, China Medical Science Press, Beijing, 2020.
- [180] X. Shi, X. Gan, X. Wang, et al., Rapid detection of *Ganoderma lucidum* spore powder adulterated with dyed starch by NIR spectroscopy and chemometrics, LWT-Food Sci. Technol. 167 (2022) 113829. https://doi. org/10.1016/j.lwt.2022.113829.
- [181] The United States Pharmacopeial Convention, The United States Pharmacopeia, 40th Edition ed, 2017.
- [182] N. Krone, B.A. Hughes, G.G. Lavery, et al., Gas chromatography/mass spectrometry (GC/MS) remains a pre-eminent discovery tool in clinical steroid investigations even in the era of fast liquid chromatography tandem mass spectrometry (LC/MS/MS), J. Steroid. Biochem. Mol. Biol. 121 (2010) 496-504. https://doi.org/10.1016/j.jsbmb.2010.04.010.
- [183] H. Messai, M. Farman, A. Sarraj-Laabidi, et al., Chemometrics methods for specificity, authenticity and traceability analysis of olive oils: principles, classifications and applications, Foods 5 (2016) 77. https://doi.org/10.3390/ foods5040077.
- [184] D. Cör, Ž. Knez, M. Knez Hrnčič, Antitumour, antimicrobial, antioxidant and antiacetylcholinesterase effect of *Ganoderma lucidum* terpenoids and polysaccharides: a review, Molecules 23 (2018) 649. https://doi.org/10.3390/ molecules23030649.
- [185] T. Miyazaki, M. Nishijima, Studies on fungal polysaccharides. XXVII. Structural examination of a water-soluble, antitumor polysaccharide of *Ganoderma lucidum*, Chem. Pharm. Bull. 29 (1981) 3611-3616. https://doi. org/10.1248/cpb.29.3611.
- [186] J.G. Wang, Z.C. Ma, L.N. Zhang, et al., Structure and chain conformation of water-soluble heteropolysaccharides from *Ganoderma lucidum*, Carbohydr. Polym. 86 (2011) 844-851. https://doi.org/10.1016/j.carbpol.2011.05.031.
- [187] H. Zhang, J.Q. Wang, S.P. Nie, et al., Sulfated modification, characterization and property of a water-insoluble polysaccharide from *Ganoderma atrum*, Int. J. Biol. Macromol. 79 (2015) 248-255. https://doi.org/10.1016/ j.ijbiomac.2015.04.070.
- [188] L. Zhao, Y. Dong, G. Chen, et al., Extraction, purification, characterization and antitumor activity of polysaccharides from *Ganoderma lucidum*, Carbohydr. Polym. 80 (2010) 783-789. https://doi.org/10.1016/ j.carbpol.2009.12.029.
- [189] S.Q. Huang, J.W. Li, Y.Q. Li, et al., Purification and structural characterization of a new water-soluble neutral polysaccharide GLP-F1-1 from *Ganoderma lucidum*, Int. J. Biol. Macromol. 48 (2011) 165-169. https://doi.org/10.1016/j.ijbiomac.2010.10.015.
- [190] S.Q. Huang, Z.X. Ning, Extraction of polysaccharide from *Ganoderma lucidum* and its immune enhancement activity, Int. J. Biol. Macromol. 47 (2010) 336-341. https://doi.org/10.1016/j.ijbiomac.2010.03.019.
- [191] M. Shi, Y. Yang, X. Hu, et al., Effect of ultrasonic extraction conditions on antioxidative and immunomodulatory activities of a *Ganoderma lucidum* polysaccharide originated from fermented soybean curd residue, Food Chem. 155 (2014) 50-56. https://doi.org/10.1016/j.foodchem.2014.01.037.

- [192] M. Shi, Z. Zhang, Y. Yang, Antioxidant and immunoregulatory activity of *Ganoderma lucidum* polysaccharide (GLP), Carbohydr. Polym. 95 (2013) 200-206. https://doi.org/10.1016/j.carbpol.2013.02.081.
- [193] Y. Li, L. Fang, K. Zhang, Structure and bioactivities of a galactose rich extracellular polysaccharide from submergedly cultured *Ganoderma lucidum*, Carbohydr. Polym. 68 (2007) 323-328. https://doi.org/10.1016/ j.carbpol.2006.12.001.
- [194] W. Liu, H. Wang, X. Pang, et al., Characterization and antioxidant activity of two low-molecular-weight polysaccharides purified from the fruiting bodies of *Ganoderma lucidum*, Int. J. Biol. Macromol. 46 (2010) 451-457. https:// doi.org/10.1016/j.ijbiomac.2010.02.006.
- [195] W. Liu, J. Xu, P. Jing, et al., Preparation of a hydroxypropyl *Ganoderma lucidum* polysaccharide and its physicochemical properties, Food Chem. 122 (2010) 965-971. https://doi.org/10.1016/j.foodchem.2009.11.087.
- [196] Z.L. Zhao, T.M. Yu, L.P. Zhang, Chemical study on the water soluble polysaccharide from spores of *Ganoderma lucidum*, Nat. Prod. Res. Dev. 17 (2005) 182-185. https://doi.org/10.16333/j.1001 -6880.2005.02.017.
- [197] C. Huang, X.D. Gao, X.B. Pang, et al., Isolation, purification, composition and activity of *Ganoderma lucidum* polysaccharide, Chin. J. Biochem. Pharm. 26 (2005) 221-223. https://doi.org/10.3969/ j.issn.1005-1678.2005.04.011.
- [198] X. Huang, H. Wu, F. Huang, et al., Analysis of polysaccharide from broken cellular wall and unbroken spore of *Ganoderma lucidum*, Zhong Cao Yao 37 (2006) 813-816. https://doi.org/10.1360/yc-006-1325.
- [199] Y. Sone, R. Okuda, N. Wada, et al., Structures and antitumor activities of the polysaccharides isolated from fruiting body and the growing culture of mycelium of *Ganoderma lucidum*, Agric. Biol. Chem. 49 (1985) 2641-2653. https://doi.org/10.1080/00021369.1985.10867134.
- [200] J. Xu, W. Liu, W. Yao, et al., Carboxymethylation of a polysaccharide extracted from *Ganoderma lucidum* enhances its antioxidant activities *in vitro*, Carbohydr. Polym. 78 (2009) 227-234. https://doi.org/10.1016/ j.carbpol.2009.03.028.
- [201] L. Lai, D. Yang, Rheological properties of the hot-water extracted polysaccharides in Ling-Zhi (*Ganoderma lucidum*), Food Hydrocoll. 21 (2007) 739-746. https://doi.org/10.1016/j.foodhyd.2006.09.009.
- [202] X. Di, K.K. Chan, H.W. Leung, et al., Fingerprint profiling of acid hydrolyzates of polysaccharides extracted from the fruiting bodies and spores of Lingzhi by high-performance thin-layer chromatography, J. Chromatogr. A. 1018 (2003) 85-95. https://doi.org/10.1016/j.chroma.2003.07.015.
- [203] D.T. Wu, J. Xie, D.J. Hu, et al., Characterization of polysaccharides from *Ganoderma* spp. using saccharide mapping, Carbohydr. Polym. 97 (2013) 398-405. https://doi.org/10.1016/j.carbpol.2013.04.101.
- [204] J. Xie, J. Zhao, D.J. Hu, et al., Comparison of polysaccharides from two species of *Ganoderma*, Molecules 17 (2012) 740-752. https://doi. org/10.3390/molecules17010740.
- [205] D. Pan, L. Wang, C. Chen, et al., Structure characterization of a novel neutral polysaccharide isolated from *Ganoderma lucidum* fruiting bodies, Food Chem. 135 (2012) 1097-1103. https://doi.org/10.1016/ j.foodchem.2012.05.071.
- [206] X. Sun, H. Wang, X. Han, et al., Fingerprint analysis of polysaccharides from different *Ganoderma* by HPLC combined with chemometrics methods, Carbohydr. Polym. 114 (2014) 432-439. https://doi.org/10.1016/ j.carbpol.2014.08.048.
- [207] Y. Xu, X. Zhang, X.H. Yan, et al., Characterization, hypolipidemic and antioxidant activities of degraded polysaccharides from *Ganoderma lucidum*, Int. J. Biol. Macromol. 135 (2019) 706-716. https://doi.org/10.1016/ j.ijbiomac.2019.05.166.
- [208] H. Zhao, Q. Zhang, L. Zhao, et al., Spore powder of *Ganoderma lucidum* improves cancer-related fatigue in breast cancer patients undergoing endocrine therapy: a pilot clinical trial, Evid. Based Complement Alternat. Med. 2012 (2012) 809614. https://doi.org/10.1155/2012/809614.
- [209] I. Boldizsar, K. Horvath, G. Szedlay, et al., Simultaneous GC-MS quantitation of acids and sugars in the hydrolyzates of immunostimulant, water-soluble polysaccharides of basidiomycetes, Chromatographia 47 (1998) 413. https://doi.org/10.1007/BF02466472.
- [210] O.O. Orole, GC-MS evaluation, phytochemical and antinutritional screening of *Ganoderma lucidum*, J. Adv. Biol. Biotechnol. 5 (2016) 1-10. https://doi. org/10.9734/JABB/2016/24261.

- [211] J.Z. He, P. Shao, X.H. Men, et al., Analysis of structural characteristics of polysaccharide from *Ganoderma lucidum*, Chinese J. Anal. Chem. 38 (2010) 372-376. https://doi.org/10.3724/SP.J.1096.2010.00372.
- [212] H. Zhao, C.J. Lai, Y. Yu, et al., Acidic hydrolysate fingerprints based on HILIC-ELSD/MS combined with multivariate analysis for investigating the quality of *Ganoderma lucidum* polysaccharides, Int. J. Biol. Macromol. 163 (2020) 476-484. https://doi.org/10.1016/j.ijbiomac.2020.06.206.
- [213] Y. Huang, F. Xu, W. Zhang, et al., Progress for pharmacometabolomics and its applications, J. China Pharm. Univ. 44 (2013) 105-112. https://doi. org/10.11665/j.issn.1000-5048.20130202.
- [214] S.D. Milhorini, D.D. Bellan, M. Zavadinack, et al., Antimelanoma effect of a fucoxylomannan isolated from *Ganoderma lucidum* fruiting bodies, Carbohydr. Polym. 294 (2022) 119823. https://doi.org/10.1016/ j.carbpol.2022.119823.
- [215] S. Wachtel-Galor, J. Yuen, J.A. Buswell, et al., *Ganoderma lucidum* (Lingzhi or Reishi): a medicinal mushroom, 2nd edition ed, CRC Press/Taylor & Francis, Boca Raton, 2011.
- [216] H. Luo, D.C. Tan, B. Peng, et al., The pharmacological rationales and molecular mechanisms of *Ganoderma lucidum* polysaccharides for the therapeutic applications of multiple diseases, Am. J. Chin. Med. 50 (2022) 53-90. https://doi.org/10.1142/s0192415x22500033.
- [217] H. Pan, Y. Wang, K. Na, et al., Autophagic flux disruption contributes to Ganoderma lucidum polysaccharide-induced apoptosis in human colorectal cancer cells via MAPK/ERK activation, Cell Death Dis. 10 (2019) 456. https://doi.org/10.1038/s41419-019-1653-7.
- [218] X. Dan, W. Liu, J.H. Wong, et al., A ribonuclease isolated from wild Ganoderma lucidum suppressed autophagy and triggered apoptosis in colorectal cancer cells, Front. Pharmacol. 7 (2016) 217. https://doi. org/10.3389/fphar.2016.00217.
- [219] A. Thyagarajan, A. Jedinak, H. Nguyen, et al., Triterpenes from *Ganoderma lucidum* induce autophagy in colon cancer through the inhibition of p38 mitogen-activated kinase (p38 MAPK), Nutr. Cancer 62 (2010) 630-640. https://doi.org/10.1080/01635580903532390.
- [220] F.S. Reis, R.T. Lima, P. Morales, et al., Methanolic extract of *Ganoderma lucidum* induces autophagy of AGS human gastric tumor cells, Molecules 20 (2015) 17872-17882. https://doi.org/10.3390/molecules201017872.
- [221] W. Zhang, Z. Lei, J. Meng, et al., Water Extract of Sporoderm-Broken Spores of *Ganoderma lucidum* induces osteosarcoma apoptosis and restricts autophagic glux, Onco. Targets Ther. 12 (2019) 11651-11665. https://doi. org/10.2147/ott.s226850.
- [222] X. Liu, Y. Xu, Y. Li, et al., *Ganoderma lucidum* fruiting body extracts inhibit colorectal cancer by inducing apoptosis, autophagy, and G0/G1 phase cell cycle arrest *in vitro* and *in vivo*, Am. J. Transl. Res. 12 (2020) 2675-2684.
- [223] L.X. Sun, Z.B. Lin, X.J. Li, et al., Promoting effects of Ganoderma lucidum polysaccharides on B16F10 cells to activate lymphocytes, Basic Clin. Pharmacol. Toxicol. 108 (2011) 149-154. https://doi.org/10.1111/j.1742-7843.2010.00632.x.
- [224] C. Wang, S. Shi, Q. Chen, et al., Antitumor and immunomodulatory activities of *Ganoderma lucidum* polysaccharides in gliomabearing rats, Integr. Cancer Ther. 17 (2018) 674-683. https://doi. org/10.1177/1534735418762537.
- [225] L.X. Sun, W.D. Li, Z.B. Lin, et al., Protection against lung cancer patient plasma-induced lymphocyte suppression by *Ganoderma lucidum* polysaccharides, Cell Physiol. Biochem. 33 (2014) 289-299. https://doi. org/10.1159/000356669.
- [226] J. Shen, H.S. Park, Y.M. Xia, et al., The polysaccharides from fermented Ganoderma lucidum mycelia induced miRNAs regulation in suppressed HepG2 cells, Carbohydr. Polym. 103 (2014) 319-324. https://doi. org/10.1016/j.carbpol.2013.12.044.
- [227] G. Wang, L. Wang, J. Zhou, et al., The possible role of PD-1 protein in *Ganoderma lucidum*-mediated immunomodulation and cancer treatment, Integr. Cancer Ther. 18 (2019) 1-13. https://doi. org/10.1177/1534735419880275.
- [228] R. Rubel, H.S.D. Santa, L.F. Dos Santos, et al., Immunomodulatory and antitumoral properties of *Ganoderma lucidum* and *Agaricus brasiliensis* (Agaricomycetes) medicinal mushrooms, Int. J. Med. Mushrooms. 20 (2018) 393-403. https://doi.org/10.1615/IntJMedMushrooms.2018025979.

- [229] J. Su, L. Su, D. Li, et al., Antitumor activity of extract from the sporodermbreaking spore of *Ganoderma lucidum*: restoration on exhausted cytotoxic T cell with gut microbiota remodeling, Front. Immunol. 9 (2018) 1765. https:// doi.org/10.3389/fimmu.2018.01765.
- [230] A. Opattova, J. Horak, S. Vodenkova, et al., Ganoderma lucidum induces oxidative DNA damage and enhances the effect of 5-fluorouracil in colorectal cancer in vitro and in vivo, Mutat. Res. 845 (2019) 403065. https:// doi.org/10.1016/j.mrgentox.2019.06.001.
- [231] J. Su, D. Li, Q. Chen, et al., Anti-breast cancer enhancement of a polysaccharide from spore of *Ganoderma lucidum* with paclitaxel: suppression on tumor metabolism with gut microbiota reshaping, Front. Microbiol. 9 (2018) 3099. https://doi.org/10.3389/fmicb.2018.03099.
- [232] D.L. Liu, Y.J. Li, D.H. Yang, et al., *Ganoderma lucidum* derived ganoderenic acid B reverses ABCB1-mediated multidrug resistance in HepG2/ADM cells, Int. J. Oncol. 46 (2015) 2029-2038. https://doi.org/10.3892/ijo.2015.2925.
- [233] Q.P. Wu, Y.Z. Xie, Z. Deng, et al., Ergosterol peroxide isolated from *Ganoderma lucidum* abolishes microRNA miR-378-mediated tumor cells on chemoresistance, PLoS One 7 (2012) e44579. https://doi.org/10.1371/ journal.pone.0044579.
- [234] D. Li, L. Gao, M. Li, et al., Polysaccharide from spore of *Ganoderma lucidum* ameliorates paclitaxel-induced intestinal barrier injury: apoptosis inhibition by reversing microtubule polymerization, Biomed. Pharmacother. 130 (2020) 110539. https://doi.org/10.1016/j.biopha.2020.110539.
- [235] H. Yu, Y. Yang, T. Jiang, et al., Effective radiotherapy in tumor assisted by *Ganoderma lucidum* polysaccharide-conjugated bismuth sulfide nanoparticles through radiosensitization and dendritic cell activation, ACS Appl. Mater Interfaces. 11 (2019) 27536-27547. https://doi.org/10.1021/ acsami.9b07804.
- [236] T.P. Smina, S. De, T.P. Devasagayam, et al., Ganoderma lucidum total triterpenes prevent radiation-induced DNA damage and apoptosis in splenic lymphocytes in vitro, Mutat. Res. 726 (2011) 188-194. https://doi. org/10.1016/j.mrgentox.2011.09.005.
- [237] S. Dai, J. Liu, X. Sun, et al., *Ganoderma lucidum* inhibits proliferation of human ovarian cancer cells by suppressing VEGF expression and upregulating the expression of connexin 43, BMC Complement. Altern. Med. 14 (2014) 434. https://doi.org/10.1186/1472-6882-14-434.
- [238] M. Kong, Y. Yao, H. Zhang, Antitumor activity of enzymatically hydrolyzed Ganoderma lucidum polysaccharide on U14 cervical carcinoma-bearing mice, Int. J. Immunopathol. Pharmacol. 33 (2019) 1-8. https://doi. org/10.1177/2058738419869489.
- [239] V.T. Nguyen, N.T. Tung, T.D. Cuong, et al., Cytotoxic and anti-angiogenic effects of lanostane triterpenoids from *Ganoderma lucidum*, Phytochem. Lett. 12 (2015) 69-74. https://doi.org/https://doi.org/10.1016/j.phytol.2015.02.012.
- [240] S. Chen, T. Yong, Y. Zhang, et al., Anti-tumor and anti-angiogenic ergosterols from *Ganoderma lucidum*, Front. Chem. 5 (2017) 85. https://doi. org/10.3389/fchem.2017.00085.
- [241] P.R. Guo, Y.W. Sheng, B. Liu, et al., Influence of *Ganoderma lucidum* polysaccharide on the inhibitory effects of cisplatin on the tumor growth and angiogenesis in bladder cancer (T24) cells-bearing nude mice, Jie Fang Jun Yi Xue Za Zhi 39 (2014) 470-474. https://doi.org/10.11855/ j.issn.0577-7402.2014.06.09.
- [242] A. Acevedo-Díaz, G. Ortiz-Soto, I.J. Suárez-Arroyo, et al., Ganoderma lucidum extract reduces the motility of breast cancer cells mediated by the RAC-lamellipodin axis, Nutrients 11 (2019) 1116. https://doi.org/10.3390/ nu11051116.
- [243] J. Loganathan, J. Jiang, A. Smith, et al., The mushroom *Ganoderma lucidum* suppresses breast-to-lung cancer metastasis through the inhibition of proinvasive genes, Int. J. Oncol. 44 (2014) 2009-2015. https://doi.org/10.3892/ ijo.2014.2375.
- [244] K. Na, K. Li, T. Sang, et al., Anticarcinogenic effects of water extract of sporodermbroken spores of *Ganoderma lucidum* on colorectal cancer *in vitro* and *in vivo*, Int. J. Oncol. 50 (2017) 1541-1554. https://doi.org/10.3892/ijo.2017.3939.
- [245] L. Zheng, Y.S. Wong, M. Shao, et al., Apoptosis induced by 9,11-dehydroergosterol peroxide from *Ganoderma lucidum* mycelium in human malignant melanoma cells is Mcl-1 dependent, Mol. Med. Rep. 18 (2018) 938-944. https://doi.org/10.3892/mmr.2018.9035.
- [246] K. Li, K. Na, T. Sang, et al., The ethanol extracts of sporoderm-broken spores of *Ganoderma lucidum* inhibit colorectal cancer *in vitro* and *in vivo*, Oncol. Rep. 38 (2017) 2803-2813. https://doi.org/10.3892/or.2017.6010.

- [247] D. Sohretoglu, C. Zhang, J. Luo, et al., ReishiMax inhibits mTORC1/2 by activating AMPK and inhibiting IGFR/PI3K/Rheb in tumor cells, Signal Transduct. Target Ther. 4 (2019) 21. https://doi.org/10.1038/s41392-019-0056-7.
- [248] A.Y. Cheng, Y.C. Chien, H.C. Lee, et al., Water-extracted Ganoderma lucidum induces apoptosis and S-phase arrest via cyclin-CDK2 pathway in glioblastoma cells, Molecules 25 (2020) 3585. https://doi.org/10.3390/ molecules25163585.
- [249] C. Jiao, W. Chen, X. Tan, et al., Ganoderma lucidum spore oil induces apoptosis of breast cancer cells in vitro and in vivo by activating caspase-3 and caspase-9, J. Ethnopharmacol. 247 (2020) 112256. https://doi. org/10.1016/j.jep.2019.112256.
- [250] C.S. Shao, X.H. Zhou, X.X. Zheng, et al., Ganoderic acid D induces synergistic autophagic cell death except for apoptosis in ESCC cells, J. Ethnopharmacol. 262 (2020) 113213. https://doi.org/10.1016/ j.jep.2020.113213.
- [251] C. Bal, Antioxidant and antimicrobial capacities of Ganoderma lucidum, J. Bacteriol. Mycol. 7 (2019) 5-7. https://doi.org/10.15406/ jbmoa.2019.07.00232.
- [252] S. Quereshi, A.K. Pandey, S.S. Sandhu, Evaluation of antibacterial activity of different *Ganoderma lucidum* extracts, J. Sci. Res. 3 (2010) 9-13. https:// doi.org/10.22159/ajpcr.2019.v12i7.33714.
- [253] R. Ghobadi, R. Mohammadi, J. Chabavizade, et al., Effect of *Ganoderma lucidum* powder on oxidative stability, microbial and sensory properties of emulsion type sausage, Adv. Biomed. Res. 7 (2018) 24. https://doi.org/10.4103/2277-9175.225595.
- [254] J. Mishra, A. Joshi, R. Rajput, et al., Phenolic rich fractions from mycelium and fruiting body of *Ganoderma lucidum* inhibit bacterial pathogens mediated by generation of reactive oxygen species and protein leakage and modulate hypoxic stress in HEK 293 cell line, Adv. Pharmacol. Sci. 2018 (2018) 6285615. https://doi.org/10.1155/2018/6285615.
- [255] N. Hoque, A.A. Faysal, I. Ahmed, et al., *In vitro* antioxidant, antimicrobial and cytotoxic activities of the various extracts of *Ganoderma lucidum* available in Bangladesh, J. Pharmacogn. Phytochem. 4 (2015) 42-46. https:// doi.org/10.4016/9522.01.
- [256] G. Celk, In vitro Antimicrobial and antioxidant properties of Ganoderma lucidum extracts grown in turkey, Eur. J. Med. Plants. 4 (2014) 709-722.
- [257] M. Erawati, M. Andriany, N.S.D. Kusumaningrum, The Potential of Ganoderma lucidum as antimicrobial agent for multidrug-resistant mycobacterium tuberculosis, Antiinfect. Agents. 16 (2018) 11-14. https:// doi.org/10.2174/2211352516666180227135043.
- [258] L. Hleba, N. Vuković, J. Petrová, et al., Antimicrobial activity of crude methanolic extracts from *Ganoderma lucidum* and *Trametes versicolor*, Sci Pap: Anim Sci Biotechnol. 47 (2014) 89-93. https://doi.org/10.15407/ ukrbotj72.04.393.
- [259] J. Mishra, R. Rajput, K. Singh, et al., Antibacterial natural peptide fractions from Indian *Ganoderma lucidum*, Int. J. Pept. Res. Ther. 24 (2017) 543-554. https://doi.org/10.1007/s10989-017-9643-z.
- [260] P. Sa-Ard, R. Sarnthima, S. Khammuang, et al., Antioxidant, antibacterial and DNA protective activities of protein extracts from *Ganoderma lucidum*, J. Food Sci. Technol. 52 (2015) 2966-2973. https://doi.org/10.1007/s13197-014-1343-5.
- [261] S.A. Heleno, I.C. Ferreira, A.P. Esteves, et al., Antimicrobial and demelanizing activity of *Ganoderma lucidum* extract, *p*-hydroxybenzoic and cinnamic acids and their synthetic acetylated glucuronide methyl esters, Food Chem. Toxicol. 58 (2013) 95-100. https://doi.org/10.1016/j.fct.2013.04.025.
- [262] H. Kaur, S. Sharma, P.K. Khanna, et al., Evaluation of *Ganoderma lucidum* strains for the production of bioactive components and their potential use as antimicrobial agents, J. Appl. Nat. Sci. 7 (2015) 298-303. https://doi. org/10.31018/JANS.V711.605.
- [263] W.A.A.Q.I. Wan-Mohtar, L. Young, G.M. Abbott, et al., Antimicrobial properties and cytotoxicity of sulfated (1,3)-β-D-glucan from the mycelium of the mushroom *Ganoderma lucidum*, J. Microbiol. Biotechnol. 26 (2016) 999-1010. https://doi.org/10.4014/jmb.1510.10018.
- [264] S. Savin, O. Craciunescu, A. Oancea, et al., Antioxidant, cytotoxic and antimicrobial activity of chitosan preparations extracted from *Ganoderma lucidum* mushroom, Chem. Biodivers. 17 (2020) e2000175. https://doi. org/10.1002/cbdv.202000175.

- [265] S. Mahendran, S. Saravana, P. Vijayabaskar, et al., Antibacterial potential of microbial exopolysaccharide from *Ganoderma lucidum* and *Lysinibacillus fusiformis*, Int. J. Recent Sci. Res 4 (2013) 501-505. https://doi.org/10.1021/ acs.jafc.9b01195.s001.
- [266] D.S. Stojkovic, L. Barros, R.C. Calhelha, et al., A detailed comparative study between chemical and bioactive properties of *Ganoderma lucidum* from different origins, Int. J. Food Sci. Nutr. 65 (2014) 42-47. https://doi.org/10.3 109/09637486.2013.832173.
- [267] Swati, A. Tiwari, P. S. Negi, et al., A Comparative evaluation of *in vitro* anti-inflammatory and antifungal activity of *Ganoderma lucidum* strains DARL-4 and MS-1, Int. J. Green Pharm. 12 (2018) S126-S130. https://doi. org/10.5204/thesis.eprints.116592.
- [268] M.A. Arias-Londoño, P.A. Zapata-Ocampo, Á.R. Mosquera-Arévalo, et al., Antifungal protein determination for submerged cultures of the medicinal mushroom *Ganoderma lucidum* (Ganodermataceae) with activity over the phytopathogen fungus *Mycosphaerella fijiensis* (Mycosphaerellaceae), Actual. Biol. 41 (2020) 53-64. https://doi.org/10.17533/udea.acbi. v41n111a04.
- [269] Y.Q. Li, S.F. Wang, Anti-hepatitis B activities of ganoderic acid from Ganoderma lucidum, Biotechnol. Lett. 28 (2006) 837-841. https://doi. org/10.1007/s10529-006-9007-9.
- [270] Y. Li, Y. Yang, L. Fang, et al., Anti-hepatitis activities in the broth of *Ganoderma lucidum* supplemented with a Chinese herbal medicine, Am. J. Chin. Med. 34 (2006) 341-349. https://doi.org/10.1142/s0192415x06003874.
- [271] Z. Li, J. Liu, Y. Zhao, Possible mechanism underlying the antiherpetic activity of a proteoglycan isolated from the mycelia of *Ganoderma lucidum in vitro*, J. Biochem. Mol. Biol. 38 (2005) 34-40. https://doi.org/10.5483/ bmbrep.2005.38.1.034.
- [272] Y. Hijikata, A. Yasuhara, Y. Sahashi, Effect of an herbal formula containing *Ganoderma lucidum* on reduction of herpes zoster pain: a pilot clinical trial, Am. J. Chin. Med. 33 (2005) 517-523. https://doi.org/10.1142/ s0192415x05003120.
- [273] S. Bharadwaj, K.E. Lee, V.D. Dwivedi, et al., Discovery of *Ganoderma lucidum* triterpenoids as potential inhibitors against Dengue virus NS2B-NS3 protease, Sci. Rep. 9 (2019) 19059. https://doi.org/10.1038/s41598-019-55723-5.
- [274] W.Z. Lim, P.G. Cheng, A.Y. Abdulrahman, et al., The identification of active compounds in *Ganoderma lucidum* var. antler extract inhibiting dengue virus serine protease and its computational studies, J. Biomol. Struct. Dyn. 38 (2020) 4273-4288. https://doi.org/10.1080/07391102.2019.1678523.
- [275] P.J. Farrell, Epstein-Barr Virus and Cancer, In Annu Rev Pathol, A. K., Abbas; J. C., Aster; M. B., Feany, Eds. 2019.
- [276] S. Huh, S. Lee, S.J. Choi, et al., Quercetin synergistically inhibit ebvassociated gastric carcinoma with *Ganoderma lucidum* extracts, Molecules 24 (2019) 3834. https://doi.org/10.3390/molecules24213834.
- [277] D.S. Zheng, L.S. Chen, Triterpenoids from *Ganoderma lucidum* inhibit the activation of EBV antigens as telomerase inhibitors, Exp. Ther. Med. 14 (2017) 3273-3278. https://doi.org/10.3892/etm.2017.4883.
- [278] B. Donatini, Control of oral human papillomavirus (HPV) by medicinal mushrooms, *Trametes versicolor* and *Ganoderma lucidum*: a preliminary clinical trial, Int. J. Med. Mushrooms 16 (2014) 497-498. https://doi. org/10.1615/intjmedmushrooms.v16.i5.80.
- [279] E. Hernández-Márquez, A. Lagunas-Martínez, V.H. Bermudez-Morales, et al., Inhibitory activity of Lingzhi or Reishi medicinal mushroom, *Ganoderma lucidum* (higher Basidiomycetes) on transformed cells by human papillomavirus, Int. J. Med. Mushrooms 16 (2014) 179-187. https://doi. org/10.1615/intjmedmushr.v16.i2.80.
- [280] W. Zhang, J. Tao, X. Yang, et al., Antiviral effects of two Ganoderma lucidum triterpenoids against enterovirus 71 infection, Biochem. Biophys. Res. Commun. 449 (2014) 307-312. https://doi.org/10.1016/ j.bbrc.2014.05.019.
- [281] T.L. Hsu, S.C. Cheng, W.B. Yang, et al., Profiling carbohydrate-receptor interaction with recombinant innate immunity receptor-Fc fusion proteins, J. Biol. Chem. 284 (2009) 34479-34489. https://doi.org/10.1074/jbc. M109.065961.
- [282] A.V. Avtonomova, L.M. Krasnopolskaya, Antiviral properties of basidiomycetes metabolites, Antibiot. Khimioter. 59 (2014) 41-48. https:// doi.org/10.1007/978-981-16-4779-6_15.

- [283] R. Akbar, W.K. Yam, Interaction of ganoderic acid on HIV related target: molecular docking studies, Bioinformation 7 (2011) 413-417. https://doi. org/10.6026/97320630007413.
- [284] H.X. Wang, T.B. Ng, A laccase from the medicinal mushroom *Ganoderma lucidum*, Appl. Microbiol. Biotechnol. 72 (2006) 508-513. https://doi.org/10.1007/s00253-006-0314-9.
- [285] N. Suwannarach, J. Kumla, K. Sujarit, et al., Natural bioactive compounds from fungi as potential candidates for protease inhibitors and immunomodulators to apply for coronaviruses, Molecules 25 (2020) https:// doi.org/10.3390/molecules25081800.
- [286] N.A. ElSayed, G. Aleppo, V.R. Aroda, et al., Pharmacologic approaches to glycemic treatment: standards of medical care in diabetes-2020, Diabetes Care 43 (2020) S98-S110. https://doi.org/10.2337/dc20-S009.
- [287] L. Xu, Y. Li, Y. Dai, et al., Natural products for the treatment of type 2 diabetes mellitus: pharmacology and mechanisms, Pharmacol. Res. 130 (2018) 451-465. https://doi.org/10.1016/j.phrs.2018.01.015.
- [288] D.H. Ryu, J.Y. Cho, N.B. Sadiq, et al., Optimization of antioxidant, antidiabetic, and anti-inflammatory activities and ganoderic acid content of differentially dried *Ganoderma lucidum* using response surface methodology, Food Chem. 335 (2021) 127645. https://doi.org/10.1016/ j.foodchem.2020.127645.
- [289] M.Y. Chen, D. Xiao, W. Liu, et al., Intake of *Ganoderma lucidum* polysaccharides reverses the disturbed gut microbiota and metabolism in type 2 diabetic rats, Int. J. Biol. Macromol. 155 (2020) 890-902. https://doi. org/10.1016/j.ijbiomac.2019.11.047.
- [290] H.N. Li, L.L. Zhao, D.Y. Zhou, et al., *Ganoderma lucidum* polysaccharides ameliorates hepatic steatosis and oxidative stress in *db/db* mice via targeting nuclear factor E2 (erythroid-derived 2)-related factor-2/heme oxygenase-1 (HO-1) pathway, Med. Sci. Monit. 26 (2020) e921905. https://doi. org/10.12659/msm.921905.
- [291] S.D. Chen, T.Q. Yong, Y.F. Zhang, et al., Inhibitory effect of five *Ganoderma* species (Agaricomycetes) against key digestive enzymes related to type 2 diabetes mellitus, Int. J. Med. Mushrooms 21 (2019) 703-711. https://doi.org/10.1615/IntJMedMushrooms.v21.i7.70.
- [292] S.F. Shen, L.F. Zhu, Z.J. Wu, et al., Production of triterpenoid compounds from *Ganoderma lucidum* spore powder using ultrasound-assisted extraction, Prep. Biochem. Biotechnol. 50 (2020) 302-315. https://doi.org/10.1080/1082 6068.2019.1692218.
- [293] H. Xiao, Z. Fang, X. He, et al., Recombinant Ling Zhi-8 enhances Tregs function to restore glycemic control in streptozocin-induced diabetic rats, J. Pharm. Pharmacol. 72 (2020) 1946-1955. https://doi.org/10.1111/ jphp.13360.
- [294] H. Liang, Y. Pan, Y. Teng, et al., A proteoglycan extract from *Ganoderma lucidum* protects pancreatic beta-cells against STZ-induced apoptosis, Biosci. Biotechnol. Biochem. (2020) 1-8. https://doi.org/10.1080/09168451.2020.1 805718.
- [295] Z. Yang, C. Chen, J. Zhao, et al., Hypoglycemic mechanism of a novel proteoglycan, extracted from *Ganoderma lucidum*, in hepatocytes, Eur. J. Pharmacol. 820 (2018) 77-85. https://doi.org/10.1016/j.ejphar.2017.12.020.
- [296] L. Li, J.X. Xu, Y.J. Cao, et al., Preparation of Ganoderma lucidum polysaccharide-chromium (III) complex and its hypoglycemic and hypolipidemic activities in high-fat and high-fructose diet-induced prediabetic mice, Int. J. Biol. Macromol. 140 (2019) 782-793. https://doi. org/10.1016/j.ijbiomac.2019.08.072.
- [297] N.L. Klupp, H. Kiat, A. Bensoussan, et al., A double-blind, randomised, placebo-controlled trial of *Ganoderma lucidum* for the treatment of cardiovascular risk factors of metabolic syndrome, Sci. Rep. 6 (2016) 29540. https://doi.org/10.1038/srep29540.
- [298] T.Y. Vitak, S.P. Wasser, E. Nevo, et al., Enzymatic system of antioxidant protection of erythrocytes in diabetic rats treated with medicinal mushrooms agaricus brasiliensis and *Ganoderma lucidum* (Agaricomycetes), Int. J. Med. Mushrooms 19 (2017) 697-708. https://doi.org/10.1615/ IntJMedMushrooms.2017021305.
- [299] T.A. Wihastuti, R. Amiruddin, F.Y. Cesa, et al., Decreasing angiogenesis vasa vasorum through Lp-PLA2 and H2O2 inhibition by PSP from *Ganoderma lucidum* in atherosclerosis: *in vivo* diabetes mellitus type 2, J. Basic Clin. Physiol. Pharmacol. 30 (2020) 1-6. https://doi.org/10.1515/ jbcpp-2019-0349.

- [300] T. Heriansyah, W. Nurwidyaningtyas, D. Sargowo, et al., Polysaccharide peptide (PsP) *Ganoderma lucidum*: a potential inducer for vascular repair in type 2 diabetes mellitus model, Vasc. Health Risk Manage 15 (2019) 419-427. https://doi.org/10.2147/VHRM.S205996.
- [301] C. Xiao, Q. Wu, J. Zhang, et al., Antidiabetic activity of *Ganoderma lucidum* polysaccharides F31 down-regulated hepatic glucose regulatory enzymes in diabetic mice, J. Ethnopharmacol. 196 (2017) 47-57. https://doi.org/10.1016/ j.jep.2016.11.044.
- [302] Y. Gao, H. Gao, E. Chan, et al., Protective effect of *Ganoderma* (a mushroom with medicinal properties) against various liver injuries, Food Rev. Int. 21 (2005) 27-52. https://doi.org/10.1081/fri-200040586.
- [303] Y. Liu, C. Zhang, J. Du, et al., Protective effect of *Ganoderma lucidum* polysaccharide against carbon tetrachloride-induced hepatic damage in precision-cut carp liver slices, Fish Physiol. Biochem. 43 (2017) 1209-1221. https://doi.org/10.1007/s10695-016-0333-0.
- [304] Y. Shi, J. Sun, H. He, et al., Hepatoprotective effects of Ganoderma lucidum peptides against D-galactosamine-induced liver injury in mice, J. Ethnopharmacol. 117 (2008) 415-419. https://doi.org/10.1016/ j.jep.2008.02.023.
- [305] H.F. Han, N. Nakamura, M. Hattori, Protective effects of an acidic polysaccharide isolated from fruiting bodies of *Ganoderma lucidum* against murine hepatic injury induced by *Propionibacterium acnes* and lipopolysaccharide, J. Nat. Med. 60 (2006) 295-302. https://doi.org/10.1007/ s11418-006-0004-z.
- [306] N.P. Sudheesh, T.A. Ajith, J. Mathew, et al., *Ganoderma lucidum* protects liver mitochondrial oxidative stress and improves the activity of electron transport chain in carbon tetrachloride intoxicated rats, Hepatol. Res. 42 (2012) 181-191. https://doi.org/10.1111/j.1872-034X.2011.00906.x.
- [307] X.L. Li, A.G. Zhou, X.M. Li, Inhibition of *Lycium barbarum* polysaccharides and *Ganoderma lucidum* polysaccharides against oxidative injury induced by γ-irradiation in rat liver mitochondria, Carbohydr. Polym. 69 (2007) 172-178. https://doi.org/10.1016/j.carbpol.2006.09.021.
- [308] R.J.K. Susilo, D. Winarni, S.A. Husen, et al., Hepatoprotective effect of crude polysaccharides extracted from *Ganoderma lucidum* against carbon tetrachloride-induced liver injury in mice, Vet. World. 12 (2019) 1987-1991. https://doi.org/10.14202/vetworld.2019.1987-1991.
- [309] Z. Hu, R. Du, L. Xiu, et al., Protective effect of triterpenes of *Ganoderma lucidum* on lipopolysaccharide-induced inflammatory responses and acute liver injury, Cytokine 127 (2020) 154917. https://doi.org/10.1016/j.cyto.2019.154917.
- [310] C. Zhao, J. Fan, Y. Liu, et al., Hepatoprotective activity of *Ganoderma lucidum* triterpenoids in alcohol-induced liver injury in mice, an iTRAQ-based proteomic analysis, Food Chem. 271 (2019) 148-156. https://doi.org/10.1016/j.foodchem.2018.07.115.
- [311] Y.J. Liu, J.L. Du, L.P. Cao, et al., Anti-inflammatory and hepatoprotective effects of *Ganoderma lucidum* polysaccharides on carbon tetrachlorideinduced hepatocyte damage in common carp (*Cyprinus carpio* L.), Int Immunopharmacol. 25 (2015) 112-20. https://doi.org/10.1016/ j.intimp.2015.01.023.
- [312] M. Oliveira, F.S. Reis, D. Sousa, et al., A methanolic extract of *Ganoderma lucidum* fruiting body inhibits the growth of a gastric cancer cell line and affects cellular autophagy and cell cycle, Food Funct. 5 (2014) 1389-1394. https://doi.org/10.1039/c4fo00258j.
- [313] C.Y. Liang, H.R. Li, H. Zhou, et al., Recombinant Lz-8 from *Ganoderma lucidum* induces endoplasmic reticulum stress-mediated autophagic cell death in SGC-7901 human gastric cancer cells, Oncol. Rep. 27 (2012) 1079-1089. https://doi.org/10.3892/or.2011.1593.
- [314] A.A. Shaito, D.T.B. Thuan, H.T. Phu, et al., Herbal medicine for cardiovascular diseases: efficacy, mechanisms, and safety, Front. Pharmacol. 11 (2020) 422. https://doi.org/10.3389/fphar.2020.00422.
- [315] W.L. Shi, H. Han, G.Z. Chen, et al., Extraction, characterization of the polysaccharide extracts from Se-enriched *G. lucidum* (Se-GLP) and its inhibition against oxidative damage in ischemic reperfusion mice, Carbohydr. Polym. 80 (2010) 774-778. https://doi.org/10.1016/j.carbpol.2009.12.027.
- [316] X. Zhang, C. Xiao, H. Liu, Ganoderic acid a protects rat H9c2 cardiomyocytes from hypoxia-induced injury via up-regulating miR-182-5p, Cell Physiol. Biochem. 50 (2018) 2086-2096. https://doi. org/10.1159/000495053.

- [317] Y.Z. Xie, F. Yang, W. Tan, et al., The anti-cancer components of *Ganoderma lucidum* possesses cardiovascular protective effect by regulating circular RNA expression, Oncoence 28 (2016) 7-8. https://doi.org/10.18632/ oncoscience.316.
- [318] B.E. Klein, R. Klein, K.E. Lee, Components of the metabolic syndrome and risk of cardiovascular disease and diabetes in Beaver Dam, Diabetes Care 25 (2002) 1790. https://doi.org/10.2337/diacare.25.10.1790.
- [319] C.S. Fox, M.J. Pencina, P.W. Wilson, et al., Lifetime risk of cardiovascular disease among individuals with and without diabetes stratified by obesity status in the Framingham heart study, Diabetes Care. 31 (2008) 1582-1584. https://doi.org/10.2337/dc08-0025.
- [320] B.R. Baiju, R. Retnakaran, G.L. Booth, Increased risk of cardiovascular disease in young women following gestational diabetes mellitus, Diabetes Care 31 (2008) 1668-1669. https://doi.org/10.2337/dc08-0706.
- [321] T.V. Lasukova, A.G. Arbuzov, L.N. Maslov, et al., *Ganoderma lucidum* extract in cardiac diastolic dysfunction and irreversible cardiomyocytic damage in ischemia and reperfusion of the isolated heart, Patol. Fiziol. Eksp. Ter. (2008) 22-25. https://doi.org/10.1007/s10517-015-2851-7.
- [322] F. Wang, Z. Zhou, X. Ren, et al., Effect of *Ganoderma lucidum* spores intervention on glucose and lipid metabolism gene expression profiles in type 2 diabetic rats, Lipids Health Dis. 14 (2015) 1-9. https://doi.org/10.1186/ s12944-015-0045-y.
- [323] T.A. Wihastuti, T. Heriansyah, The inhibitory effects of polysaccharide peptides (PsP) of *Ganoderma lucidum* against atherosclerosis in rats with dyslipidemia, Heart Int. 12 (2017) e1-e7. https://doi.org/10.5301/ heartint.5000234.
- [324] L. Zengenni, Y. Zhihang, L. Gaoyang, et al., Hypolipidemic, antioxidant, and antiapoptotic effects of polysaccharides extracted from reishi mushroom, *Ganoderma lucidum* (Leysser: Fr) Karst, in mice fed a high-fat diet, J. Med. Food 21 (2018) 1218-1227. https://doi.org/10.1089/jmf.2018.4182.
- [325] Y. Gao, X. Dai, G. Chen, et al., A randomized, placebo-controlled, multicenter study of *Ganoderma lucidum* (W. Curt.: Fr.) Lloyd (Aphyllophoromycetideae) polysaccharides (Ganopoly®) in patients with advanced lung cancer, Int. J. Med. Mushrooms 5 (2003) 369-382. https://doi. org/10.1615/InterJMedicMush.v5.i4.40.
- [326] W. He, J. Yi, Study of clinical efficacy of Lingzhi spore capsule on tumour patients with chemotherapy/radiotherapy, Clin. J. Trad. Chin. Med. 9 (1997) 292-293. https://doi.org/CNKI:SUN:AHLC.0.1997-06-010.
- [327] M. Lu, K. Leng, Investigation of ZhengQing Lingzhi liquid as adjuvant treatment on patients with colon cancer, J. Guiyang Coll. Tradit. Chin. Med. 28 (2003) 1. https://doi.org/10.3969/j.issn.1000-2707.2003.05.021.
- [328] B. Yan, Y. Wei, Y. Li, Effect of Laojunxian Lingzhi oral liquid combined with chemotherapy on non-parvicellular lung cancer at stages II and III, Tradit. Chin. Drug Res. Clin. Pharmacol. 9 (1998) 78-80.
- [329] X. Zhang, Y. Jia, Q. Li, et al., Clinical curative effect investigation of Lingzhi tablet on lung cancer, Chin. Tradit. Patent. Med. 22 (2000) 486-488. https://doi.org/10.3969/j.issn.1001-1528.2000.07.012.
- [330] N. Lin, J. Su, Z. Zhu, et al., Analysis on 66 cases of cancer patients treated by chemotherapy with extract of *Ganoderma lucidum*, J. Pract. Tradit. Chin. Intern. Med. 18 (2004) 457-454. https://doi.org/10.3969/ j.issn.1671-7813.2004.05.071.
- [331] J. Zhou, Q. Zhang, Effect of *Ganoderma lucidum* spore on T cell subtype and VEGF of peripheral blood in old patients with carcinoma of uterine cervix, Matern Child Health Care China. 29 (2014) 2021-2022.
- [332] Z. Zhen, F. Wang, G. Fan, et al., Effect of *Ganoderma lucidum* spore to the immunological function of patients with hepatocellular carcinoma after operation, Chin. J. Hepat. Surg. (Electron Ed). 2 (2013) 171-174. https://doi. org/10.3877/cma.j.issn.2095-3232.2013.03.008.
- [333] Y. Benkui, W. Yanju, L. Yuqiang, Effect of Laojunxian Lingzhi oral liquid combined with chemotherapy on non_parvicellular lung cancer at stages II and III, Tradit. Chin. Drug Res. Pharmacol. 9 (1998) 13-16.
- [334] Z. Lin, B. Yang, *Ganoderma* and health: pharmacology and clinical application, Springer Nature, 2019.
- [335] H. Zhao, Q. Zhang, L. Zhao, et al., Spore powder of *Ganoderma lucidum* improves cancer-related fatigue in breast cancer patients undergoing endocrine therapy: a pilot clinical trial, Evid. Based Complement. Alternat. Med. 2012 (2012).

- [336] Y.H. Shieh, C.F. Liu, Y.K. Huang, et al., Evaluation of the hepatic and renalprotective effects of *Ganoderma lucidum* in mice, Am. J. Chin. Med. 29 (2001) 501-507. https://doi.org/10.1142/S0192415X01000526.
- [337] N. Futrakul, M. Boongen, P. Tosukhowong, et al., Treatment with vasodilators and crude extract of *Ganoderma lucidum* suppresses proteinuria in nephrosis with focal segmental glomerulosclerosis, Nephron 92 (2002) 719-720. https://doi.org/10.1159/000064082.
- [338] G. Xiao, F. Liu, Z. Chen, Clinical observation on treatment of Russula subnigricans poisoning patients by *Ganoderma lucidum* decoction, Zhongguo Zhong Xi Yi Jie He Za Zhi 23 (2003) 278-280.
- [339] N. Futrakul, T. Panichakul, P. Butthep, et al., *Ganoderma lucidum* suppresses endothelial cell cytotoxicity and proteinuria in persistent proteinuric focal segmental glomerulosclerosis (FSGS) nephrosis, Clin. Hemorheol. Microcirc. 31 (2004) 267-272.
- [340] Y. Hijikata, A. Yasuhara, Y. Sahashi, Effect of an herbal formula containing *Ganoderma lucidum* on reduction of herpes zoster pain: a pilot clinical trial, Am. J. Chinese Med. 33 (2005) 517-523.
- [341] G.H. Wang, X. Li, W.H. Cao, et al., A retrospective study of *Ganoderma lucidum* spore powder for patients with epilepsy, Medicine 97 (2018) e10941. https://doi.org/10.1097/MD.00000000010941.
- [342] L. Zou, H. Zhang, Research advance of morinda officinalis oligosaccharides in treatment of depression, Chin. J. New Drugs 21 (2012) 1889-1945.
- [343] L.H. Qin, C. Wang, L.W. Qin, et al., Spore powder of *Ganoderma lucidum* for Alzheimer's disease: a protocol for systematic review, Medicine 98 (2019) e14382. https://doi.org/10.1097/MD.000000000014382.
- [344] E.K. Li, L.S. Tam, C.K. Wong, et al., Safety and efficacy of *Ganoderma lucidum* (lingzhi) and San Miao San supplementation in patients with rheumatoid arthritis: a double-blind, randomized, placebo-controlled pilot trial, Arthritis. Care Res. 57 (2007) 1143-1150. https://doi.org/10.1002/art.22994.

- [345] J.P. Gau, C.K. Lin, S.S. Lee, et al., The lack of antiplatelet effect of crude extracts from *Ganoderma lucidum* on HIV-positive hemophiliacs, Am. J. Chin. Med. 18 (1990) 175-179. https://doi.org/10.1142/ S0192415X90000228.
- [346] H. Xu, J. Xue, Applications "Ji 731 Solution" treatment of atrophic rhinitis preliminary summary, J. Peking Univ. 3 (1979) 180.
- [347] A.M. Faruque, Ganoderma lucidum: persuasive biologically active constituents and their health endorsement, Biomed. Pharmacother. 107 (2018) 507-519. https://doi.org/10.1016/j.biopha.2018.08.036.
- [348] T.R. Smina, J. Matheu, K.K. Janardhanan, et al., Antioxidant activity and toxicity profile of total triterpenes isolated from *Ganoderma lucidum* (Fr.) P. Karst occurring in South India, Environ. Toxicol. Pharmacol. 32 (2011) 438-446. https://doi.org/10.1016/j.etap.2011.08.011.
- [349] J. Zhang, X. Gao, Y. Pan, et al., Toxicology and immunology of Ganoderma lucidum polysaccharides in Kunming mice and Wistar rats, Int. J. Biol. Macromol. 85 (2016) 302-310. https://doi.org/10.1016/ j.ijbiomac.2015.12.090.
- [350] Y. Kwok, K.F.J. Ng, C.C.F. Li, et al., A prospective, randomized, doubleblind, placebo-controlled study of the platelet and global hemostatic effects of *Ganoderma lucidum* (Ling-Zhi) in healthy volunteers, Anesth. Analg. 101 (2005) 423-426. https://doi.org/10.1213/01.ANE.0000155286.20467.28.
- [351] J. Tao, K.Y. Feng, Experimental and clinical studies on inhibitory effect of *Ganoderma lucidum* on platelet aggregation, J. Tongji Med. Univ. 10 (1990) 240-243. https://doi.org/10.1007/BF02887938.
- [352] S.M. Wicks, R. Tong, C.Z. Wang, et al., Safety and tolerability of *Ganoderma lucidum* in healthy subjects: a double-blind randomized placebocontrolled trial, Am. J. Chinese Med. 35 (2007) 407-414. https://doi. org/10.1142/S0192415X07004928.